

Meiotic Behavior In Treated *Vicia Faba* Plant With Diprofos Drug

Salha. M. S. AL-Shamrani

Faculty of Science (Girls), King Abdulaziz University, Jeddah, Kingdom of Saudi Arabia.

Corresponding author: sal-2006@hotmail.com

Abstract: The cytotoxicity of the corticosteroid drug "diprofos" (0.50, 1, 2 and 4ml /100ml) were examined on *Vicia faba* meiotic behavior. *Vicia faba* plants at the flowering stage spraying with the obvious diprofos concentrations to study the meiotic behavior after 24h. and 48h. from spraying. All diprofos treatments showed highly significant increased of abnormal pollen mother cells (PMC_s) which increased as the concentration and period duration increased. Abnormalities % at the second meiotic division were lower than those recorded in the first meiotic division in most treatments as a result of recovery in this cell age. On the other hand, the abnormalities were present in metaphase and anaphase stages at the first and second meiotic division. Stickiness and disturbed chromosomes were the most common abnormalities found in all phases of meiosis after diprofos treatments. In addition, laggard, bridges, breaks and micronuclei, which recorded with low percentages in some treatments. Results strongly suggest that diprofos drug has a cytotoxic effect on *Vicia faba* plant.

[Salha. M. S. AL-Shamrani. **Meiotic Behavior In Treated *Vicia Faba* Plant With Diprofos Drug.** *Life Sci J* 2015;12(5):41-45]. (ISSN:1097-8135). <http://www.lifesciencesite.com>. 5

Keywords: Mitotic activity, chromosomal abnormalities, Lagging chromosomes, bridges, fragments and Stickiness.

1.Introduction

Corticosteroids are contra-indicated in patients with peptic ulcer, osteoporosis, psychoses, or sever psychoneuroses. Because of the interference with inflammatory and immunological response, corticosteroids should not be usually given in presence of cute infections. Life vaccines should be avoided in patients receiving corticosteroids. Corticosteroids should not be given during pregnancy or lactation. prolonged use of beta-methasone in high doses may cause pituitary suppression, acute adrenal insufficiency, fluid and electrolyte disturbance, hyperglycemia and glucosuria, increased susceptibility to infections, peptic ulcers, osteoporosis, arrest of growth, cushin's habitus (moon- face,buffalo hump), behavioral disturbance and cataract,Henein, (2004). Chromosomal aberration test (CA) is one of the sensitive and important tests for evaluating genetic hazard of drugs and / or carcinogens because CA is known to have a serious impact on human health. There is a clear association between chromosomal aberration and certain types of cancer. Many drugs such as; Trifluo perazine, aspirin, indometacin, theophylline, novalgin, oxicam, ampicilin, piroxicam, carbenilin, phenoparetal, clomiditens, nitroimidazol, acetyl salicylic acid, bohemine roscovitine, fenaton, diazebam, megazol, penta brominaled and nitroso-methylurea were studied in different biological systems (e.g., mice, salmonella, yeast, *Drosophila melanogaster*, human lymphocytes, *Allium cepa* and *Helianthus annus*.)El- Bayoumi et al., 1984; Atefet al., 2011). The present study aimed to evaluate the mutagenic effect of diprofos drug with different

concentration at exposure times on *Vicia faba* meiotic division behavior.

2.Materials And Methods:

2.1.Materials:

2.1.1.Sample:

Vicia faba plants (Var. Giza 40) at the flowering stage were used in this study

2.1.2.Tested drug:

The teated drug was Diprofos drug It is produced by Schering Plough Company, USA. This drug is present in ampoule form,each one ml ampoule contains 7 mg betamethasone.

2.2.Methods:

2.2.1.Treatment:

- *Vicia faba* plants were sprayed with four medical preparations of Diprofos drug.The recommended dose for this drug depends on the disease type. Generally, Dosage for adults: short term (1-2) ml daily for the first few days, subsequently reducing the daily dosage by 0.50-1 ml every 2-5 days.

- Four diprofos concentrations were used:
0.50, 1, 2, 4 ml/100 ml water.

- Control plants were sprayed with distilled water.

Ten flower buds from ten different plants were gathered after (24 and 48) hours from spraying for study.

