

Feeding European Sea Bass (*Dicentrarchus labrax*) With Trash Fish

1- Growth Performance and Reproductive Histology

Ahmed K.I.El-Hammady¹; Seham A. Ibrahim²; Mohammed A.I.Wafa¹ and Fawzia A. El-Ghamadi³

¹ Fish Nutrition Lab, National Institute of Oceanography and Fisheries, Cairo, Egypt.

²Department of Zoology, Faculty of Science, Benha University, Benha, Egypt.

³Department of Zoology, Faculty of Science, King Abd El-Aziz University Jeddah, KSA

akelhammady@yahoo.com

Abstract: The effects of exposing commercial feeds for European sea bass (*Dicentrarchus labrax*) were examined to lower prices diet. Trash fish used as aqua feed were submitted to six feeding schedules in net pen cultivation (40 fish/net pen). Each treatment had two replicate net pen. The present study was performed to investigate the influence of individual fed local trash fish (8.6% of body weight) or dry pellet diet (2.2% of body weight) with four feeding schedules (Fs) as fed European sea bass (a range initial body weight from 19.57 to 24.85g) for 140 days. Fs1: 1.1% of body weight (BW) dry pellet at 9.00am and 4.3% of BW. Local trash fish at 14.00pm; Fs2: one day 2.2% of BW. Dry pellet diet consecutive another day 8.6% of BW. Local trash fish; Fs3: one day 2.2% of BW Consecutive two days 8.6% of BW Local trash fish; Fs4: two days 2.2% of BW. Dry pellet diet consecutive one day 8.6% of BW local trash fish. Growth performance, survival rate, proximate analysis of dorsal musculature and some somatic parameters of European sea bass were measured to assess four feeding schedules compared to fish fed only local trash fish or dry pellet diet. The improved of weight gain percentage, geometric mean were obtained in fish fed with feeding schedule No. 4(2 day fed dry pellet diet with one day feed local trash fish) compared to other feeding schedules. The survival rate of *D. labrax* feeding schedule No. 4were higher compared to other fish feeding schedule 1, 2, 3. Proximate analysis of dorsal musculature of different feeding schedules showed significantly ($P < 0.05$) concentrations of lipid compared to fish fed only local trash fish as dry pellet diet. Small, round and transparent oocytes with a central nucleus were observed in histological sections of ovaries. Though, testes based on microscopic observation were distinguished for male *D.labrax* and spermatogonia and primary spermatocytes were the dominant cells of this stage. The results of this study revealed that the overlap between local trash fish with dry pellet diet under feeding schedule No.4, could be used in fed European sea bass (*D. labrax*) to give better growth performance.

[Ahmed K.I.El-Hammady; Seham A. Ibrahim; Mohammed A.I.Wafa and Fawzia A. El-Ghamadi: **Feeding European Sea Bass (*Dicentrarchus labrax*) With Trash Fish. 1. Growth Performance and Response Reproductive.** *Life Sci J* 2014;11(9):568-583]. (ISSN:1097-8135). <http://www.lifesciencesite.com>. 94

Key Words: *Dicentrarchus labrax*, artificial diet, trash fish, growth performance, histology of ovary and testis.

1. Introduction:

Economically productive aquaculture systems depend upon an adequate supply of low-cost feeds with high nutritional quality. Feed is considered as the most expensive single factor since feed cost constitutes at least 50% of the total production (Posadas, 1988).

Trash fish, as commonly defined is that portion of the catch that by virtue of their small size or low consumer preference has little or no value (Sugiyama *et al.*, 2004). The same authors, showed that the use of term trash fish varies from country to country and can change both seasonally and with locations. Xu *et al.* (2007) mentioned that trash fish, mainly from capture fisheries, are not consumed by human due to their low protein content and relative unpalatability. Trash fish are often discarded as by-catch, with potential environmental and aesthetic problems (Li *et al.*, 2004). In aquaculture, trash fish fed directly

to carnivorous fish or as an ingredient of artificial feed are considered better alternatives (Xu *et al.*, 2007).

Bunlipatanon *et al.* (2004) reported that better cost-benefit and resource use, to fed Asian sea bass (*Lates calcarifer*) and tiger grouper (*Epinephelus fuscoguttatus*) were recorded for fish reared on trash fish. Hung and Mao (2010) studied the effect of different trash fish with alginate binding on growth and body composition of juvenile cobia (*Rachycentron conadum*). They ground raw fish and extruded by an extruder. Sodium alginate was used as a binder for all moist diets at a concentration of 3%. After extrusion, the moist diets were submersed in 10% CaCl₂ solution to gel through the strong binding of calcium and alginate for 10 minutes. Feeds were sealed in vacuum packed bags and stored frozen (-20°C) until feeding. Peres and Oliva-Teles, (1999) found that the optimum protein level in the diets for sea bass juveniles was estimated to be around 50%.

An important number of studies has been developed on the relations between nutrition and reproduction in several fish farmed species and some effort has been paid on the development of fish farming (Asturiano *et al.*, 2006). In several studies have shown that n-3 series enriched diets have profound effects on female reproduction, influencing pattern of gonadal development, plasma levels of lipids and sex steroids, egg quality and lipid levels, fecundity, hatching and survival rates of European sea bass (Asturiano *et al.*, 2006). Though, Asturiano *et al.* (2001) described the influence of series n-6 PUFAs on reproduction of teleost fish. Chaitanawisuti *et al.* (2011) fed the broodstock spotted Babylon (*Babylonia aerolata*) by trash fish once daily at 10:00h with the daily amount calculated as 15% of the total broodstock biomass per tank (excess diet was removed and feeding rate was adjusted based on weight gain after each sampling, which was done every (2 weeks). The mechanism linking gonad development and reproductive performance in order to improve culture techniques (Cek and Yilmaz, 2007). The same authors showed that one of the most important factors necessary in the successful culturing of a fish species is obtaining a basic understanding of its key biological processes. The most important of these biological processes is the fish's reproductive cycle and formation of gametes.

A number of reports have demonstrated that gonad development in fishes is highly sensitive to exposure to substances with oestrogenic activity (Maack and Segner, 2003). The structural alterations induced by exposure to environmental oestrogens can include the development of ovo-testes, the manifestation of gonadal malformations and histopathological alterations, or the increased frequency of atretic oocytes (Nolan *et al.*, 2001). Mahmoud (2009) reported that a thorough study of gonad anatomy and histology is required for proper management of the fishery.

The present work tries to define the course of local trash fish and dry pellet diet as fed in European sea bass (*Dicentrarchus labrax*) according to changes in four feeding schedules. This is considered in association to lowering the costs of feeding and the environmental risks.

2. Materials and Methods

Experimental fish and rearing condition

The experiment was carried out in twelve (12) net pen culture (diameter 4.0m, depth water 1.2m.) inside commercial farm (earthen pond) at the Port Said governorate, Egypt. Juvenile European sea bass (*Dicentrarchus labrax*) were obtained from a local commercial fish farm 480 homogeneous fish with a range of initial body weight from 24.85 to

19.57g and distributed in 12 net pen at 40 fish per net pen. Fifteen-days acclimated to the commencement of the experimental period. During the acclimation period fish were gradually fed the dry pellet diet by starving them, followed by feeding local trash fish plus dry pellets, then progressively reducing the trash fish until dry pellets were completely accepted as has been by Stirling (1977). Fish were hand fed in plastic box. The feeding trials were conducted for 140 days (from June to October, 2012).

Experimental diets and feeding schedule

The experiment was set up a six different diets fed in duplicate. The experimental diets consisted of one pelleted diet and one local trash fish with four different feeding schedule. The feeding rate from dry pellet diet was 2.2% of body weight and to compensate feeding rate from local trash fish 8.6% of body weight (for the higher water content), respectively. Daily ration were fed twice a day (equal meal at 9.00 am and 14.00pm h) according to feeding rates. Two test diets contained dry pellet diet and local trash fish, whilst in four feeding schedule were as follows:

Feeding schedule one (FS₁) was daily ration fed twice (1.1% of body weight dry pellet diet at 9.00am and 4.3% of body weight local trash fish at 14.00pm).

Feeding schedule two (FS₂) was daily ration, one day 2.2% of body weight dry pellet diet and another day 8.6% of body weight local trash fish.

Feeding schedule three (FS₃) was ration one day 2.2% of body weight, dry pellet diet and follow two day 8.6% of body weight /day, of local trash fish.

Feeding schedule four (FS₄) was ration two day 2.2% of body weight dry pellet diet and follow one day 8.6% of body weight local trash fish.

Diet formulation and trash fish preparation

Local trash fish were collected from fishermen of Lake Manzala (belonging to Port Said Governorate, Northern Egypt). Local trash fish were frozen in a cold storage at -10C° respectively. Before feeding, carcasses of frozen fish were minced by meat grinder into pieces and thawed for two hour respectively before feeding. Ingredients diet (obtained from a local feed company) Proximate composition and formulation (Table 1,2). Proximate composition of the local trash fish and formulate diet are shown in Table (3). All ingredients were exposed to grinding, mixture and processed into a California pellet Meal machine (CPM) as dry pelleted diet.

