

Antibacterial, antifungal activity and phytochemical analysis of some desert plants against human pathogenic bacteria and fungi

Mohamed E Zain¹, Amani S. Awaad², Monerah R. Al Othman¹, and Sahar K. Al-Dosary³

¹Botany and Microbiology Department, College of Science, King Saud University, Riyadh, KSA.

²Pharmacognosy Department, College of Pharmacy, Salman Bin Abdulaziz University, Al-Kharj, KSA.

³Biology Department, College of Science, Dammam University, Dammam, KSA.

mzain@ksu.edu.sa

Abstract: Five desert plants; namely, *Bidens bipinnata*, *Cyperus alternifolius*, *Desmostachya bipinnata*, *Glossostemon bruguieri*, and *Schouwia thebica* were investigated for their antimicrobial activity. The ethanolic extract inhibited the growth of some pathogenic bacteria including *Enterococcus faecalis*, *Escherichia coli*, *Moraxella lacunata*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Serratia marcescens*, *Bacillus subtilis*, *Micrococcus luteus*, *Sarcina ventriculi*, *Staphylococcus aureus* and pathogenic fungi including *Candida albicans*, *Candida tropicalis*, *Aspergillus flavus*, *A. fumigatus* and *Penicillium chrysogenum*. The extracts of *Cyperus alternifolius* and *Desmostachya bipinnata* were similar in their phytochemical constituents, while the chemical groups of *Bidens bipinnata* were similar to those in *Schouwia thebica*. Traces of saponins were only present in *Glossostemon bruguieri*.

[Mohamed E. Zain, Amani S. Awaad, Monerah R. Al Othman, and Sahar K. Al-Dosary. **Antibacterial, antifungal activity and phytochemical analysis of some desert plants against human pathogenic bacteria and fungi.** *Life Sci J* 2014;11(7):343-349]. (ISSN:1097-8135). <http://www.lifesciencesite.com>. 39

Keywords: Bioactive compounds, antibacterial and antifungal agents, human pathogenic microorganisms, desert plants.

1. Introduction

Plants are a source of great economic value all over the world. Nature has given us a very rich botanical wealth and large number of diverse types of plants grow in different parts of the country. Interestingly, the use of medicinal plants as a source for relief from illness can be traced back over five millennia to written documents of the early civilization in China, India and the Near East, but it is doubtless an art as old as mankind.

Plants contain large varieties of chemical substances, which possess important therapeutic properties that can be utilized in the treatment of human diseases. The studies of plants used in folkloric remedies have attracted the attention of many scientists in finding solution to the problems of multiple resistances to the existing synthetic antibiotics. Most of the synthetic antibiotics now available in the market have major setback due to the multiple resistance developed by pathogenic microorganisms against their drugs (**Akinpelu et al., 2008**).

Natural products perform various functions, and many of them have interesting and useful biological activities (**Harvey, 1999**). There are more than 35,000 plant species being used in various human cultures around the world for medicinal purpose. Researchers are increasingly turning their attention to natural products looking for new leads to develop better drugs against cancer, as well as viral

and microbial infections (**Srinivasan et al., 2001; Harvey, 1999; Hoffmann et al., 1993**).

Now a day's maximum number of plants are being screened for their possible pharmacological value. However, the plant kingdom still holds many plant species containing substance of medicinal value which have yet to be discovered (**Tambe et al., 2013**). The medicinal value of plants lies in some chemical substances that produce a definite physiologic action on the human body. The most important of these phytochemicals of plants are alkaloids, flavonoids, tannins and phenolic compounds (**Vedhanarayanan et al., 2013**). The current study was mainly concerned with investigating the antibacterial and antifungal activities of some plants against representatives of human pathogenic bacteria and fungi.

2. Materials and Methods

Plant Materials

Plant samples of *Bidens bipinnata*, *Cyperus alternifolius*, *Desmostachya bipinnata*, *Glossostemon bruguieri*, and *Schouwia thebica* were collected during in March 2013 from desert around Riyadh. The plants were identified by Dr. Jacob T. Pandalayil, Assistant Professor of Plant Taxonomy, Botany and Microbiology Department, College of Science, King Saud University and by comparison with plant description (**Migahid, 1996**). A voucher specimen has been deposited in the herbarium of College of Science, King Saud University.