2.2.2. Meiotic studies:

The appropriate flower buds were collected and fixed in carnoy's solution (ethyl alcohol absolute and glacial acetic acid with ratio 3:1 for 24h., and then transferred to 70% ethyl alcohol and kept in refrigerator. The cytological analysis were carried. by

using 2% aceto-carmin stain) as described by Darlington and La Cour, 1979).

2.2.3. Statistical analysis:

The data recorded for different treatments were statistically analyzed using t-test for determining significantly of differences between these treatments.

3. Results And Discussion

3.1- Total Meiotic Abnormalities Percentages:

A wide spectrum of meiotic abnormalities were recorded in ten flower buds from different plants after different treatments with diprofos. Data in (table1)

shows that total abnormal PMCs % was highly significant increased as the diprofos concentration increased and by the increasing of period duration from 24h. to 28h. in all treatments. Whereas, the total abnormalities ranged from (26.34% - 51.87 %) at the 24 h. period, while it ranged from (35.00% - 60.60 %) at the 48 h. period. The induction of meiotic abnormalities appears to be a common effect of most chemicals by many investigators (Ann et al, 2012; Rina et al., 2012; Madhuri et al, 2012; Sefa et al, 2012).

Table(1): Percentages of abnormal PMC, in the 1st & 2nd meiotic divisions and total mean of meiotic abnormalities after spraying of *Vicia faba* plant with diprofos for 24 and 48 hours.

Conc. ml/100ml	Times hours	% abnormal In PMCs meiotic division			% abnormal in 1st division			% abnormal in 2nd division		
		dividing Cells No.	abnormal Cells No.	Abn.% PMCs ± S.E	Dividing Cells No	abnormal Cells No	abn.%	dividing Cells No.	abnormal Cells No.	Abn.%
Control		510	18	3.44 ± 1.07	276	6	2.17	234	12	5.13
0.50	24	540	148	26.34 ± 2.02**	186	42	22.58	354	106	29.94
	48	531	186	35.00 ± 1.04**	209	74	35.41	322	112	34.78
1	24	500	153	30.44 ± 0.85**	265	120	45.28	235	33	14.04
	48	559	202	36.09 ± 0.51**	310	138	44.52	249	64	25.70
2	24	515	246	47.81 ± 0.90**	209	120	57.42	306	126	41.18
	48	491	244	49.59 ± 0.55**	251	144	57.37	240	100	41.67
4	24	462	240	51.87 ± 2.62**	246	130	52.85	216	110	50.93
	48	475	288	60.60 ± 0.74**	285	176	61.75	190	112	58.95

PMCs: pollen mother cells. * significant (0.05), ** highly significant (0.01) (T- Test).

3.2- First and Second Meiotic Abnormalities Percentage

Table (1) revealed that the percentages of abnormalities in the first meiotic division increased as diprofos concentration increased and by the increasing of the duration period in the most treatments, which it ranged from (22.58% – 57.42%) and (35.41% – 61.75%) at the duration period 24h. and 48h. respectively.

Also, this trait in the second meiotic division increased as the increasing of both diprofos concentration and period duration except for diprofos concentration 1ml / 100ml which it showed the lower percentage in second meiotic abnormalities; 14.04% and 25.70% at the duration period 24h. and 48h. Respectively. While the highest value of this trait were 50.93% and 58.95% at the 4ml/100ml diprofos concentration at the duration period 24h. and 48h. respectively.

On other hand, the percentages of the abnormalities in the second meiotic division were

lower than those recorded in the first meiotic division in all treatments except for the (0.50ml/100ml) diprofos at the 24h. as a result of recovery in this cell age (table 1).

Results from table (2) and figure (1) revealed that abnormal% (met, ana and telo) phases in the second meiotic division ranged from: (5.53% – 21.05%, 5.53% – 21.05% and 0.56% – 11.11%) respectively lower than those recorded in the first meiotic division that ranged from: (5.38% – 27.37%, 13.98% – 27.37% and 3.23% – 12.20%) respectively as a result of recovery in this cell age except for two treatments: 0.50 ml/100ml at 24 h. and 1ml/100ml at 24h. for abnormal (meta and telo) phases respectively. However, abnormalities didn't show in prophase 1 but it observed in prophase 2 with percentage (3.11% – 13.68%). These results were in agreement with many researches, (Priti et al., 2009; Haroun, 2010; Asmahan and Salha, 2012).