Growth trial

At the beginning of growth trial the feeding rate was adjusted based on weight gain after each sampling, which was done every 28 day.

A representative sample of ten fish was withdrawn and kept frozen (-4C°). Growth trial was

conducted for 20 weeks (June-October) and every fourth weeks fish in each net pen were bulk-weight and counted to follow, growth and feed intake. At the end of the growth study and after an overnight fast, all fish from each net pen were individually weighed, total length measured and calculated to determine survival rate (%). Though fish from each net pen were randomly sacrificed and pooled for dorsal **musculature** composition analysis, weighed liver, and gonad (ovary, testis) for **determining** somatic-index and **examined** histological **histologically** for gonad **and** intestine.

Chemical analysis

Samples of feed ingredients diets and fish muscle were analyzed in triplicate using standard methods (AOAC, 1997). Dry matter was determined by drying in an oven at 105°C for 24 h. Nitrogen (N) was determined by the kjeldahl method and crude protein (CP) was calculated as NX 6.25. Crude fat (EE) content was analyzed using the soxhlet method with petroleum ether (bp 40 to 60°C) crude fiber (CF) content was determined by standard method (AOAC, 2000). Ash content was determined by incineration in a muffle furnace at 550°C for 12 h. Gross energy was calculated by Martinez – Liorens *et al.* (2007) as energy coefficients: protein 23.9kJ/g, lipid 39.8kJ/g and carbohydrate 17.6kJ/g. Nitrosen free extract (NFE) was computed by taking the sum values for crude protein, lipid, ash, fiber and moisture, and subtracting this from 100.

Gonads were preserved after dissection in 10% formalin, dated and labeled with all the necessary data for subsequent examination. Paraffin embedded samples of the ovary and testis were sectioned to 6µm in thickness, and stained with Haematoxylin and eosin (H and E) Bernet *et al.* (1999). They were examined and photographed by a built in Camera.

Water quality monitoring

Water temperature at 20cm depth (TC°) was recorded **daily** with a temperature meter at 9.00h, range **ranged** (31.9-25.3 C°).

Dissolved oxygen (DOmg.L⁻¹) was measured by instruction Manual, Hanna instruments (HI9146), USA (Dissolved oxygen and temperature meter) a weekly (range 6.3-4.5mg/L). Salinity (‰) was measured by Manual Handle ding consort (C 860) bvba, Belgium. a weekly (Range 22.1-14.5‰); pH was measured a weekly by Adwa instruments (A Dlllo and A Dlll) pH meters Hungary a weekly (range 7.9-7.2)

Calculations

The following calculations were **used**:

Weight gain (%) =

$$\frac{(Final\ body\ weight - Initial\ body\ weight) \times 100}{Initial\ body\ weight}$$

Feed efficiency (%) =

$$\frac{(Fish\ weight\ gain \times 100)}{Feed\ intake\ (dry\ matter)}$$

Daily feed intake (percentage of body weight) =

$$\frac{Feed\ intake\ (dry\ matter) \times 100}{[(initial\ fish\ wt.\ final\ fish\ wt.) \times days\ fed / 2]}$$

Protein efficiency ratio (PER) =

$$\frac{Fish\ weight\ gain}{Protein\ intake}$$

Condition factor (CF)

$$\frac{(Body\ weight \times 100)}{Total\ body\ length\ (cm)^3}$$

Specific growth rate (SGR %) =

$$\frac{[(Ln\ Wf - Ln\ Wi) / T] \times 100}{}$$

Where Wf and Wi refer to the mean final body weight and the mean initial weight, and T is the feeding trial period in days.

Survival rate (%) =

$$\frac{(TNf / TNi) \times 100}{}$$

Where the TNf is total number of fish at finish and TNi is total number of fish at start.

Daily growth index, cited by Kaushik *et al.* (2004),(DGI)=

$$\frac{100 \times (Final\ body\ weight^{1/3} - Initial\ body\ weight^{1/3})}{Time\ days}$$

Mean weight (Geo metric mean),cited by Lupatsch *et al.* (2001),=

$$\frac{[(Initial\ body\ weight\ (g)) \times (Final\ body\ weight\ (g))]^{0.5}}{}$$

Total feed intake per fish =

$$\frac{Total\ feed\ intake\ (g)}{Number\ of\ fish}$$

Protein intake =

$$[Feed\ intake\ (g)] \times [Protein\ in\ the\ diet\ (%)]$$

Hepato-somatic index (HSI) =

$$\frac{100 \times (Liver\ weight\ (g))}{Whole\ body\ weight\ (g)}$$

Gonado – somatic index (GSI) =

$$\frac{100 \times (Weight\ of\ gonads(g))}{Whole\ body\ weight(g)}$$

Statistical analysis

All data on fish growth performance, feed utilisation and muscle traits were statistically analysed by one-way analysis of variance (ANOVA), using **tests performed by Duncan (1955)** for individual comparison (P< 0.05) level of significance). Statistical analysis were carried out using the **IBM SPSS statistic (2011)** programme, version 19.

3. Results and Discussion:

The interpretation of data should be done with care because of some major problems encountered which led to experimental errors that could have masked the treatment effect. Fishery products, either in the form of low-value trash fish or rendered as fish meal, are presently the major sources of protein in the grow-out culture of most fish species and constitutes up to 70% by weight of their diet (Tacon, 1995). A dependable supply of cost-effective, non-marine alternative sources of protein must be provided for fish farming to be profitable (Millamena, 2002).

The results presented in table (4) demonstrated effect trash fish on growth during 140 days (20 weeks). One reason for this duration, **Yigit *et al.* (2003)** showed that studies of protein and amino

acid requirements of fish are usually conducted for an experimental period of 8-12 weeks. It is that statistical differences in the commonly measured growth criteria may not become apparent. Other responses such as feed intake and feed efficiency are not reliable criteria because of the difficult in collecting accurate data (Lovell, 1989).

Specific growth rate, weight gain percentage, mean weight (geometric mean) and daily growth index were equation to examine growth performance of European sea bass fed the dry pellet or local trash fish with four different feeding schedule (Table 4). The final body weight and gain of sea bass fed a treatment (FS 5.4) was significantly increased ($P < 0.05$) than fish fed only local trash fish treatment (6, fed only trash fish). However, **there** was no significant differences ($P > 0.05$) among treatment 5, feeding schedule 4, and treat fish fed only dry pellet diet (DP) respectively.

Literature data on the growth of Mediterranean marine **finfish** species appear to be rather highly variable, probably due to differences in fish strains, water quality and temperature, oxygen availability, biomass density and biological value of dietary protein and non-protein energy substrates (Company *et al.* 1999). In the present study, SGR (1.194-1.140% day) of European sea bass (Table 4) were better or within the range of previous study (1% by Russell *et al.* 1996; 0.7-0.8% by Dias *et al.* 1998, however 1.6-2% by Perez *et al.*, 1997).

In this connection, Tubongbanua (1987) study development of artificial feeds for sea bass (*Lates calcarifer*) who found that the mean biomass production of sea bass fed a trash fish diet was 71.7 kg/h for pasteurized trash fish diet, 70.5 kg/ha for sun-dried trash fish diet, 70.5 kg/ha for sun-dried trash fish, and 63.4 kg/ha for raw fish. **They also** showed that sea bass fed the pelleted fish meal had a production of 7.04 kg/ha while mashed fish meal gave only 56.4 kg/ha. These results appeared that the pasteurized trash fish had a better production compared to raw and sun-dried trash fish, this may be attributed to the deactivation of enzyme thiaminase and destruction of some harmful bacteria during heating, thus improving the feed quality of pasteurized trash fish over that of sun-dried and raw trash fish. This observation was also reported by Lovell (1979) for catfish fed with pasteurized, raw and sun-dried catfish waste. This may be due to some unknown growth factors in trash fish which could have been degraded during processing it into fish meal, the differences in enzyme thiaminase activity, or due to differences in the organoleptic attractiveness of the two products (Tubongbanua, 1987). Though, Fagbenro (1994) found that tilapia fed dried fermented fish silage, can assimilate protein as amino

acids and short peptides, so the protein breakdown during the treatment could have a positive effect on the digestibility and assimilation of this nutrient. In the present study considering the high acceptability of the trash fish diets, the use of trash fish in diets might act as a natural feed stimulant (Plascencia-Jatomea *et al.*, 2002). Though, they reported that small amounts of silage improved the growth efficiency of Atlantic salmon. This behavior was explained by the authors through the low levels of proteolytic enzymes within the gastro intestinal tract of the fry.