Plant Extracts

The plant samples were washed 2-3 times with running fresh water and then air-dried under shade. After complete shade drying, the plant materials (300 g) was grinded with mechanical grinder and the powder was kept in tightly closed containers. The extract was prepared by percolation in 90% ethanol (Awaad et al., 2012) at room temperature for two days. The ethanol extract was filtered and the residues were re-percolated for four times. The total ethanol extract was concentrated under reduced pressure at a temperature not exceeding 35°C.

Antimicrobial Activity Assay

Test organisms

Eleven strains of pathogenic bacteria; *Enterococcus faecalis*, *Escherichiacoli*, *Moraxella lacunata*, *Proteus merabiles*, *Serratia marcesens*, *Pseudomons aeruginosa*, *Salmonella typhi*, *Bacillus subtilis*, *Micrococcus luteus*, *Sarcina ventricull*, *Staphylococcus aureus* and five strains of pathogenic fungi; *Candida albicans*, *Candida tropicalis*, *Aspergillus flavus*, *Aspergillus fumigates*, *Penicillium chrysogenum* were obtained from the Microbiology Lab, Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, Egypt and use as test organisms.

Antibacterial activity

The antibacterial activity of the investigate plants was determined by well diffusion method according to National Committee for Clinical Laboratory Standards (NCCLS, 1993). Petri plates containing 20 ml of Nutrient Agar medium were seeded with 24 hrs. old cultures of bacterial inoculum (standardized inoculums $1-2 \times 10^7$ cfu/ml 0.5 Mcfarland standard). Wells (6 mm in diameter) were cut off from agar and 100 µl of plant extracts were tested in a concentration of 5 mg/ml and incubated at 37°C for 24 hrs. The antibacterial activity was determined by measuring the inhibition zone formed around the well.

Antifungal activity

The antifungal activity was determined by well diffusion method according to National Committee for Clinical Laboratory Standards (NCCLS, 1993). Petri plates containing 20 ml of Malt extract Agar medium were seeded with 3 day cultures of fungal inoculums (standardized inoculums $1-2 \times 10^7$ cfu/ml 0.5 Mcfarland standard). Wells (5 mm in diameter) were cut off from agar and 50 µl of plant extracts were tested in a concentration of 5 mg/ml and incubated at 37°C for 4 days. The antifungal activity was determined by measuring the inhibition zone formed around the well.

Phytochemical Analysis

The extracts of *Bidens bipinnata*, *Cyperus alternifolius*, *Desmostachya bipinnata*, *Glossostemon bruguieri*, and *Schouwia thebicawere* chemically tested for the presence of different phytochemical constituents such as carbohydrates and/or glycosides (Majaw et al., 2009), flavonoides, tannins, sterols and/or triterpenes (Jack and Okorosaye-Orubite, 2008), proteins and/or amino acids, alkaloids and/or nitrogenous bases (Kumar et al., 2009), saponins (Edeoga et al., 2005), anthraquinones (Majaw et al., 2009), cardinolides (Sonibare et al., 2009) and oxidase enzyme (Wagner and Baldt, 2001).

3. Results

The antibacterial and antifungal activities of five desert plants were determined. Moreover, the chemical constituents of each plant were investigated. The ethanolic extract of each plant was investigated for its ability to inhibit the growth of human bacteria; *Enterococcus faecalis*, *Escherichiacoli*, *Moraxella lacunata*, *Proteus merabiles*, *Serratia marcesens*, *Pseudomons aeruginosa*, *Salmonella typhi*, *Bacillus subtilis*, *Micrococcus luteus*, *Sarcina ventricull*, *Staphylococcus aureus* and pathogenic fungi; *Candida albicans*, *Candida tropicalis*, *Aspergillus flavus*, *Aspergillus fumigates*, *Penicillium chrysogenum*.

Bidens bipinnata

The obtained results revealed that the ethanolic extract of *Bidens bipinnata* has antibacterial activity against the investigated human pathogenic bacteria (Table 1). The highest antibacterial activity (13 mm) was obtained against *Proteus merabiles*, *Bacillus subtilis*, and *Staphylococcus aureus*. However, good antibacterial activity (12, 11, and 10 mm) was obtained against *Serratia marcesens*, *Escherichia coli*, and *Enterococcus faecalis*, respectively. The lowest antiabacterial activity of *Bidens bipinnata* (6 mm) was obtained against *Salmonella typhi* and *Micrococcus luteus* (Table 1).