Table (2): Abnormal meiotic phases percentages in *Vicia faba* plants after spraying with diprofos for 24 and 48hours

Conc. MI/100 ML	Time Hours	First Meiotic Division			Second Meiotic Division			
		Abn. Metaphase %	Abn. Anaphase%	Abn. Telophase%	Abn. Prophase%	Abn. Metaphase%	Abn. Anaphase%	Abn. Telophase %
Control		1.45	0.72	----	----	5.13	----	----
0.50	24	5.38	13.98	3.23	3.39	14.69	11.30	0.56
	48	17.22	14.35	3.83	3.11	11.18	18.01	2.48
1	24	20.38	18.87	6.04	4.26	5.53	4.26	----
	48	22.58	18.06	3.87	3.21	8.84	12.85	0.80
2	24	23.92	25.84	7.66	6.54	18.30	10.46	5.88
	48	27.09	24.70	5.58	6.67	15.83	11.67	7.50
4	24	17.89	22.76	12.20	8.33	16.67	14.81	11.11
	48	27.37	27.37	7.02	13.68	21.05	16.84	7.37

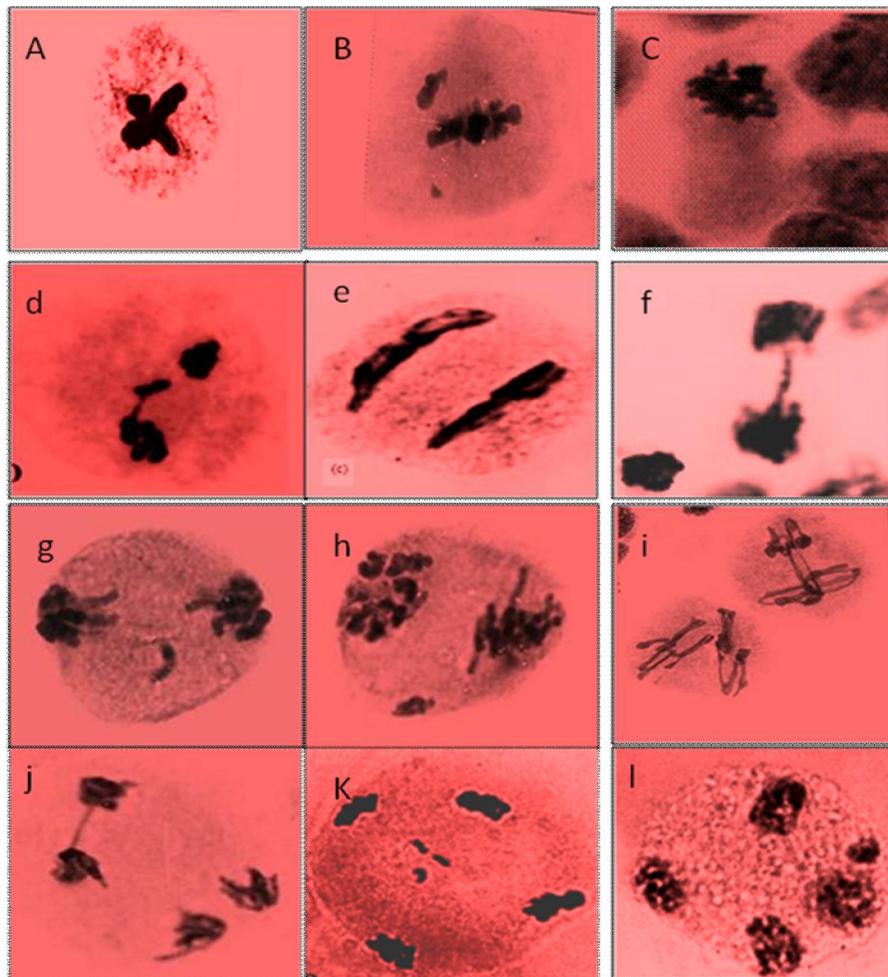


Fig.1: Different types of abnormalities in treated *Vicia faba* plant after diprofos treatments. a: Sticky(M_1); b: Sticky, Lagging and break (M_1); c: Sticky and disturbed (M_1); d: Sticky and Lagging (A_1); e: Sticky(M_2); f: Sticky and bridges(A_1); g: Lagging (T_1); h: micronuclei (P_2); i: disturbed (M_2); j: bridges (A_2); k: Lagging (T_2); l: micronuclei(T_2).