Results presented in table (4) showed that specific growth rate (1.168%/day) of fish had dry only dry pellet diet (Treat 1) control (1) decreased than treat 2,3,5 respectively. This is may be due to increased dry pellet intake $168.84 \pm 7.48g$ as fed (Table 5) which **contained** some vegetable ingredients (Table, 2). A similar trend was reported by Geay *et al.* (2010), **who** examined regulation of FADS2 expression and activity in European sea bass (*Dicentrarchus labrax*, L.) fed a vegetable diet. They observed a significant difference ($P < 0.05$) in fish final weight between dietary treatments with values of + 17.5% for fish fed fish diet by comparison fish fed 100% vegetable diet. This diminution of growth performance can be correlated to the significant decrease in n-3 HUFA content observed in flesh of fish fed vegetable diet (Parpoura and Alexis 2001). Another reason could be linked to the low arachidonic acid content of the vegetable diet compared to fish diet. Indeed, an arachidonic acid requirement has been reported for growth in different fish such as turbot (Castell *et al.*, 1994). Though, the supply of protein as vegetable meals could be another reason explaining the lower final weight of the European sea bass (Geay *et al.*, 2010), since it has been previously shown that fish meal replacement by vegetable protein can reduce **growth** in this species (Tibaldi *et al.* 2006). The lowest growth trend was obtained with treatment (6 fish fed only local trash fish) (Table 4) may be to increase chitin (scales) have an adverse effect on body weight as shown in studies reported by Plascencia-Jatomea *et al.* (2002).

Results presented in table (4) showed that final body weight, weight gain and specific growth rate were slightly decrease in sea bass (*D. labrax*) fed only dry pellet diet than fish fed schedule FS4(2 day dry pellet diet+ 1 day trash fish). This is may be due to increase carbohydrate intake (Fig.1) with fish fed only dry pellet diet(DP) than other treatment. There is a general consideration that carnivorous fish like European sea bass do not digest carbohydrates very efficiently (Krogdahl *et al.* 2005). Moreover, dietary starch level may negatively affect carbohydrate digestibility and may also interact with the digestibility of other dietary nutrients (Stone, 2003).

According to **Enes et al. (2006)** who did not observe any growth improvement in European sea bass juveniles fed diets including carbohydrates comparatively to fish fed a carbohydrate-free diet. Also, **Altan and Yildirmkorkut (2010)** noted that dietary starch level (10-30%) did not affect growth with sea bass (*D. labrax*).

The gain and the specific growth rate was significantly ($P < 0.05$) lower in fish fed only trash fish (T6) (Table 4), it could be due to the high level of lipid intake (94.46g) (Fig.1) which inhibiting the transamination of amino acid, this has also been reported by **Giri et al(2000)**.

However, a lower of growth (gain and SGR) was noticed in fish fed on only pellet dry diets reported that it may cause decreasing the enzyme activity (Protease activity) with fish fed on plant protein (**Venkatesh et al., 1985**). Though, a decrease growth in treat (1) may be due to increase carbohydrate/protein intake ratio 0.273 in treat (1) (Fig.1) which correlated inversely with apparent digestibility coefficient (**Fernandez et al, 1998**).

Differences in weight gain and SGR between of the sea bass fed only trash fish for (Treat. 6) or fed only dry pellet diets (Treat 1) (Table 4) were significantly ($P < 0.05$) which may be affected by dietary moisture content. (Table 3). In this connection, effect of dietary moisture content on performance of fish varies depending on fish species (**Lee et al., 2000**) who reported that increase of moisture up to 30% of the diet reduced weight gain of Juvenile Atlantic salmon.

Though, **Ekanem 1996** reported that estuarine catfish fry (*Chrysihthys nigrodigitatus*) grew better when fed a dry diet than moist diet, which probably results in improved water quality in the rearing pond. **Munsiri and Lovell (1993)** showed that the properties of the diet such as its composition of ingredients or its moisture content is affected on the gastric content of fish. Moreover, **Higgs et al. (1985)** reported that marine fish prefer moist feed to dry feed for its osmoregulation between body and medium, so that dietary moisture content may affect the performance of fish.

These facts reveal the increased weight gain of European sea bass (Table 4) treated with the feeding schedule 4(2 day dry pellet diet with 1 day trash fish) are mainly attributed to using a blend of dry pellet diet and trash fish which can be used to balance dietary essential amino acid content. In this connection, essential amino acid (EAA) deficiency is one of the factors limiting the utilization of economics protein sources as fish meal substitutes (**Glencross et al., 2007**). Compared to fish meal, poultry by products meal (BM) and, meat and bone meal (MBM) are lower methionine and lysine contents and feather mea

(FM) lower in methionine, lysine and histidine, (**Hertrampf and Piedad-Pascual, 2000**). Also, blood meal is rich in lysine and can be used to balance dietary lysine content when poultry by product, meat and bone meal and feather meal are used, alone or in combination, as fish meal substitutes (**Milliamena, 2002; Guo et al., 2007**).

Decrease weight gain (Table 4) of *Dicentrarchus labrax* fed only dry pellet diet (T1) may be attributed to incorporated meat and bone meal (MBM) in diet. **Robaina et al. (1995)** reported gilthead sea bream had low digestibility to the diets incorporated in MBM. **Millamena (2002)** suggested that high ash content in diets (content meat meals) may lower the digestibility of the diets that may have further caused the lowering in growth rates. However, **Xue and Cui (2001)** found that destroyed palatability has been demonstrated responsible to the reduced feed intake and growth of fish fed the diets in which high levels of dietary fish meal was replaced with economic plant or animal protein ingredients.

The reduced growth of *D.labrax* fed only dry pellet diet (Treat, 1) Table (4) than fish fed schedule 4 may be attributed to increase feed consume from fiber (11.929) for fish treat.2(Fig.1). **Oberleas and Harland, (1977)** showed that a higher fiber, content in diet which can decrease transit time of intestinal contents (increase fecal nitrogen and lipid excretion as reflected by lowered protein and energy digestibility. Though, **Poston (1986)** observed for rainbow trout a growth depression with a level of cellulose incorporation above 8%. **Shiau and Liang (1994)** reported a better utilization of dietary protein when they tested agar supplementation at two dietary protein levels in hybrid tilapia. They suggested that this effect could be related to a slower feed passage time through the gastro intestinal tract, that might enhance overall nutrient absorption. However, **Dias et al. (1998)** found that the dietary of silica, cellulose or natural zeolite as bulk agents at 10 and 20% level had no adverse effect on sea bass (*D. labrax*) growth.

In the present study, fluctuation in water salinity (range 22.1-14.5per thousand) may affect on growth and feed consumption. In their study, **Partridge and Jenkins(2002)** found that, the European sea bass fingerlings grew about 20-30% more in sea water than those reared in fresh water. They show that the reason for this is thought to be due to the fact that fish in fresh water spend more energy than those in sea water for osmoregulation as freshwater species have to excrete higher amount of water from body to the hypo-osmotic environment. Though, different kind trash fish may effect on growth performance. **Hung and Mao (2010)** found that different kind trash fish (used three species of trash fish: A-anchovy, L-Lizard fish; C-cardinal fish) and

those combinations (50% A+ 50%L; 50% A + 50% C; 50% L + 50% C). **These** different diets had significant differences on weight gain; specific growth rate, feed conversion ratio and survival of juvenile cobia ($P < 0.05$). Though, whole body proximate composition (crude protein, lipid and moisture) of cobia was affected as different fed trash fish. **Asturiano et al. (2001)** fed male European sea bass (*D. labrax*) on a wet diet (WD) consisting of trash fish 71% crude protein bogue (*Boops boops* L.), squid 79% crude protein (*Loligo vulgaris*) and fed two commercial **pelleted** diet (53% crude protein) enriched with polyunsaturated fatty acids (PUFA). They reported that the improved growth of the WD fed fish could be related to the higher dietary protein level of their food. Though, the same authors found that the WD diet consisted of two components bogue and squid, with the total percentages of saturated, monounsaturated and polyunsaturated fatty acids similar respectively in both components and the total percentage of PUFAs was highest in WD which also contained the largest percentage of total n-3 PUFAs as compared to two commercial pelleted diet. They added that since total lipid content was similar in three diets (wD and two commercial diets), it is possible that the PUFA lipid composition may be influencing reproductive performance.

A little variation in the survival rate in **the** treatments (Table 4) was observed. **Teng and Chua (1978)** noted that starvation occurs among fish populations where size hierarchy was established. The physical presence of large individuals inhibited the smaller fish from feeding satisfactorily and the dominance of feeding and space by a few larger fish in a restricted culture can cause death of smaller individuals due to starvation. Further, it was also possible that the larger sea bass (*Lates calcarifer*) preyed on the smaller ones thus resulting in a decline in survival on some of the treatments (**Tubongbanua, 1987**).

The proximate analysis of **musculature** of the sea bass (*Dicentrarchus labrax*) at different feeding schedules are shown in Table (5), insignificant effect on the moisture, protein and lipid content ($P > 0.05$) in both treatments (treat.6 or treat.1) (Table 5). With increase dietary moisture content in fresh fish (T.6) comparatively by sea bass (*D. labrax*) fed only dry pellet Table 3) diet (T.1) had no significant effect ($P > 0.05$) on proximate composition of dorsal **musculature** (Table 5).

A similar trend was found by **Lee et al. (2000)** who **examined** the effect of dietary moisture content on growth body composition and gastric evacuation of juvenile Korean rockfish (*Sebastes schlegeli*). **Also, they showed** that dietary moisture

content had no significant effect on protein and lipid content of muscle ($P > 0.05$).