The antifungal activity of *Bidens bipinnata* (10 mm) was obtained against *Candida albicans* and *C. tropicalis*, (9 mm) against *Aspergillus flavus*, (8 mm) against *Aspergillus fumigates*, and (6 mm) against *Penicillium chrysogenum* (Table 1).

On the other hand, the obtained results of phytochemical analysis of *Bidens bipinnata* revealed the presence of tannins, flavonoids (free aglycone and glycoside), unsaturated and/or triterpenoides, carbohydrates or glycosides, and proteins and/or amino acids. The crystalline sublimate, volatile oil, alkaloides and/or nitrogenous bases, cardinolides, saponins, anthraquinones, and oxidase enzymes were absent (Table 1).

Cyperus alternifolius

The ethanolic extract of *Cyperus*

alternifolius has antibacterial activity against the investigated human pathogenic bacteria (Table 2). The highest antibacterial activity (13 and 11 mm) was obtained against *Pseudomonas aeruginosa* and *Serratia marcescens*, respectively. However, good antibacterial activity (10 mm) was obtained against *Moraxella lacunata*, *Bacillus subtilis*, and *Micrococcus luteus*; (9 mm) against *Proteus merabiles* and (8 mm) against *Enterococcus faecalis*, *Escherichia coli*, and *Sarcina ventricull*. The extract of *Cyperus alternifolius* showed no activity against *Salmonella typhi* and *Staphylococcus aureus* (Table

2).

On the other hand, the results of antifungal activity of *Cyperus alternifolius* revealed that the highest activity (15 and 13 mm) was obtained against *Candida albicans* and *C. tropicalis*, respectively (Table 2). Moreover, moderate antifungal activity (9 and 7 mm) was obtained against *Aspergillus flavus* and *Aspergillus fumigates*, respectively. The lowest antifungal activity (6 mm) was obtained against *Penicillium chrysogenum* (Table 2).

Table 1. The antibacterial, antifungal activities and chemical constituents of *Bidens bipinnata*.

Antibacterial activity		Antifungal activity		Phytochemical analysis	
Bacteria	Zone of inhibition (mm)	fungi	Zone of inhibition (mm)	Test	result
Gram-negative		Unicellular		Crystalline sublimate	-
<i>Enterococcus faecalis</i>	10	<i>Candida albicans</i>	10	Volatile oil	-
<i>Escherichia coli</i>	11	<i>Candida tropicalis</i>	10	Tannins	+
<i>Moraxella lacunata</i>	10	Filamentous		Flavonoids	
<i>Proteus merabiles</i>	13	<i>Aspergillus flavus</i>	09	a. free aglycone	+
<i>Serratia marcescens</i>	12	<i>Aspergillus fumigates</i>	08	b. glycoside	+
<i>Pseudomonas aeruginosa</i>	09	<i>Penicillium chrysogenum</i>	06	Alkaloids and/or nitrogenous bases	-
<i>Salmonella typhi</i>	06			Cardinolides	-
Gram-positive				Unsaturated sterols and/or triterpenoides	+
<i>Bacillus subtilis</i>	13			Saponins	-
<i>Micrococcus luteus</i>	06			Carbohydrates or glycosides	+
<i>Sarcina ventricull</i>	09			Anthraquinones	-
<i>Staphylococcus aureus</i>	13			Proteins and/or amino acids.	+
				oxidase enzyme	-

(-), absent; (+), present

Table 2. The antibacterial, antifungal activities and chemical constituents of *Cyperus alternifolius*.

Antibacterial activity		Antifungal activity		Phytochemical analysis	
Bacteria	Zone of inhibition (mm)	fungi	Zone of inhibition (mm)	Test	result
Gram-negative		Unicellular		Crystalline sublimate	+
<i>Enterococcus faecalis</i>	08	<i>Candida albicans</i>	15	Volatile oil	-
<i>Escherichia coli</i>	08	<i>Candida tropicalis</i>	13	Tannins	+
<i>Moraxella lacunata</i>	10	Filamentous		Flavonoids	
<i>Proteus merabiles</i>	09	<i>Aspergillus flavus</i>	09	a. free aglycone	+
<i>Serratia marcescens</i>	11	<i>Aspergillus fumigates</i>	07	b. glycoside	+
<i>Pseudomonas aeruginosa</i>	13	<i>Penicillium chrysogenum</i>	06	Alkaloids and/or nitrogenous bases	-
<i>Salmonella typhi</i>	00			Cardinolides	-
Gram-positive				Unsaturated sterols and/or triterpenoides	+
<i>Bacillus subtilis</i>	10			Saponins	-
<i>Micrococcus luteus</i>	10			Carbohydrates or glycosides	+
<i>Sarcina ventricull</i>	08			Anthraquinones	-
<i>Staphylococcus aureus</i>	00			Proteins and/or amino acids.	+
				oxidase enzyme	-