{ M_1, A_1, T_1 : first (meta, ana)phase}; P_2, M_2, A_2, T_2 : second (pro, meta, ana, telo)phase
 Fig.1: Different types of abnormalities in treated *Vicia faba* plant after diprofos treatments. a: Sticky(M_1); b: Sticky, Lagging and break (M_1); c: Sticky and disturbed (M_1); d: Sticky and Lagging (A_1); e: Sticky(M_2); f: Sticky and bridges(A_1); g: Lagging (T_1); h: micronuclei (P_2); i: disturbed (M_2); j: bridges (A_2); k: Lagging (T_2); l: micronuclei(T_2).

{ M_1, A_1, T_1 : first (meta, ana)phase}; P_2, M_2, A_2, T_2 : second (pro, meta, ana, telo)phase}

Table (3): Percentages of different meiotic abnormalities in the 1st and 2nd meiotic divisions after spraying *Vicia faba* plants with diprofos for 24 and 48 hours.

Conc. MI/100ML	Time Hours	1st Meiotic Division					2nd Meiotic Division				
		Stick-iness	Disturb-ed	Lagg-ards	Bridges	Breaks	Stick-iness	Disturbed	Lagg-ards	Bridges	Micro-nuclei
Control		----	1.18	----	----	----	0.39	1.96	----	----	----
0.50	24	2.59	5.19	----	----	----	10	9.26	----	----	0.37
	48	4.90	9.04	----	----	----	12.05	7.53	----	----	1.51
1	24	12	10.80	0.40	0.80	----	2.60	4	----	----	----
	48	11.45	11.81	0.36	1.07	----	8.59	2.86	----	----	----
2	24	15.53	6.99	0.39	0.39	----	18.25	3.88	1.94	----	0.39
	48	22.81	3.67	1.63	1.22	----	15.89	2.04	0.81	0.81	0.81
4	24	26.84	----	0.43	0.87	----	22.08	----	0.43	0.43	0.87
	48	30.74	1.26	2.11	1.68	1.26	18.11	1.26	1.68	0.84	1.68

3.3-Types of different meiotic abnormalities:

Stickiness and disturbed chromosomes were the most common abnormalities found in all phases of meiosis after diprofos treatments (table 3- fig:1). Number of sticky cells increase as the diprofos concentration increased or per longed the period duration in the most treatments.

On the other hand, sticky cells in the second meiotic division decreased than those recorded in the first meiotic division at these diprofos treatments: 1 and 4ml / 100ml (at 24h. and 48h.) and 2ml / 100ml at 24h. (Table 3). Sticky chromosomes were shown in various meiotic division (Fig.1:a,b,c,d,e and f). Our results are in agreement with the results of many investigators(Ann et al, 2012; Rina et al., 2012; Madhuri et al, 2012; Sefa et al, 2012; Asmahan, and Salha 2012), who suggested that chromosome stickiness may results from breakage and exchange between chromatin fibers on the surface of adjoining chromosomes.

The second type of abnormalities is the disturbed which observed in metaphase and anaphase in all treatments (Fig.1: c, f and i), and the percentage of this trait ranged from: (1.26% – 11.81%) and (1.26% – 9.26%) in the first and second meiotic division respectively (Table 3).

This abnormality was observed by many researcher (Maju et al.,1984; El-Ashmawy et al.,1992; Obrecht et al.,1996; Giri et al.,1999; Alia,et al.,2001; Poli et al., 2002; Evandri et al.,2003; Arkhipchuk et al., 2004; Sadiq and Al-Quraishe 2004; Usciat et al., 2004; Singh et al.,2005; Ganguly et al.,2010; Haroun, 2010; Gabriele et al.,2010; Min et al.,2010; Saulo et al., 2010; Atef et al., 2011) after many chemicals treatments, they suggested that the chromosomes disturbed may results from the effect of the chemical treatments on proteins constituting the spindle apparatus.

