Survey of methicillin-resistant Strains of *Staphylococci* from Neonatal Septicemia for *mecA* gene

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Abstract: septicemia is still prevalent among neonates and it is a major medical problem. The aim of this study was survey of methicillin-resistant strains of *Staphylococci* from blood culture in neonate for *mecA* gene distribution. 138 blood cultures samples performed from neonates and identified the *Staphylococci spp*. These isolates were tested for antimicrobial susceptibility according to CLSI. Detection of mecA gene was performed by PCR. Among 138 samples from neonates 31.8% were positive blood culture for *Staphylococci* strains; of which 54.5% and 45.5% were *Coagulase negative Staphylococci* (CONS) and *Staphylococcus aureus* respectively. total 24 samples of CONS were biotyped, S. epidermidis (62.5%) and *S. saprophyticus*(37.5%). maximum resistance was seen with Ampicillin and minimum Resistance with Ciprofloxacin. prevalence of MRCONS was 55.6% and MRSA was 55%. The *mecA* gene was detected in 87% of the isolated CONS and 70% of *S. aureus* isolates. This study show that the high prevalence of methicillin resistance.

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Key words: Blood culture, Neonates, CONS, mec A, MRSA

Introduction

Septicemia, is a common condition in neonates with a resultant high mortality and morbidity rate in spite of new advances in antibiotic therapy(Yalaz,2006). Staphylococci are the most abundant isolated bacteria from blood. Since septicemia with Staphylococcus aureus is associated with a high mortality and an increased length of stay in hospital ,timely detection and identification of or coagulase-negative staphylococci S.aureus (CONS) including methicillin resistance from the patient's blood has great therapeutic, economic and prognostic significance(Gröbner and Kempf,2007). These strains carry the mecA gene, which encodes a modified penicillin-binding protein (PBP2a) that is responsible for resistance to B_-lactam antibiotics(Livermore,2000). Identification of methicillin resistance is performed by phenotypic and methods(Martins,2007).Today, genotypic the phenotypic method such as disk diffusion is used in most laboratories that Various factors affect on the growth of bacteria and results(Mirsalehian A,2003).Hence a sensitive and exact method is necessary that be independent from conditions of culture media. Isolation mecA gene is a useful marker for identification of resistance to oxacillin in Staphylococcus spp.PCR is a rapid and accurate method for isolation this gene.(De Giusti,1999) Hence, the present study was undertaken to survey of methicillin-resistant blood culture strains of *Staphylococci* from neonates for *mecA* gene distribution in Beasat hospital, Sanandaj, Iran.

Materials and Methods

Total of 138 blood samples were taken from neonates with symptoms suggestive of neonatal sepsis from the neonate ward were included in this study. The antimicrobial susceptibility testing for all the *Staphylococci* strains to various antimicrobial agents were determined according to the Clinical Laboratory Standards Institute.((.A,2007)

Oxacillin disc Diffusion test

To determine the antibiotic methicillin resistant strains of *Staphylococcus aureus*, strains on Mueller-Hinton agar medium 4% salt were cultured. Resistance to methicillin by disk diffusion method using oxacillin disk company mast Examined. The pattern of antibiotic resistant strains *MRSA* according to CLSI isolated.(.A,2007)

PCR application of the mecA gene

Detection of *mecA* gene was performed by PCR (Shubhra .S, 2009). Genomic DNA was extracted by a commercial extraction kit, Sina Gene Company, Tehran, Iran using the Eppendorf Master cycler. Both forward and reverse primer pair was used. The forward primer is GGAATGCAGAAAGACCAAAG while the reverse primer is CTTTGGTCTTTCTGCATTCCTG. Amplification was done using a thermal regime of 35 cycles of amplification at 95° C for 5 min, and 95° C for 45s which was for denaturation. Annealing temperature was set at 58 $^{\circ}$ C for 45s. The extension phase was done using a temperature of 72 $^{\circ}$ C for 1 min and the 2nd one was 72 $^{\circ}$ C for 10 min. A positive result was inferred by detection of a 500 bp band representing part of the *mecA* gene by electrophoresis on a 1% agarose.