(-), absent; (+), present

The phytochemical constituents of *Cyperus alternifolius* contained crystalline sublimate, tannins, flavonoids (free aglycone and glycoside), unsaturated and/or triterpenoides, carbohydrates or glycosides, and proteins and/or amino acids. However, volatile oil, alkaloids and/or nitrogenous bases, cardiolides, saponins, anthraquinones, and oxidase enzymes were absent (Table 2).

Desmostachya bipinnata

The obtained results revealed that the ethanolic extract of *Desmostachya bipinnata* has antibacterial activity against the investigated human

pathogenic bacteria (Table 3). The highest antibacterial activity (13 and 12 mm) was obtained against *Micrococcus luteus* and *Bacillus subtilis*, respectively. However, good antibacterial activity (10 mm) was obtained against *Proteus merabiles*, *Pseudomons aeruginosa*, and *Salmonella typhi*; and (9 mm) against *Sarcina ventricull* and *Staphylococcus aureus*. The lowest antibacterial activity of *Desmostachya bipinnata* was obtained against *Salmonella typhi* and *Micrococcus luteus* (Table 3).

Table 3. The antibacterial, antifungal activities and chemical constituents of *Desmostachya bipinnata*.

Antibacterial activity		Antifungal activity		Phytochemical analysis	
Bacteria	Zone of inhibition (mm)	fungi	Zone of inhibition (mm)	Test	result
Gram-negative		Unicellular		Crystalline sublimate	+
<i>Enterococcus faecalis</i>	07	<i>Candida albicans</i>	10	Volatile oil	-
<i>Escherichia coli</i>	07	<i>Candida tropicalis</i>	11	Tannins	+
<i>Moraxella lacunata</i>	08	Filamentous		Flavonoids	
<i>Proteus merabiles</i>	10	<i>Aspergillus flavus</i>	07	a. free aglycone	+
<i>Serratia marcesens</i>	07	<i>Aspergillus fumigates</i>	08	b. glycoside	+
<i>Pseudomons aeruginosa</i>	10	<i>Penicillium chrysogenum</i>	07	Alkaloids and/or nitrogenous bases	-
<i>Salmonella typhi</i>	10			Cardiolides	-
Gram-positive				Unsaturated sterols and/or triterpenoides	+
<i>Bacillus subtilis</i>	12			Saponins	-
<i>Micrococcus luteus</i>	13			Carbohydrates or glycosides	+
<i>Sarcina ventricull</i>	09			Anthraquinones	-
<i>Staphylococcus aureus</i>	09			Proteins and/or amino acids.	+
				oxidase enzyme	-

(-), absent; (+), present

The highest antifungal activity of *Desmostachya bipinnata* was (11 and 10 mm) was obtained against *Candida tropicalis* and *C. albicans*, respectively. While the antifungal activity (8 mm) was obtained against *Aspergillus fumigates*. The lowest antifungal activity (7 mm) was obtained against *Aspergillus flavus* and *Penicillium chrysogenum* (Table 3).

The obtained results of phytochemical analysis of *Desmostachya bipinnata* revealed the presence of crystalline sublimate, tannins, flavonoids (free aglycone and glycoside), unsaturated and/or triterpenoides, carbohydrates or glycosides, and proteins and/or amino acids. The volatile oil, alkaloids and/or nitrogenous bases, cardiolides, saponins, anthraquinones, and oxidase enzymes were absent (Table 3).

Glossostemon bruguieri

The ethanolic extract of *Glossostemon bruguieri* has antibacterial activity against the investigated human pathogenic bacteria (Table 4).