In addition to previous common abnormalities, it was observed another abnormalities on meiotic division including: laggards, bridges, breaks and micronuclei, which recorded with low percentages in some

treatments (Table 3). Laggard chromosomes were observed in some treatments in (meta, ana, and telo) phases in the first and second meiotic division (Fig.1, b, d, k and g). Laggard at metaphase could be attributed to failure of the spindle apparatus to organize and function in a normal way (Haroun, 2010). These Laggards may be distributed randomly to either poles at both anaphase and telophase (I and II) which result ultimately in aneuploidy (Ann, *et al*, 2012) or may give for micronuclei at telophase (Rina, *et al.*, 2012).The induction of Laggard chromosomes could be attributed to irregular orientation of chromosomes (Min, *et al.*,2010).

On the other hand, breaks appeared in metaphase I only in 4ml 100ml at 48h.(Table 3, Fig.1: b). While, bridges were induced in some treatments, (Table 3, Fig.1:f and j) and they could be due to the breakage and reunion (Atef, *et al.*,2011) or due to the general stickiness of chromosomes (Sefa, *et al.*, 2012). While, micronuclei was also recorded in some treatments (Table 3, Fig.1: h and l) and our results are in agreement with the results of (Haroun, 2010).

Finally, the induction of these chromosomal abnormalities were pointed to the mutagenic potential of the diprofos drug.

References

1. Henein, W.H. (2004): Atlas1: Everything about drugs form A to Z. First Edition. El-Nasr Modern Library, Cairo, Egypt, 24-26.
2. El- Bayoumi, A.S.; Habib, A.A. and Haliem, A.S. (1984): Mitotic disturbances induced by the drug " Novalgin " in *Allium cepa* root tips. *Qatar Uni. Sci. Bull.* 4: 95-115.
3. Maju, M.; Jaju, M. and Ahuja, Y.R. (1984): Evaluation of genotoxicity of ampicilin and carbenicillin on human lymphocytes in vitro chromosome aberration, mitotic index, cell cycle Kintice, satellite associations of acrocentric chromosomes and sister chromatid exchanges. *Hum. Toxicol.* 3 (3): 173-191.
4. El-Ashmawy, S. H; Abdel-Rahem and Abu Salha, A.E. (1992): cytological effects of two colchicines