Muscle body proximate composition of European sea bass (*D. labrax*) was presented in Table (5). Compared with body composition of sea bass (*D. labrax*) at beginning trial, crude protein, and lipid concentration increased while ash and moisture concentration decreased at the end of experiment when fish fed only trash fish. On the contrary, **Hung and Mao (2010)** who noted that whole body proximate composition of juvenile cobia fed trash fish was differences at beginning trial than at the end of experiment. According to **Huy (2002)** who used trash fish that had low lipid content, caused lipid reducing in muscle of cobia. **Grigorakis et al. (2003)** show that muscle proximate composition of wild fish gilthead sea bream (*Sparus aurata*) were found to have lower lipid and higher water content than cultured fish.

With increase feed intake of lipid in *D. labrax* fed only trash fish (Table 5) had decreased concentrations of body fat (Table 5). **Alvarez et al. (1998)** found that European sea bass **revealed** no significant effect of dietary fat (8-18%) with respect to the total and neutral intramuscular fat content in the dorsal muscle was detected.

Crude protein content of muscle European sea bass (Table 5) was not affected by the various fed schedule treatments. This agrees with **the** finding where by feed compositions were observed to have relatively little effect on the whole-body protein of humpback groupers (**Shapawi et al., 2011**).

Feeding fish with local trash fish (Treat. 5) resulted in slightly higher body moisture and the lowest of body lipid content compared to European sea bass fed only the pelleted feeds (treat. 1), table (5). Moreover, these results corroborate previous reported data (**Shapawi et al. 2011**). Concerning the effects of feeding consume (dry basis), protein and lipid content in muscle *D. labrax* was slightly **increased** with increase feeding consume for fish treat 5 (FS.4) comparatively by treat.1(DP) (Fig.1). No clear trend in moisture and ash content in sea bass **musculature** was observed at the end of the growth period (Table 5). This situation has already been demonstrated with sea bass by **Hidalgo et al. (1987)**, in which case, an increase in feeding rate from 0.74 to 1.45% bw day⁻¹ caused a rise in the lipid content of the fingerlings at 15C°, whereas no significant effect of ration on the lipid content (1.0% to 2.6 bw day⁻¹) was detected at 20C°. **Mihelakakis et al. (2002)** and **Van Ham et al. (2003)** found that body lipid is reduced when the feeding rate is lowered. A similar trend was reported by, **Hillested and Johnsen (1994)** who noted the fat in the fillet was higher as protein content of the diets decreased to 35%, which corresponded to a Digestible crude protein/ Digestible energy ratio of 14.8. Under protein

restriction, **Schwarz et al. (1985)** found that carp accumulated proportionally more dry matter, fat and energy. **Kaushik and Luquet, (1984)** who fed trout adipitium with a non-protein energy source and restricted protein led to increasing levels of fat in the carcass.

Indices of condition, such as condition factor (K) and hepato-somatic index (HSI) are often used to assess the nutritional status of fish because they can be determined easily and may provide an indication of physiological condition (**Mihelakakis et al. 2002**). In the current study at the end of the experiment, considering the result of condition factor, (Table 6) showed that *D. labrax* with FS.4 consumed more feed have slightly increase in CF than other treat. **Eroldogan et al. (2004)**, show that an increase in feeding rate was reflect insignificantly increase in condition factor due to deeper body shape.

In the present study (Table 6) HSI with fish treat.5 (FS.4) had increased significantly ($P < 0.05$ respectively) with increase food supply (Fig.1). Similar observations on HSI have been reported in *Sparus aurata* (**Mihelakakis et al. 2002**) who found that HSI were significantly higher with feeding rate 2% / day, comparatively with feeding rate up to 0.57 % / day. Hepato-somatic index (HSI) in fish usually suggests problems in nutrition because the relative size of the liver is correlated with nutritional state of the fish. A similar decrease in HSI was observed in striped bass when feeding rate was reduced from 1.0 to 0.5 BW.d (**Hung et al., 1993**). On the other hand, hepato-somatic index in treat.1 (fed only dry pellet diet) was significantly ($P < 0.05$) increased than treat (6) fed only trash fish (Table 7). This may be due to decrease fishmeal as protein source in dry pellet diet and substitute by plant protein source, soybean and corn gluten meal, (Table 2). **Dias, (1999)** showed that hepatic fat deposition indeed is high, there is evidence that replacement of fish meal by plant protein source such as corn gluten meal or soy protein concentrates affects hepatic lipogenic enzyme activities variably in sea bass, while the activity of malic enzyme decrease, that of fatty acid synthetase increased significantly with high level of corn gluten meal in the diet. **Shapawi et al. (2011)** stated that HSI is related to the nutritional state of fish and may directly related to energy requirements for growth and found that poor growth of grouper (*Cromileptes altivelis*) as a result of lower protein content and palatability of the feed probably had contributed to a higher value of HSI. However, they found that feeding fish with trash fish resulted in lower HSI compared to fish fed the pelleted feeds. In this trend, **Hernandez et al. (2007)** suggested that diets containing high amounts of carbohydrates should result in higher HSI index. On

the other hand, feeding rate and feed consume not only feed conversion efficiency but also condition factor and hepato-somatic index, which are used to assess the nutritional status of fish and are good predictors of physiological condition of aquatic animals (**Ng et al., 2000; Mihelakakis et al., 2002**). Though, a similar trend with hepato-somatic index in the present study (Table 6). **Hung et al. (1993)** in striped bass and **Mihelakakis et al. (2002)** in gilthead sea bream, observed similar differences in the condition in dices of fish fed suboptimal and adequate rations were reported when feeding rate was reduced from 2.0% to 0.5% body weight/day.

At the end of the 140 days of the growth trial, all groups of *D. labrax* showed mean individual gonad somatic index ranging from 0.281 to 0.27% for ovary, and 0.0597 to 0.0566% for testes, (Table 6) with no significantly differences among groups. Though, results presented in table (6) showed that gonado-somatic index (ovary and testes) for *D. labrax* fed dry pellet diet (treat.1) or fed only local trash fish (treat.6) alone was decreased insignificantly ($P > 0.05$) than fish fed combination of dry pellet diet and local trash fish (FS.4). A similar tendency was also observed by **Teruel et al. (2001)** who reported that a higher amount of essential nutrients in the artificial diets such as protein, lipid and the highly unsaturated fatty acids, for example, 20:4 n-6, 20: 5-n-3, 22: 6n-3 in *Haliotis asinina* fed artificial diet alone and a combination of natural diet and artificial diet influenced the increased reproductive performance (gonado- somatic index). In this connection **Kaushik et al. (2004)** found *Dicentrarchus labrax* fish meal replacement by plant ingredients (fish meal was decreased gradually from 100% to about 2% and replaced by plant protein sources were formulated have a slight increased in gonado-somatic index (GSI) 0.52, 0.72, 1.41. When final body weight 330.8; 333.2, 317.2g; with fish med was decreased gradually u to 25% (Fish meal 52, 40, 25) replaced by plant protein sources. **Cerdá et al. (1995)** with female bass (*D. labrax*) showed that fish fed with krill before or during the spawning season produced egg and larvae with a better quality than fish fed trash fish. This positive effect has been attributed to the presence of phosphatidy choline and astaxanthin from polar and nonpolar lipid fractions of raw krill, respectively (**Watanabe et al., 1991 a,b**).

Previous studies involving examination of female (*Dicentrarchus labrax*) reproductive performance, the wet diet proved to be the most beneficial (**Navas et al., 1998**) contrary to the results obtained by **Asturiano et al. (2001)** for sea bass (*D. labrax*) males. Since steroidogenesis and eicosanoids are essential for testicular function, it is possible that contrary to what has been reported in females, a reduction in these n-3 dietary PUFAs and an increase

in AA (arachidonic acid) resulting in an altered n-3: n-6 PUFA ratio may improve male European sea bass reproductive performance. Though, the same authors found that European sea bass fed commercial PUFA-enriched diets exhibited enhanced reproductive performance as compared to fish fed a wet diet. Studies have shown that male European sea bass fed diets with varying levels of n-3 and n-6 PUFAs exhibit depressed testicular steroidogenesis which may result in a delay in timing and extent of spermiation (Cerdá *et al.*, 1997). Moreover, *in vitro* studies (Wade *et al.*, 1994) showing the regulatory effects of AA, arachidonic acid, EPA, eicosapentaenoic acid, and DHA, docosahexaenoic acid, on testicular function are taken into account. The same authors shown that AA stimulates testosterone production, whereas n-3 PUFAs, particularly EPA, may function as inhibitory regulators. Recent *in vitro* studies in European sea bass testis demonstrated that AA-stimulated PG (prostaglandins) series E (PGE2) production was enhanced in the presence of gonadotropin, while EPA and DHA were ineffective (Asturiano *et al.*, 2000). It has been proposed that the inhibitory actions EPA and DHA may in part be due to inhibition of PGE2 formation from AA (Wade *et al.*, 1994). In addition, high cellular levels of EPA and DHA are known to displace AA and compete for cyclooxygenase enzymes which convert these PUFAs to eicosanoids which are less effective than PGE2 in reproductive events (Murdoch *et al.* 1993). In this regard, results by Asturiano *et al.* (2001) show that sea bass (*D. labrax*) fed diets lower in EPA and DHA composition and n-3:n-6 ratios had enhanced reproductive performance. Chaitanawisuti *et al.* (2011) attempted to condition broodstock spotted babylon (*Babylonia areolata*) using formulated diets than a local trash fish. They found that the levels of protein content of the formulated diet did not differ significantly among the local trash fish, but lipid content and total unsaturated fatty acids of the formulated diet was significantly higher than those of the local trash fish. On the other hand, they showed that the proximate composition of egg capsules (Protein and lipid) revealed that there were no significant differences fed both formulated diet and local trash fish but significant differences in fatty acid composition (higher levels of EPA, DHA and ARA) than those of broodstock fed the trash fish.