Results

Among 138 blood samples from neonates, 44 were positive blood culture for *Staphylococci spp*,that 24 (54.5%) and 20 (45.5%) were *Coagulase negative Staphylococci* and *Staphylococcus aureus* respectively. (Table1). Total 24 samples of *CONS* were biotyped *S. epidermidis* 15 (62.5%) was the

most common species followed by *S. saprophyticus* 9 (37.5%).

The antibiotic susceptibility patterns of *Staphylococci* isolates are shown in Table(2). In Among *CONS* and *Staphylococcus aureus* 62.5% and 75% resistance was seen maximum with Ampicillin and minimum with Ciprofloxacin for both *CONS* and *S. aureus* which was 8.3% and 15% respectively. Among 24 *CONS*, 19 (79.1%) were meticiline resistance(MR) and among 20 *S. aureus* strains, 16 (80%) were MR by routine disc diffusion test using Oxacillin disc. prevalence of *MRCONS* was 55.6% and *MRSA* was 55%. Screening for *mecA* gene by using PCR method revealed that 21 (87%.) and 14 (70%) strains of *CONS* and *S. aureus* were positive for *mecA* gene respectively.

Table 1. Abundance distribution of Staphylococci spp from the neonate ward at Beasat hospital, Sanandaj, Iran

Isolated bacteria	Number of isolates	Percentage
Coagulase-negative staphylococci	24	54.5
S. aureus	20	45.5
Total	44	100

Table 2. Percentage of resistance of isolated <i>Staphylococci spp</i> from the neonate ward at Beasat hospital, Sanandaj,				
Iran				

	CoNS	S. aureus
Antibiotic	Rate of resistance (%)	
Sulfamethoxazole-trimethoprim	34.1	22.7
Gentamicin	29.5	34.0
Erythromycin	36.3	34.0
Tetracycline	29.5	36.3
Vancomycin	18.1	20.4
Ciprofloxacin	8.3	15.0
Clindamycin	25.0	30.0
Ampicillin	62.5	75.0

Discussion

Bacterial pathogens particularly Staphylococci spp pose a significant threat to human health generally neonates. (Shubhra .S,2009) In current study, from 138 blood cultures, 44 (31.9%) Staphylococci spp were isolated and identified, which 24 (54.5%) and 20 (45.5%) were *Coagulase negative* Staphylococci (CONS) and Staphylococcus aureus respectively. Many studies from elsewhere in the world still report that CONS are the most common organisms associated with neonatal sepsis(AlFaleh,2010). In previous study alfaleh (AlFaleh,2010) 55.11% and ghieb (Gheibi,2008)54% CONS isolated which is comparable to present study, while in study of Iran, kalantar et.al 65.78% CONS reported(Kalantar, 2007). Antimicrobial sensitivity pattern differs in different studies as well as at different times Iran and overseas in studies(Rahbar,2005, Yadegar,2009). In this study

the most common resistance in staphylococci spp was to ampicilin that About 75% for S.aureuse and 65% for CONS, also 80% S.aureus were resistant to Oxacillin. In study of torret S. aureuse resistance to ampicilin was 85%(Yano,2009) and in Philippine, rate of Oxacillin-resistant S. aureus from clinical specimens was 66 % (Arakama,2010). Moreover, because of widespread methicillin resistance among Staphylococci spp, the most frequent causative microorganism among neonates, and empiric Staphylococcal infection treatment of with vancomycin is advocated strongly in many neonatal wards (Kalantar, Motlagh, 2007, Gheib.S 2008). In our study, we observed that more than 20% resistance to vancomycin in S.aureus and in CONS 18%. Saderi reported Prevalence of Staphylococcus strains resistant to vancomvcin in iran (Shahrbanooie.2005).

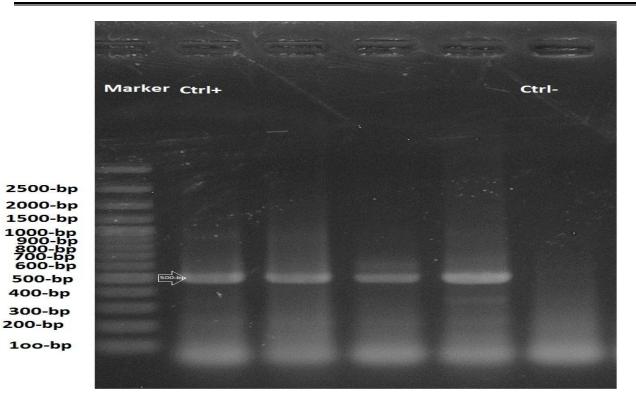


Figure 1. PCR analysis for the *mecA* gene among *Staphylococci spp* isolated from the neonate ward at Beasat hospital, Sanandaj, Iran. Line 1, DNA marker; line 2, 3,4 and 5 have *mecA* gene; Line 6 negative control, lacking *mecA* gene.