- drugs on *Allium cepa* root tips Egypt. *J. Genet. Cytol.*, 21: 133-141.
5. Obreeht, P. S.; Grosse, Y.; Pfohl-Leskowicz, A. and Dirheimer, G. (1996): Protection by indomethacin and aspirin against genotoxicity of ochratoxin A, particularly in the urinary bladder and kidney. *Arch. Toxicol.*, 70 (3-4): 244-8.
 6. Giri, A.K.; Das, M.; Reddy, V.G. And Pal, A.K. (1999): Mutagenic and genotoxic effects of theopylline and the obromine in Salmonella assay and in vivo sister chromatid exchanges in bone marrow cells of mice. *Mut. Res.* 444 (1): 17-23.
 7. Alia, A. E; Ezzat, S. E; Fatma, M. I. B. And Maha, M. A. E. (2001): Cytogenetic effects of two antiepileptic drugs on mice bone marrow cells and human a-mitotic cell cultures. *Egypt. J. Genet. Cytol.*, 29: 115-129.
 8. Poli, P.; Aline, M. M.; Buschini, A.; Mortara, R.A.; Northfleet, A. C.; Silva, S.; Rossi, C. and Zucchi, T.M. (2002): Cytotoxic and genotoxic effects of megazol, an anti-Chagas' disease drug, assessed by different short term tests. *Biochem. Pharmacol.* 1, 01 (11): 1617-1627.
 9. Evandri, M.G., Mastrangelo, L., G and Costand, P.B. (2003): *In vitro* assessment of mutagenicity and clastogenicity of penta brominated diphenyl ether flame retardant. *Environ. Mol. Mutagen*; 42 (2): 85-90.
 10. Arkhipchuk, V. V., Goncharuk, V. V., Charnkh V.P., Malshtan, I. N. and Gristenke, I. S. (2004): Use of a complex approach for assessment of metamizole sodium and acetyl-salicylic acid toxicity. *Genotoxicity And Cytotoxicity, Appl. Toxicol.*; 24 (5): 401-7.
 11. Sadiq, M.F. And Al-Quraishe, F.A. (2004): the effect of trifluoperazine on the induction of sex-linked recessive lethal by cyclophosphamide in *Drosophila melanogaster*. *Drug Chem. Toxicol.*, 27 (1): 169-78.
 12. Singh, G.; Dricver, P.H. And Sander, J.W. (2005): Cancer risk in people with epilepsy: the role of antiepileptic drugs. *Brain*, 28 (1): 7-17.
 13. Usciat, M.; Codaccioni, M. and Guern, J. (2004): Early cytological and biochemical events induced by a 6 benzylaminopurine application on inhibited auxiliary buds of *Cicer arietinum* plants. *Journal of Experimental Botany*, 23(4): 1009-1020
 14. Priti, K., Lalit, M.T.; Tapan, K.N.; Tarun, K.B.; Lelit, S. and Bibhesh, K. S. (2009): Chromosomal abnormalities arising under the action of antibiotics in *Pisum sativum*. *Nature and Science*, 7 (3): 104-112.
 15. Ganguly, S.; Bhattacharya, S.; Mandi, S. and Tarafdar, J. (2010): Biological detection and analysis of toxicity of organophosphate and azadirachtin-based insecticides in *Lathyrus sativus* L. *Ecotoxicology*, 19: 85-95.
 16. Haroun, S.A. (2010): Mutagenic effects of kockia indica extraction on *Vicia faba* L. *Journal of American Science*, 6 (7): 292-299.
 17. Gabriele, J.; Svetla, G.; Mila, S. and Stanislava, K. (2010): Cytotoxic and genotoxic effects of paraquat in *Hordeum vulgare* and human Lymphocytes *in vitro*. *Environ. Toxicol.*, 25: 294-303.
 18. Min, Yi; Huilan, Yi; Honghai, Li and Lihua, Wu. (2010): Aluminum induces chromosome aberrations, micronuclei, and cell cycle dysfunction in root cells of *Vicia faba*. *Environ. Toxicol.*, 25: 124-129.
 19. Saulo, M., S.; Pamela, S. S. and Lyderson, F. V. (2010): Cytogenotoxicity of *Cymbopogon. citrates* (DC) stapf (lemom grass) aqueous extracts in vegetal test systems. *Anais da Academia Brasileira de Ciencias*, 82 (2): 305-311.
 20. Atef, A.A.H.; Abd EL-Hamid N.R.; Abd EL-Hady, E.A. and AL- Ansary, A.M. (2011): Cytogenetic effect of insecticide Tellition and Fungicide Dithane M 45 on meiotic cells and seed storage proteins of *Vicia faba*. *Journal of American Science*, 7 (1): 19-25.
 21. Drlington, C.D. and Lacour, E.(1976): The handling of chromosomes. Sixth edition George Allan and Unwin Lid., London,50-52.
 22. Ann, T. D; Julie, H and Mike, O. D. (2012): Chromosome aberration frequency in rat peripheral lymphocytes increases with repeated dosing with hexa methyl phosphoramidate or cyclophosphamide. *Mutagenesis*, 27(5): 533-539.
 23. Rina, T.;Shyam, S. P. and Pankaj, T. (2012): Genotoxicity of ibuprofen in mouse bone marrow cells *in vivo*. *Drug and Chemical Toxicology* (27)940-951.
 24. Madhuri, K. V.; Rabbani, S. J.; Kumar, N. and Khalid, A. (2012): Assessment of drug induced genotoxicity in gastric cancer patients. *African J. of Biotechnology*, vol. 11 (4):974-978.
 25. Sefa Pl.; Atila, Cr. and Bahattin, Ai. (2012): The toxic and environmental evaluation of pyrolytic liquids by *Allium cepa* test. *Chemistry and Ecology*, vol.28 (1): 65-73.
 26. Asmahan, A. M. A. and Salha, M. A. (2012): Cytogenetic and molecular evaluation of genetic effect of the clomid drug on *vicia faba* plant. *J. Rad. Res. Appl. Sci.*, Vol., No.2, pp.50-62.