The highest antibacterial activity (15, 12 and 11 mm) was obtained against *Staphylococcus aureus*, *Bacillus subtilis* and *Proteus merabiles*, respectively. However, good antibacterial activity (10 mm) was obtained against *Moraxella lacunata* and *Serratia marcesens*; (9 mm) against *Escherichia coli* and (8 mm) against *Enterococcus faecalis*, *Micrococcus luteus*, and *Sarcina ventricull*; (7 mm) against *Pseudomons aeruginosa*. The extract of *Glossostemon bruguieri* showed no activity against *Salmonella typhi* (Table 4).

On the other hand, the results of antifungal activity of *Glossostemon bruguieri* revealed that the highest activity (11 mm) was obtained against *Aspergillus flavus* and *Aspergillus fumigates*; (10 mm) against *Penicillium chrysogenum* (Table 4). There was no activity against *Candida albicans* and *C. tropicalis*.

The phytochemical analysis of *Glossostemon bruguieri* showed the presence of tannins, flavonoids (free aglycone and glycoside),

unsaturated and/or triterpenoides, traces of saponins, carbohydrates or glycosides, and proteins and/or amino acids. The crystalline sublimate, volatile oil, alkaloids and/or nitrogenous bases, cardanolides, saponins, anthraquinones, and oxidase enzymes were absent (Table 4).

Schouwia thebica

The obtained results revealed that the ethanolic extract of *Schouwia thebica* has antibacterial activity against the investigated human pathogenic bacteria (Table 5). The highest

antibacterial activity (11 mm) was obtained against *Enterococcus faecalis* and *Bacillus subtilis*. A good antibacterial activity (10 mm) was obtained against *Moraxella lacunata*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*; (9 mm) against *Escherichia coli*, *Proteus merabiles* and *Sarcina ventricull*. The lowest antibacterial activity of *Schouwia thebica* (7 mm) was obtained against *Salmonella typhi* and *Micrococcus luteus*. There was no activity against *Serratia marcesens* (Table 5).

Table 4. The antibacterial, antifungal activities and chemical constituents of *Glossostemon bruguieri*.

Antibacterial activity		Antifungal activity		Phytochemical analysis	
Bacteria	Zone of inhibition (mm)	fungi	Zone of inhibition (mm)	Test	result
Gram-negative		Unicellular		Crystalline sublimate	
<i>Enterococcus faecalis</i>	08	<i>Candida albicans</i>	00	Volatile oil	-
<i>Escherichia coli</i>	09	<i>Candida tropicalis</i>	00	Tannins	+
<i>Moraxella lacunata</i>	10	Filamentous		Flavonoids	
<i>Proteus merabiles</i>	11	<i>Aspergillus flavus</i>	11	a. free aglycone	+
<i>Serratia marcesens</i>	10	<i>Aspergillus fumigates</i>	11	b. glycoside	+
<i>Pseudomonas aeruginosa</i>	07	<i>Penicillium chrysogenum</i>	10	Alkaloids and/or nitrogenous bases	-
<i>Salmonella typhi</i>	00			Cardanolides	-
Gram-positive				Unsaturated sterols and/or triterpenoides	+
<i>Bacillus subtilis</i>	12			Saponins	±
<i>Micrococcus luteus</i>	08			Carbohydrates or glycosides	+
<i>Sarcina ventricull</i>	08			Anthraquinones	-
<i>Staphylococcus aureus</i>	15			Proteins and/or amino acids.	+
				oxidase enzyme	-

(-), absent; (+), present

Table 5. The antibacterial, antifungal activities and chemical constituents of *Schouwia thebica*.

Antibacterial activity		Antifungal activity		Phytochemical analysis	
Bacteria	Zone of inhibition (mm)	fungi	Zone of inhibition (mm)	Test	result
Gram-negative		Unicellular		Crystalline sublimate	
<i>Enterococcus faecalis</i>	11	<i>Candida albicans</i>	09	Volatile oil	-
<i>Escherichia coli</i>	09	<i>Candida tropicalis</i>	09	Tannins	+
<i>Moraxella lacunata</i>	10	Filamentous		Flavonoids	
<i>Proteus merabiles</i>	09	<i>Aspergillus flavus</i>	07	a. free aglycone	+
<i>Serratia marcesens</i>	00	<i>Aspergillus fumigates</i>	11	b. glycoside	+
<i>Pseudomonas aeruginosa</i>	10	<i>Penicillium chrysogenum</i>	08	Alkaloids and/or nitrogenous bases	-
<i>Salmonella typhi</i>	07			Cardanolides	-
Gram-positive				Unsaturated sterols and/or triterpenoides	+
<i>Bacillus subtilis</i>	11			Saponins	-
<i>Micrococcus luteus</i>	07			Carbohydrates or glycosides	+
<i>Sarcina ventricull</i>	09			Anthraquinones	-
<i>Staphylococcus aureus</i>	10			Proteins and/or amino acids.	+
				oxidase enzyme	-