In Britain, France, the United state reports of outbreak strains VRSA observed(Tenover FC,2001) In our study, prevalence methicillin resistant rate in *MRCONS* was 55.6% that lower than from other previous reports done in other countries such as Turkey (74.4%),France (71%) (Khadri and Alzohairy,2010), and Iran 70% (Davoodi,2012). On the other hand, in this study, prevalence of *MRSA* was 55%.According to statistics in the United States, the prevalence of *MRSA* was 2% in 1980 and in 2004 was 60%(Lin Y,2007).frequency of *MRSA* in Asian countries such as India 44% (Kupfer,2010),Saudi Arabia was 8%(Broens,2011), and iran56%(21).

The high prevalence of *MRSA* and *MRCONS* isolates in Iran can be due insufficient infection control measures in hospitals and inappropriate use of methiciilin. We tested all isolated *Staphylococcus spp* for detection of *mecA* gene. The highest *mecA* gene carriage was found in *CONS* strains. 21 (87%) of 24 *CONS* strains from the neonates were *mecA* positive (Figure 1). Disk diffusion method could not recognized total staph strains that have *mecA* gene.One of the most reliable method for the identification of meticiline resistance is detection of *mec A* gene by PCR(Velasco,2005). In this study, 20 samples of *S.aureuse*, 14 strain were positive for *mec A* gene and 16 strain were meticiline resistance.

kolbert observed similar cases in *CONS* Despite the lack of *mecA* gene were meticiline resistance (Kolbert,1995) .This can due be production high amounts B-lactamase in strains. *CONS* predominated as the cause of methicillin resistance in our study. Our results shown that disk diffusion method is not an accurate method for the determination of methicillin susceptibility for *staphylococci spp* and a Rapid method as PCR for antibiotic susceptibility is important to institute appropriate therapy.

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References

1. Yalaz M, Çetin H, Akisu M, Aydemir S, Tunger A, Kultursay N. Neonatal nosocomial sepsis in a level-III NICU: evaluation of the causative agents and antimicrobial susceptibilities. Turkish Journal of Pediatrics. 2006;48(1):13.

- Gröbner S, Kempf V. Rapid detection of methicillinresistant *staphylococci* by real-time PCR directly from positive blood culture bottles. European Journal of Clinical Microbiology & Infectious Diseases. 2007;26(10):751-4.
- 3. Livermore DM. Antibiotic resistance in staphylococci. International Journal of Antimicrobial Agents. 2000;16:3-10.
- Martins A, DE Lourdes M, Cunha R. Methicillin resistance in *Staphylococcus aureus* and *coagulasenegative staphylococci*: Epidemio logical and molecular aspects. Microbiology and immunology. 2007;51(9):787-95.
- Mirsalehian A JF, Alizadeh S. Comparison of disk agar diffusion susceptibility testing and PCR in detection of methicillin resistant *Staphylococcus* aureus. Med J Tehran Univ Med Sc. 2003;61(6):420-
- De Giusti M, Pacifico L, Tufi D, Panero A, Boccia A, Chiesa C. Phenotypic detection of nosocomial mecApositive *coagulase-negative staphylococci* from neonates. Journal of antimicrobial chemotherapy. 1999;44(3):351-8.
- 7.. A M. performance standards for antimicrobial suseptibility testing seventeenth information suplement. 2007;27(1).
- Shubhra .S GB, S.K. Agarwal, Anuradha Rajput, Piyush Tripathi, Mala Kumar, Shraddha Singh, R.K. Singh. Prevalence of Mec A Gene positive coagulase negative *Staphylococci* in NICU of a tertiary care hospital. Biomedical Research 2009;20(2):94-8.
- 9. AlFaleh KM. Incidence of Late Onset Neonatal Sepsis in Very Low Birth Weight Infants in a Tertiary Hospital: An ongoing challenge. Sultan Qaboos University Medical Journal. 2010;10(2):227.
- Gheibi SKM. Coagulase Negative *Staphylococcus*; the Most Common Cause of Neonatal Septicemia in Urmia, Iran. iranian journal of pediatrics. 2008;18:(3)237-43.
- 11. Kalantar E, Motlagh M, Lordnejad H, Beiranvand S. The prevalence of bacteria isolatedfrom blood culturesof iranianchildrenand study of their antimicrobial suseptibilities. Jundishapur Journal of Natural Pharmaceutical Products. 2007;2008(01, Winter):1-7.
- Rahbar M, Gra-Agaji R, Hashemi S. Nosocomial blood stream infections in Imam Khomeini Hospital, Urmia, Islamic Republic of Iran, 1999-2001. East Mediterr Health J. 2005;11(3):478-84.
- 13. Yadegar A, Sattari M, Mozafari NA, Goudarzi GR. Prevalence of the genes encoding aminoglycosidemodifying enzymes and methicillin resistance among clinical isolates of *Staphylococcus aureus* in Tehran, Iran. Microbial Drug Resistance. 2009;15(2):109-13.