One of the most important factors necessary in the successful culturing of a fish species is obtaining a basic understanding of its key biological processes. The most important of these biological processes in the fish's reproductive cycle and formation of gametes. Small, round and transparent oocytes with a central nucleus were observed in histological sections of ovaries (plate 1: a, b, c, d, e, f)

and nucleoli were found in few oocytes. Also, no lipid droplets were found. These cells had basophilic cytoplasm and an acidophilic nucleus (plate 1. a, b, c, d, e, f). Though, in the present study, testes, based on microscopic observation were distinguished for male *Dicentrarchus labrax* Plate 2 (a, b, c, d, e, f) spermatogonia and primary spermatocytes were the dominant cells of this stage (plate 2). Though, spermatogonia had a light cytoplasm and a large nucleus. Some secondary spermatocytes having basophilic cytoplasm were also observed (plate 2). The results present in Table 6 and plate 1,2, show that fish fed mixed from formulated diet and local trash fish especially feeding schedule 4 have improving reproductive performance (gonado somatic index) and development tissue of gonads (ovary and testes) may be to different unsaturated acids content in both formulated diet and local trash fish. Chaitanawisuti, *et al.* (2011) found that the formulated diet contained significantly higher levels of ARA (arachidonic acid, 20:Ln-6), EPA (eicosapentaenoic acid, 20:5n-3) and DHA (docosahexaenoic acid, 22:6n-3) than those of fish fed the local trash fish. The same authors, showed the compositions of egg capsules produced significantly more ARA, EPA and DHA compared to broodstock fed the local trash fish, however, the ARA/EPA and DHA/EPA ratios in egg capsules were significantly higher in the trash fish – fed group compared to those fed the formulated diet. Lupatsch *et al.* (2001) reported that good knowledge on nutrient requirements is needed for best growth performance. Though, feed ration effects a wide range of biochemical parameter, in fish which in turn help to formulate its physiological response to environmental stimuli (Chatzifotis *et al.*, 2011). The same authors, showed that a case in point is the development and maturation of gonads which is achieved by the transfer of nutrients from the diet /or somatic tissue under the influence of daylight and water temperature. On the other hand, with sea bass (*Dicentrarchus labrax*), Papadaki *et al.* (2005) observed precocious males and females reach puberty after the first and second year of life (300- 400g), respectively, diverting a part of surplus somatic energy to reproduction instead of somatic growth. Cerda *et al.* (1994) reported that reduced feeding can delay the timing of spawning in sea bass and decrease fecundity, but with the concomitant reduction of somatic growth, however, gonadal maturation is accompanied by the transfer of lipids from energy increase of plasma lipids (Fernandez *et al.*, 1989). However, effects of starvation and re-feeding on reproductive indices long term starvation (one month feeding –three month starvation) and starvation re-feeding (two month starvation –two month re-feeding) on gonad maturation for *Dicentrarchus labrax* (12 months old,

weighing 289 ± 34 g were examined by **Chatzifotis et al (2011)**, who showed that the feeding regime had a profound effect on gonad maturation of sea bass, male and female fish in the long term starvation feeding

regime show significantly lower gonado somatic index values than those on the starvation / re-feeding, feeding regime.

Table (1): Nutrient composition (%) in dry matter of feedstuffs

Test ingredients	Dry matter	Crude protein	Crude lipid	Crude fiber	Nitrogen free extract	Ash
Fish meal	89.9	72.86	10.23	3.12	-	13.79
Meat and bone meal	92.3	50.70	7.91	3.90	16.58	20.91
Soybean meal	89.7	44.04	4.24	7.92	35.23	8.58
Corn gluten meal	88.2	56.24	5.90	4.76	26.08	7.03
Wheat flour	88.0	18.18	3.41	8.52	52.16	17.73

Table (2): Test diet formulations (pelleted diet)

Ingredient	Amount (g/100g diet as fed)
Fish meal	36.5
Meat and bone meal	20.7
Soybean meal	12.3
Corn gluten meal	8.0
Wheat flour	14.5
Fish oil	5.0
Vitamin*	1.5
Mineral**	1.5

* Vitamin premix (local) supplied the diet (g/kg) the following: Vit.A,9000000. I.U.;

D₃, 1500000 IU; E, 24g; C, 6g; K₃, 1-2 g; B₁, 0.9g; B₂, 1.5; B₆, 1.2g; Folic acid, 0.12g Niacin, 6g; Pantothenic acid, 2.76g. (Carrier, cellulose, up to 1000g).

** Mineral premix (local) consisted of (mg/kg premix) the following: 4000mg KCL (52%); 1030 mg ZnSO₄.7H₂O; 33mgKI; 1.35mg Na₂Se₂O₃; 1319mg MnSO₄.H₂O; 50mg copper sulphate (25% Cu); 5mg cobalt sulphate; 4300mg sodium sulphate (32.37% Na). (Carrier up to 1000gm).

Table (3): Proximate compositions of pelleted feeds and local trash fish

Parameters	Pelleted feed	Trash fish
Proximate composition (g/100g diet)		
Dry matter	90.77	25.46
Crude protein	49.29	61.63
Crude lipid	12.33	16.73
Crude fiber	7.78	-
Ash	17.14	19.40
Nitrogen-free extract	13.46	2.24
Gross energy (MJ/Kg ⁻¹)*	19.06	21.78
Protein energy ratio (mg protein/KJ)	25.86	28.30

Gross energy calculated using: 23.9 KJg⁻¹ protein; 39.8 KJg⁻¹ lipid; 17.6 KJg⁻¹ carbohydrates (cited by **Martinez-Lioren et al., 2007**).

D. labrax is already known to exhibit sexual growth dimorphism at commercial size (300 – 400 g). The females are larger than males at this stage of development with a relative advantage estimated at 20 - 40% (**Carrillo et al., 1995; Chatain et al., 1997**). **Saillant et al., (2003)** reported that high densities and size grading applied to intensive sea bass are not responsible for the sexual dimorphism in farmed populations. However, the feeding rates and growth conditions of fish may account for a part of

the sex ratio variation. Results obtained from this study show that there are histological features in the gonads of the males and females *D. labrax*. Sea bass culture, however, is dependent on the supply of trash fish because of the unfavourable response of the fish to artificial pelleted feed. **Suzuki, et al. (2000)** found that, the thickness of the Zona radiata is probably an adaptation to protect the developing egg from injury and from abrasion. The rhythm of deposition of yolk inclusions in the oocyte of fish differs from species to species. In *Oreochromis mossambics*, lipid vesicles and yolk granules appear in the oocyte at the same time (**Dadzie, 1974**). **Lasiak (1983)**, observed progressive thickening of the zone radiata with maturation in the mullet, *Liza richardsoni*, in connection with substratum spawning. In other teleosts, lipid vesicles are the first type of yolk inclusion to appear in vitellogenic oocytes, their appearance marking the onset of vitellogenesis (**Schackley & King 1977, Wiegand, 1982, Dadzie, et al., 2000**). **Dadzie et al., (2000)** did not observe cortical alveoli either, in the oocytes of *Pampus argenteus*. The authors, however, reported the presence of lipid-containing vacuoles in pre-vitellogenic oocytes. These vacuoles disappeared from the cells prior to the onset of vitellogenesis. It is possible, consequently, that in the present study, cortical alveoli were also transient as a distinctive phase and, therefore, not seen. **Abou-Seedo et al. (2003)**, through the analysis of the gonadosomatic indices of the fish, adduced evidence to the effect that although mature individuals occur in December, their numbers are too low to take into consideration, in terms of the management of the *Acanthopagrus latus* species in Kuwait waters, and that the greater proportion of individuals being to spawn in January. **Lasiak (1983)**, observed progressive thickening of the zona radiata with maturation in the mullet, *Liza richardsoni*, in connection with substratum spawning.

Table (4): Growth performance of *Dicentrarchus labrax* fed the dry pellet (DP) or local trash fish (TF) with different feeding schedules (FS). Values are means \pm standard error of two replicate of net pen culture.