(-), absent; (+), present

The highest antifungal activity of *Schouwia thebica* (11 mm) was obtained against *Aspergillus fumigates*. While the antifungal activity (9 mm) was obtained against *Candida tropicalis* and *C. albicans*; (8 mm) against *Penicillium chrysogenum*. The lowest antifungal activity (7 mm) was obtained against *Aspergillus flavus* (Table 5).

The obtained results of phytochemical analysis of *Schouwia thebica* revealed the presence of tannins, flavonoids (free aglycone and glycoside), unsaturated and/or triterpenoides, carbohydrates or glycosides, and proteins and/or amino acids. The crystalline sublimate, volatile oil, alkaloids and/or nitrogenous bases, cardiolides, saponins, anthraquinones, and oxidase enzymes were absent (Table 5).

4. Discussion

Natural sources including plants are important source of potentially useful structures, which could be used for the development of new chemotherapeutic agents. However, to achieve this goal, *in vitro* antibacterial and/or antifungal activity assay are required (Tona et al., 1998). Many reports are available on the antiviral, antibacterial, antifungal, anthelmintic, antimolluscal and anti-inflammatory properties of plants (Zain et al., 2014, 2012, 2009; Awaad et al., 2012; Chanda and Baravalia, 2010; Ahameethunisa and Hooper, 2010; Chattopadhyay et al., 2009; Alam et al., 2009; Govindarajan et al., 2006; Behera and Misra, 2005; Bylka et al., 2004).

In the present study five extracts of desert plants; *Bidens bipinnata*, *Cyperus alternifolius*, *Desmostachya bipinnata*, *Glossostemon bruguieri*, and *Schouwia thebica* inhibited the growth of human pathogenic bacterial and fungi. The antimicrobial activity of plants and their use in traditional medicine have been investigated and approved long time ago (Vedhanarayanan et al., 2013; Zain et al., 2012; Kambizi and Afolayan, 2008; Uniyal et al., 2006; Newman et al., 2000; Thomson, 1978).

The results of the current study revealed the presence of the investigated plants revealed the presence of different chemical compounds including tannins, flavonoids (free aglycone and glycoside), unsaturated sterols, carbohydrates or glycosides, and proteins and/or amino acids, crystalline sublimate, and alkaloids and/or nitrogenous bases. Isolation and identification of bioactive compounds from natural resources including plants were carried out by many studies (Zain et al., 2014; Awaad et al., 2013; El-Meligy et al., 2013; Malu et al., 2009; Ahmad et al., 2008; Raghavendra et al., 2006; Bylka et al., 2004; Girish et al., 2006; Mothana and Lindequist, 2005; Karaman et al., 2003).

Acknowledgment

This research project was supported by a grant from the "Research Center of the Female Scientific and Medical Colleges", Deanship of Scientific Research, King Saud University.