- 14. Yano K, Minoda Y, Sakawa A, Kuwano Y, Kondo K, Fukushima W, et al. Positive nasal culture of methicillin-resistant *Staphylococcus aureus* (MRSA) is a risk factor for surgical site infection in orthopedics. Acta Orthop. 2009 Aug;80(4):486-90.
- 15 .Arakama M-H, Mendoza M, Patrick Fernandez R, Belmonte L, Galapia Y, Pile M. Emergence of Methicillin-resistant *Staphylococcus aureus* among Patients in a Tertiary Renal Medical Center. Philippine Journal of Microbiology and Infectious Diseases. 2010;39(1).
- 16 .Gheib.S FZ, Karamyyar . M , Khashabi.J ,Ilkhanizadeh .B *Coagulase Negative Staphylococcus*; the Most Common Cause of Neonatal Septicemia in Urmia, Iran. Iran Journal Pediatric. 2008; 18 (3):237-43.
- Shahrbanooie R. Vancomycin resistance among clinical isolates of *Staphylococcus aureus*. Archives of Iranian Medicine. 2005;8(2):100-3.
- Tenover FC BJ, Lancaster MV. increasing resistance to vancomycin and other glycopeptides in Staphylococcus aureus. Emerg Infect Dis. 2001;7(2):327-32.
- 19 .Khadri H, Alzohairy M. Prevalence and antibiotic susceptibility pattern of methicillin-resistant and *coagulase-negative staphylococci* in a tertiary care hospital in India. Int J Med Med Sci. 2010;2:116-20.
- 20. Davoodi NR, Yousefi JV, Harzandi N, Hajrafi A, Rajaei B, Gerayesh-Nejad S, et al. Molecular detection of methicillin resistant *Staphylococcus aureus* (MRSA) and methicillin resistant *coagulase-negative Staphylococcus* (CoNS) in Iran. African Journal of Microbiology Research. 2012;6(16):3716-21.
- 21 .Lin Y LT, Lin H, Chen P, Cheng M. An outbreak of methicillin-resistant *Staphylococcus aureus* infection in patients of a pediatric intensive care unit and high carriage rate among health care workers. J Microbial Immunol Infect. 2007;40:325-34.
- 22. Kupfer M, Jatzwauk L, Monecke S, Mobius J, Weusten A. MRSA in a large German University Hospital: Male gender is a significant risk factor for MRSA acquisition. GMS Krankenhhyg Interdiszip. 2010;5(2).
- 23. Broens EM, Graat EA, Van der Wolf PJ, Van de Giessen AW, De Jong MC. Prevalence and risk factor analysis of livestock associated MRSA-positive pig herds in The Netherlands. Prev Vet Med. 2011 Oct 1;102(1):41-9.
- 24. Velasco D, del Mar Tomas M, Cartelle M, Beceiro A, Perez A, Molina F, et al. Evaluation of different methods for detecting methicillin (oxacillin) resistance in *Staphylococcus aureus*. Journal of Antimicrobial Chemotherapy. 2005;55(3):379-82.
- Kolbert C, Connolly J, Lee M, Persing D. Detection of the *Staphylococcal mecA* gene by chemiluminescent DNA hybridization. Journal of clinical Microbiology. 1995;33(8):2179-82.

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