Diets and Feeding schedule Growth performance	Dry Pellet diet	Feeding schedule 1	Feeding schedule 2	Feeding schedule 3	Feeding schedule 4	Local trash fish
Initial body weight (IBW) (g)	22.56 \pm 1.56	21.77 \pm 2.2	21.62 \pm 1.95	22.73 \pm 1.13	23.01 \pm 1.84	21.80 \pm 2.14
Final body weight (FBW) (g)	115.75 \pm 2.74 ^{ab}	115.90 \pm 3.07 ^a	113.71 \pm 2.87 ^b	112.13 \pm 3.11 ^{bc}	121.75 \pm 3.25 ^a	109.22 \pm 3.37 ^c
Gain (g)	93.19 \pm 2.38 ^{ab}	94.13 \pm 2.87 ^a	92.09 \pm 3.01 ^b	89.40 \pm 2.62 ^{bc}	98.74 \pm 3.11 ^a	87.42 \pm 3.25 ^c
Weight gain (%)	413.08 \pm 15.8 ^b	432.38 \pm 18.3 ^a	425.95 \pm 16.8 ^a	393.31 \pm 14.7 ^c	429.12 \pm 18.9 ^a	401.01 \pm 19.3
Specific growth rate (%/day)	1.168 \pm 0.07	1.194 \pm 0.05	1.186 \pm 0.06	1.140 \pm 0.08	1.190 \pm 0.05	1.151 \pm 0.07
Geometric mean (mean weight)	51.10 \pm 3.64 ^a	50.23 \pm 3.41 ^{ab}	49.58 \pm 2.29 ^b	50.48 \pm 3.12 ^a	52.93 \pm 4.31 ^a	48.80 \pm 3.56 ^b
Daily growth index (%)	1.46 \pm 0.08	1.49 \pm 0.07	1.47 \pm 0.05	1.42 \pm 0.007	1.51 \pm 0.05	1.42 \pm 0.08
Survival rate (%)	90.0 \pm 2.5	82.5 \pm 5.0	85.0 \pm 2.5	90.0 \pm 7.5	90.0 \pm 2.5	82.5 \pm 5.0

Mean With different superscript letters within rows are significantly differently (P < 0.05)

Table (5): Proximate analysis (%) of dorsal muscle in European sea bass on wet tissue fed the dry pellet (DP) or local trash fish (TF) with different feeding schedule (FS) (n=2, each n consist of measurement of 5 fish).

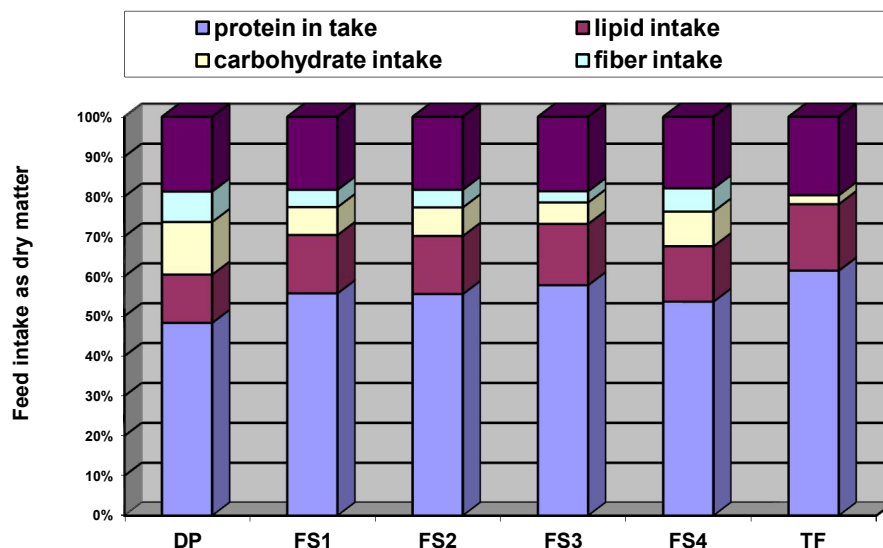
Proximate analysis (%) Diets and feeding schedule	Moisture	Crude protein	Crude fat	Ash	Gross energy (KJg ⁻¹)
Initial	76.27 \pm 2.35	18.34 \pm 0.94	2.16 \pm 0.18	3.21 \pm 0.22	524.3 \pm 23.1
Dry pellet diet (DP)	75.01 \pm 2.24	19.07 \pm 1.02	3.39 ^b \pm 0.21	2.63 \pm 0.20	590.7 \pm 19.7
Feeding schedule 1 (FS ₁)	74.36 \pm 1.96	19.11 \pm 0.87	3.85 ^a \pm 0.24	2.68 \pm 0.17	610.0 \pm 18.3
Feeding schedules 2 (FS ₂)	74.39 \pm 2.07	19.22 \pm 0.95	3.77 ^a \pm 0.19	2.62 \pm 0.21	60.4 \pm 25.01
Feeding schedule 3 (FS ₃)	74.95 \pm 2.15	19.19 \pm 1.10	3.45 ^{ab} \pm 0.22	2.40 \pm 0.19	596.0 \pm 2.23
Feeding schedule 4 (FS ₄)	74.40 \pm 1.88	19.25 \pm 0.98	3.70 ^a \pm 0.15	2.66 \pm 0.20	607.3 \pm 19.8
Local trash fish (TF)	75.58 \pm 2.17	19.10 \pm 1.21	3.18 ^b \pm 0.20	2.14 \pm 0.18	583.1 \pm 2.15

Values are mean \pm standard error values in the same row with different superscript letters are significantly different (P < 0.05); DM = dry matter

Table (6): Somatic parameters of European sea bass fed the dry pellet (DP) or local trash fish (TF) with different feeding schedule (FS). Values are mean \pm standard error of two replicates values in row with the same letter are not significant (P \leq 0.05)

Somatic parameter Diets and Feeding schedule	Condition factor (K)	Hepato-somatic index %	Gonado-somatic index	
			Ovary	Testes
Dry pellet diet (DP)	1.21 \pm 0.08	1.38 \pm 0.05 ^a	0.279 \pm 0.0012 (n = 20)	0.0584 \pm 0.0008 (n = 10)
Feeding schedule 1 (FS ₁)	1.23 \pm 0.10	1.33 \pm 0.07 ^{bc}	0.276 \pm 0.0064 (n = 24)	0.0585 \pm 0.0009 (n = 6)
Feeding schedule 2 (FS ₂)	1.22 \pm 0.11	1.35 \pm 0.04 ^{ab}	0.278 \pm 0.0004 (n = 21)	0.0579 \pm 0.0014 (n = 9)
Feeding schedule 3 (FS ₃)	1.19 \pm 0.07	1.34 \pm 0.07 ^b	0.271 \pm 0.002 (n = 23)	0.0574 \pm 0.0014 (n = 7)
Feeding schedule 4 (FS ₄)	1.25 \pm 0.08	1.41 \pm 0.08 ^a	0.281 \pm 0.0011 (n = 19)	0.0597 \pm 0.0013 (n = 11)
Local trash fish (TF)	1.20 \pm 0.09	1.30 \pm 0.06 ^c	0.275 \pm 0.003 (n=22)	0.0566 \pm 0.0016 (n = 8)