References

- Ahameethunisa, A. R. and Hooper, W. (2010). Antibacterial activity of *Artemisia nilagirica* leaf extracts against clinical and phytopathogenic bacteria. BMC Complemen. and Alternat. Med., 10: 1- 6.
- Ahmad, I.; Zahin, M.; Aqil, F.; Hasan, S.; Khan, M.S.A.; Owais, M. (2008). Bioactive compounds from *Punica granatum*, *Curcuma longa* and *Zingiber officinale* and their therapeutic potential. Drugs Fut. 33, 329.
- Akinpelu, D.A.; Aiyegoro, O.A.; Okoh, A.I. (2008). *In vitro* antimicrobial and phytochemical properties of crude extract of stem bark of *Azela africana* (Smith). Afr. J. Biotech., 7(20):3665 -3670.
- Alam, M.T.; Karim, M.M.; Shakila, K.N. (2009). Antibacterial activity of different organic extracts of *Achyranthes aspera* and *Cassia alata*. J. Sci. Res., 1: 393-398.
- Awaad, Amani S.; Soliman, G. A.; El-Sayed, Dalia F.; El-Gindi, Omimah D., and Alqasoumi, S. I. (2012). Hepatoprotective activity of *Cyperus alternifolius* on carbon tetrachloride-induced hepatotoxicity in rats. Pharmaceutical Biology, 2012; 50(2): 155-161.
- Awaad, Amani S.; Al-Jaber, Nabilah A.; Moses, John E.; El-Meligy, Reham M. and Zain, Mohamed E (2013). Antiulcerogenic Activities of the Extracts and Isolated Flavonoids of *Euphorbia cuneata* Vahl. Phytother. Res., 27: 126-130.
- Behera, S.K. and M.K. Misra, 2005. Indigenous phytotherapy for genito-urinary diseases used by the Kandha tribe of Orissa, India. J. Ethnopharmacol., 102: 319-325.
- Bylka, W., M. Szauffer-Hajdrych, I. Matalawskan and O. Goslinka, 2004. Antimicrobial activity of isocytoside and extracts of *Aquilegia vulgaris* L. Lett. Appl. Microbiol., 39: 93-97.
- Chanda, S. and Baravalia, Y. (2010). Screening of some plant extracts against some skin diseases caused by oxidative stress and microorganisms, Afr. J Biotech., 9: 3210-3217.
- Chattopadhyay, R. R.; Bhattacharya, S.K.; Medda, C.; Chanda, S. and Anvesa, B. (2009). A comparative evaluation of antibacterial potential of some plants used in Indian traditional medicine for the treatment of microbial infection, Braz. Archives of Bio. and Technol., 52: 1123-1128.
- Edeoga, H.O.; Okwu, D.E.; Mbaebie, B.O. (2005). Phytochemical constituents of some Nigerian medicinal plants. Afri. J.Biotechnol. 4:685-688.
- El-Meligy, Reham M.; Awaad, Amani S.; Zain, Mohamed E.; Alafeefy, Ahmed M. (2013). Evaluation of Selected Desert Plants as Anti-ulcerogenic Natural Products. Australian Journal of Basic and Applied Sciences, 7(4): 431-436.