Data are presented as mean \pm S.E., n = 30 fish; N= Number of fish

**Fig. (1) Feed intake as dry matter for *Dicentrarchus labrax* as different feeding schedules**

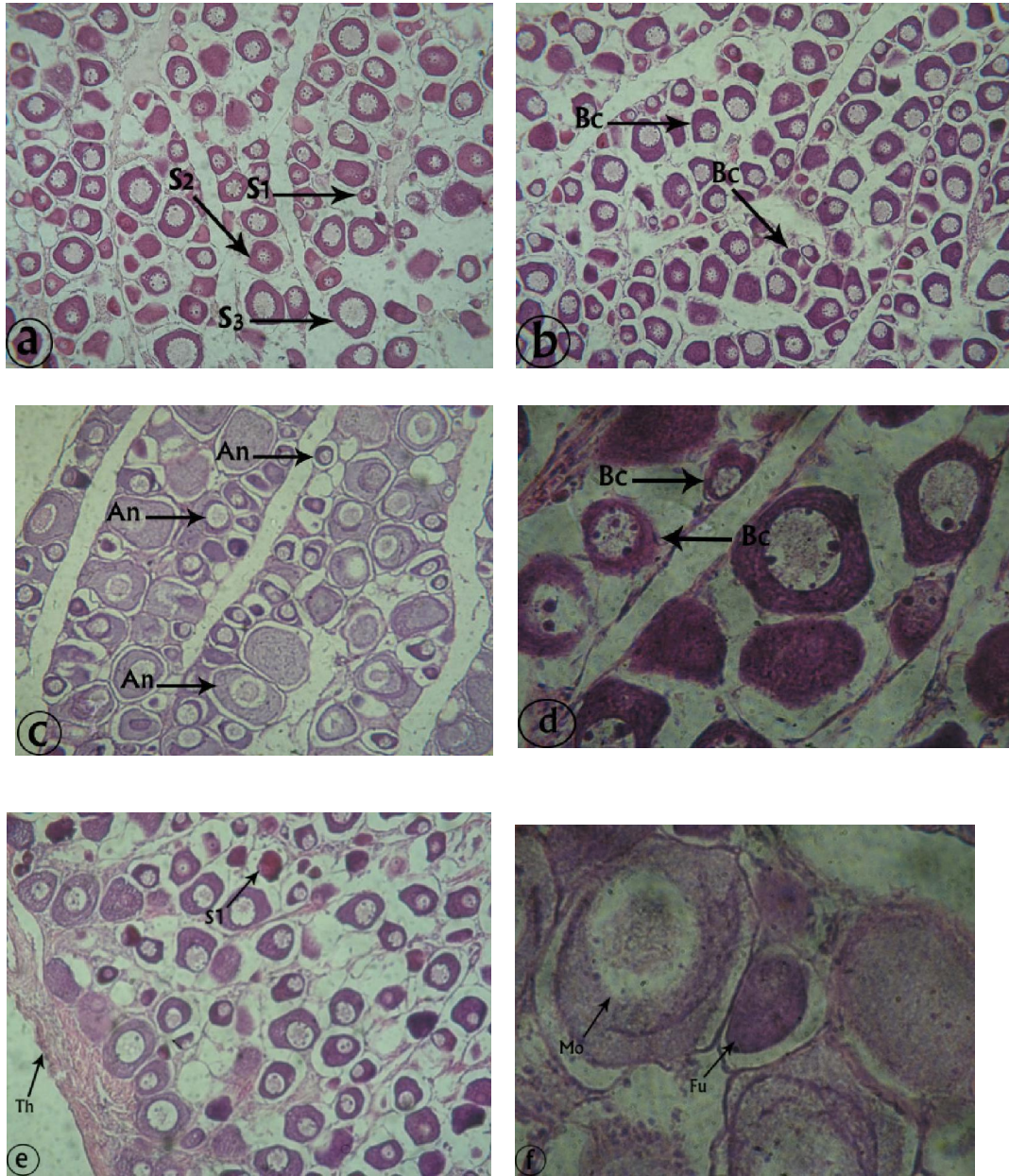


Plate (1).

Sections of *Dicentrarchus labrax* ovaries of fed different feeding schedule:

DP. a: Primary stage (S1), Secondary stage (S2) and Third stage (S3)

FS1.b: Basophilic cytoplasm(Bc) x100

FS2. c: Acidophilic nucleus (An) x100

FS3. d: Basophilic cytoplasm(Bc) x200

FS4. e: Theca layer (Th) and stage (S1) x100

TF.F: Mature ova (Mo) and fused two ova (Fu) x400

Small, round and transparent oocytes with a central nucleus were observed in histological sections of ovaries (plate-1) Nucleoli were found in few oocytes, No lipid were found, these cells had eosophilic cytoplasm and an acidophilic nucleus (plate -1,a,b,c,d,e,f).

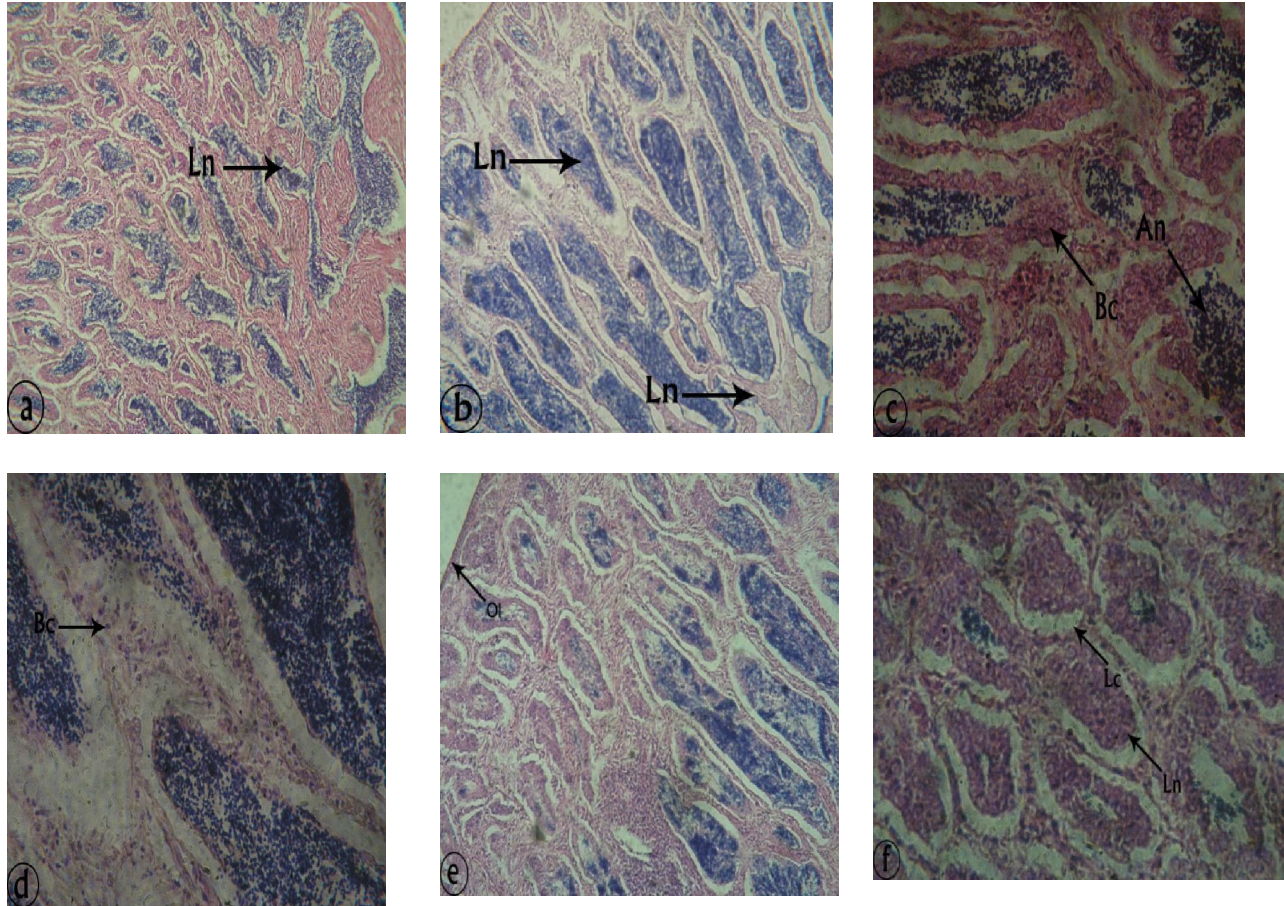


Plate (2).

Sections of *Dicentrarchus labrax* testes of fed different feeding schedule:

Dp. a: Large nucleus (Ln) x100

FS1. b: Large nucleus (Ln) x200

FS2. c: Basophilic cytoplasm (Bc) x200

FS3. d: Basophilic cytoplasm (Bc) x400

FS4. e: outer layer of testes (Ol) x200

TF. f: large nucleus (Ln) x200

Spermatogonia and primary spermatocytes were the dominant cells of this stage.

Thoughts spermatogonia had a light cytoplasm and a large nucleus, some secondary spermatocytes having basophilic cytoplasm were also observed

References

1. Abou-Seedo, F.S.; Dadzie, S, and Al-Kanaan, K.A. (2003). Sexuality, sex change and maturation patterns in the yellowfin seabream, *Acanthopagrus lotus*, (Teleostei: Sparidae) (Hostuyn, 1782) *Journal of Applied Ichthyology* 19:65-73.
2. Altan, O. and Yildirim Korkut, A. (2010). Effects of temperature and dietary carbohydrate level to growth performance and feed digestibility of European sea bass Juveniles (*Dicentrarchus Labrax*). *Bulgarian Journal of Agricultural Science*, 16 (No.6): 775-782.
3. Alvarez, M. J.; Lopez-Bote, C.J.; Diez, A.; Corraze, G.; Arzel, J.; Dias, J. and Kaushik, S. J. (1998). Dietary fish oil and digestible protein modify susceptibility to lipid peroxidation in the muscle of rainbow trout (*Oncorhynchus mykiss*) and sea bass (*Dicentrarchus labrax*). *British Journal of Nutrition*, 80: 281-289.
4. AOAC (Association of Official Analytical Chemists (2000). *Official methods of analysis*, 12th ed. Association of Official Analytical chemists, Washington, Dc. 1141 pp.
5. AOAC. (1997). *Animal feeds*. Chapter 4. in: Cunniff, P.A (Ed.). *Official Methods of Analysis*. Association of Official Chemists international. 1. AOAC. Arlington, VA. USA. P. 1102.
6. Asturiano, J. F.; Sorbera, L. A.; Carrillo, M.; Zanuy, S.; Ramos, J.; Navarro, J. C. and Bromage, N. (2001). Reproductive performance in male