13. Girish, K.S., K.D. Machiah, S. Ushanandini, H. Kumar, K.S. Nagaraju, Govindappa, M. Vedavathi and K. Kemparaju, 2006 Antimicrobial properties of a non-toxic glycoprotein (WSG) from *Withania somnifera* (Ashwagandha). *J. Basic Microbiol.*, 46: 365-374.
14. Govindarajan R, M Vijayakumar, M Singh, CHV Rao, A Shirwaikar, AKS Rawat and P Pushpangadan (2006). Antiulcer and antimicrobial activity of *Anogeissus latifolia*. *J. Ethnopharmacol.*, 106: 57-61.
15. Harvey AL (1999). Medicines from nature: are natural products still relevant to drug discovery? *Trends Pharmacol. Sci.*, 20: 196-198.
16. Hoffmann JJ, N Timmerman, R McLaughlin and H Punnapayak (1993). Potential antimicrobial activity of plants from the South Western United States. *Int. J. Pharmacolog.*, 31:101-115.
17. Jack, I.R. and Okorosaye-Orubite, K. (2008). Phytochemical analysis and antimicrobial activity of the extract of leaves of fleabane (*Conyza sumatrensis*). *J. Appl. Sci. Eeeviron. Manage.* 12 (4): 63-5.
18. Kambizi, L. and Afolayan, A.J. (2008). Extracts from *Aloe ferox* and *Withania somnifera* inhibit *Candida albicans* and *Neisseria gonorrhoea*. *African J. Biotechnol.*, 7: 12-15.
19. Karaman, I.; Sahin, F.; Gulluce, M.; Ogutcu, H.; Sengul, M.; Adiguzel, A. (2003). Antimicrobial activity of aqueous and methanol extracts of *Juniperus oxycedrus* L. *J. Ethnopharmacol.*, 85: 231-235.
20. Kumar, A., Llavaran, R., Jayachandran, T., Decaraman, M., Aravindhan, P., Padmanabhan, N., and Krishnan, M.R.V. (2009). Phytochemicals investigation on atropical plant, *syzygium cumini* from kattuppalayam Erode District, Tamil Nadu, South India. *Pakistan Journal of Nutrition*, 8, 1, p.83-5.
21. Majaw, S., and Moirangthem, J. (2009). Qualitative and Quantitative Analysis of *Clerodendron Colebrookianum* Walp. Leaves and *Zingiber Cassumunar* Roxb, Rhizomes. *Ethnobotani Cal Leaflets*, 13, p.578-89.
22. Malu, S.P., Obochi, G.O., Tawo, E.N., Nyong, B.E. (2009). Antibacterial activity and medical properties of ginger. (*Zingiber officinale*). *Global J. Pure Appl. Sci.* 15, 365-368.
23. Migahid A.M. (1996). *Flora of Saudi Arabia*. 4th edition. *King Saud University Press*. Volume 1, 127.
24. Mothana, R.A.A. and Lindequist, U. (2005). Antimicrobial activity of some medicinal plants of the island soqotra. *J. Ethnopharmacol.* 96: 177-181.
25. NCCLS, (1993). Performance Standards Antimicrobial Disc Susceptibility Tests. Approved Standard Fifth Edition. NCCLS Document M2-A5, Villanova, PA, USA.
26. Newman, D.J.; Cragg, G.M. and Snader, K.M. (2000). The influence of natural products upon drug discovery. *Nat. Prod. Res.*, 17: 215-234.
27. Raghavendra, M.P., Satish, S. and Raveesha, K.A. (2006). Phytochemical analysis and antibacterial activity of *Oxalis corniculata*: A known medicinal plant. *My. Science.*, 1: 72-78.
28. Sonibare, M.A.; Soladoye, M.O.; Esan O.O. and Sonibare, O.O. (2009). Phytochemical and Antimicrobial studies of four species of *cola schott* and *endi* (*sterculiaceae*). *Afr J Tradit Complement Altern Med*, 6, 4, p.518-25.
29. Srinivasan, D.; Sangeetha, N.; Suresh, T.; Perumalsamy, P.L. (2001). Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. *J. Ethnopharmacol.*, 74: 217-220.
30. Tambe, R.; Kulkarni, M.; Bhise, K. (2013). Preliminary phytochemical creening and HPTLC fingerprinting of bark extracts of *Symplocos racemosa*, 2(3):45-49.
31. Thomson, W.A.R. (1978). *Medicines from the Earth*. Maidenhead, United Kingdom. McGraw-Hill Book Co.
32. Tona, L., K. Kambu, N. Ngimbi, K. Cimanga and A.J. Vlietinck, 1998. Antiamoebic and phytochemical screening of some Congolese medicinal plants. *J. Ethnopharmacol.*, 61: 57-65.
33. Uniyal, S.K., K.N. Singh, P. Jamwal and B. Lal, 2006. Traditional use of medicinal plants among the tribal communities of Chhota Bhangal, Western Himalayan. *J. Ethnobiol. Ethnomed.*, 2: 1-14.
34. Vedhanarayanan, P.; Unnikannan, P.; Sundaramoorthy, P. (2013). Antimicrobial activity and phytochemical screening of *Wrightia tinctoria* (Roxb.) R.Br. *Journal of Pharmacognosy and Phytochemistry*, 2 (4): 123-125.
35. Wangner, H. and Baldt, S. (2001). *Plant drug analysis*. 2nd ed., Springer-Verlag, Berlin, Heidelberg, New York.
36. Zain M.E.; Awaad, Amani S.; Razak, A.A.; Maitland, D.J.; Khamis N.E. and Sakhawy, M.A. (2009). Secondary Metabolites of *Aureobasidium pullulans* Isolated from Egyptian Soil and Their Biological Activity. *Journal of Applied Sciences Research*, 5(10): 1582-1591.
37. Zain M. E.; Awaad, Amani S.; Al-Outhman Monerah R.; El-Meligy, Reham M. (2012). Antimicrobial activities of Saudi Arabian desert plants. *Phytopharmacology*, 2(1): 106-113.
38. Zain, M. E.; Awaad, Amani S.; Al-Othman, Monerah R.; Alafeefy, Ahmed M.; El-Meligy, Reham M. (2014). Biological activity of fungal secondary metabolites. *International Journal of Chemical and Applied Biological Sciences*, 1 (1): 14 22.