- European sea bass (*Dicentrarchus labrax*, L.) fed two PUFA. Enriched experimental diets: A comparison with males fed a wet diet. *Aquaculture*, 194: 173-190.
7. Asturiano, J. F.; Sorbera, L. A.; Zanuy, S. Carrillo, M. (2000). Effects of polyunsaturated fatty acids and gonadatropin on prostaglandin series E Production in a Primary Testis cell culture System for the European sea bass. *J. Fish Biol. (In press)*.
 8. Asturiano, J. F.; Zanuy, S.; Ramos, J.; Bruce, M. Bromag, N. and Carrillo, M. (2006). Spawning performance and eggs and larvae Quality in European sea bass (*Dicentrarchus labrax*) fed with krill or Pufa enriched diets. *Journal of animal and Veterinary Advances* 5 (12): 1133-1142.
 9. Bernet, D.; Schmidt, H.; Meier, W.; Burkhardt-Holm, P. and Wahli, T. (1999): Histopathology in fish: Proposal for a protocol to assess aquatic pollution. *J. Fish Diseases*, 22: 25-34.
 10. Bunlipatanon, P.; Songseechan, N.; Kongkeo, H.; Abery, N. W. and De Silva, S. S. (2004). Comparative efficacy of trash fish versus compounded commercial feeds in cage aquaculture of Asian seabass (*Lates calcarifer*) (Bloch) and tiger grouper (*Epinephelus Fuscoguttatus*) (Forsskal). *Aquacult. Res.* 45:373-388.
 11. Carrilo, M.; Zanuy, S.; Prat, F.; Cerda, J.; Ramos, J.; Mananos, E. and Bromage, N. (1995). Sea bass (*Dicentrarchus labrax*) In: Bromage, N.; Roberts, R.J. (Eds.), *Broodstock Management and Egg and larval Quality*. Blackwell. Oxford, pp.138 – 168.
 12. Castell, J.D.; Bell, J.G. and Tocher, D.R. (1994). Effects of purified diets containing different combinations of arachidonic and docosahexaenoic acid on survival, growth and fatty acid composition of Juvenile turbot (*Scophthalmus – maximus*). *Aquaculture*, 128: 315-333.
 13. Chatain, B.; Peruzzi, S. and Saillant, E. (1997). Sex determination in *Dicentrarchus labrax*, no evidence for male or female heterogamety. In: J.F. Baroiller, D. Guerrier and Y. Guiguen, Editors, *Proceedings of the IVE Atelier Determnisme et Differentiation du Sexe*, Ststion Commune de recherché en Ichtyophysiologie Biodiversite Environnement, INRA, Rennes, France, p.18
 14. Cek, S. and Yilmaz, E. (2007). Gonad development and sex ratio of sharptooth catfish (*Clarias gariepinus* Burchell, 1822) cultured under laboratory conditions. *Turk. J. Zool*, 31: 35-46.
 15. Cerdá, J. Carrillo, M.; Zanuy, S. Ramos, J. and Serrano, R. (1995). Short- and long – term dietary effects on female sea bass (*Dicentrarchus labrax*): seasonal changes in plasma profiles of lipids and sex steroids in relation to reproduction. *Comp. Biochem. Physiol.*, 111 B: 83-91.
 16. Cerda, J.; Carrillo, M.; Zanuy, S.; Ramos, J. and de la Higuera, M. (1994). Influence of nutritional composition of diet on sea bass, *Dicentrarchus labrax* l. reproductive performance and egg and larval quality. *Aquaculture*, 128: 345-361.
 17. Cerda, J.; Zanuy, S. and Carrilo, M. (1997). Evidence for dietary effects on plasma levels of sexual steroids during spermatogenesis in the sea bass. *Aquacult. Int.* 5, 473-477.
 18. Chaitanawisuti, N.; Sangsawangchote, S. and Piyatiratitivorakul, S. (2011). Differences in fatty acid composition of egg capsules from broodstock spotted Babylon, *Babylonia areolata*, fed a local trash fish and formulated diet under hatchery conditions. *International journal of fisheries and Aquaculture*. Vol. 3(5). PP. 89-95.
 19. Chatzifotis, S.; Papadki, M.; Despoti, S.; Roufidou, C. and Antonopoulou, E. (2011). Effect of starvation and re-feeding on reproductive indices, body weight, plasma metabolites and oxidative enzymes of sea bass (*Dicentrarchus labrax*). *Aquaculture*, 316: 53-59.
 20. Company, R.; Galduch-Giner, J.; Perez-Sanchez, J. and Kaushik, S. (1999). Protein sparing effect of dietary lipids in common dentex (*Dentex dentex*): a comparative study with sea bream (*Sparus aurata*) and sea bass (*Dicentrarchus labrax*). *Aquat. Living Resour.* 12 (1): 23-30.
 21. Dadzi, S. (1974). Oogenesis and the stages of maturation in the female cichlid fish, *Tilapia mossambica*. *Ghana Journal of Science* 14: 23 -31.
 22. Dadzie, S.; Abou-Seedo, F. and Al-Shallal, T. (2000). Histological and histochemical study of oocyte development in the silver pomfret, *Pampus argenteus* (Euphrasen) in Kuwait wastars. *Arabian Gulf Journal of scientific Research* 18:23 – 31.
 23. Dias, J. (1999). Lipid deposition in rainbow trout (*Oncorhynchus mykiss*) and European sea bass (*Dicentrarchus labrax* L.): nutritional regulation of hepatic lip-genesis. Dr thesis. Univ. porto (Portual) and Univ. Bordeaux I (France). 190. PP.
 24. Dias, J.; Huelvan, C.; Dinis, M.T. and Metailler, R. (1998). Influence of dietary bulk agents (silica, cellulose and a natural Zeolite) on protein digestibility, growth, feed intake and feed transit time in European sea bass (*Dicentrarchus labrax*) juvenile. *Aquat. Living Resour.* J1 (4): 219-226.
 25. Duncan, D. B. (1955). Multiple range and multiple F. tests *Biometrics* 11, 1-42.
 26. Ekanem, S. B. (1996). Effects of feeding frequency, moist and dry feeds on the growth of *Chrysichthys nigrodigitatus* Lacepede and on pond water quality. *Aquacult. Res.* 27: 107-112.
 27. Enes, P.; Panserat, S.; Kaushik, S. and Oliva-Teles, A. (2006). Effect of normal and waxy maize starch on growth, food utilization and hepatic glucose metabolism in European sea bass (*Dicentrarchus Labrax*) Juveniles. *Comp. Biochem. Physiol. A* 143: 89-96.
 28. Fagbenro, O.A. (1994). Dried fermented fish silage in diets for *Oreochromis niloticus*. *Isr. J. Aquacult-Bamid* 46: 140-147.

29. Fernandez, F.; Miquel, A.G.; Guinea, J. and Martinez, R. (1998). Digestion and digestibility in gilthead sea bream (*Sparus aurata*): the effect of diet composition and ration size. *Aquaculture*, 166: 67-84.
30. Fernandez, J. Gutierrez, J.; carrillo, M.; Zanuy, S. and Planas, J. (1989). Annual cycle of plasma lipids in sea bass, *Dicentrarchus labrax l.*: effects of environmental conditions and reproductive cycle. *Comparative Biochemistry and Physiology part A. physiology* 93: 407-412.
31. Geay, F.; Santigosa I Culi, E.; Corporeau, C.; Boudry, P.; Dreano, Y.; Corcos, L.; Bodin, N.; Vandeputte, M.; Zambonino-Infante, J.L.; Mazurais, D. and Cahu, C.L. (2010). Regulation of FADS2 expression and activity in European sea bass (*Dicentrarchus labrax*, L.) fed a vegetable diet. *Comparative Biochemistry and physiology. Part B: Biochemistry and Molecular Biology*. Volume 156, Issue 4, pages 237-243.
32. Giri, S.S.; Sahoo, S.K.; Sahu, A.K. and Mukhopadhyay, P.K. (2000). Nutrient digestibility and intestinal enzyme activity of *Clarias batrachus* (Linn). Juveniles fed on dried fish and chicken viscera incorporated diets. *Bioresource Technology*, 71; 97-101.
33. Glencross, B.D.; Booth, M. and Allan, G. L. (2007). A feed is only as good as its ingredients – a review of ingredient evaluation strategies for aquaculture feeds. *Aquac. Nutr.* 13: 17-34.
34. Grigorakis, K.; Taylor, K.D.A. and Alexis, M.N. (2003). Organoleptic and volatile aroma compounds comparison of wild and cultured gilthead sea bream (*Sparus aurata*): sensory differences, and possible chemical basis. *Aquaculture*, 225: 109-119.
35. Guo, J. L.; Wang, Y. and Bureau, D. P. (2007). Inclusion of rendered animal ingredients as fish meal substitutes in practical diets for cuneate drum, *Nibea miichthioides* (Chu, Lo et Wu). *Aquac. Nutr.* 13: 81-87.
36. Hernandez, M. D.; Martinez, F. J.; Jover, M. and Garcia Garica. B. (2007). Effects of partial replacement of fish meal by soybean meal in sharpnose seabream (*Diplodus puntazza*) diet. *Aquaculture* 263: 159-167.
37. Hertrampf, J. W. and Piedad-Pascual, F. (2000). *Handbook on ingredients for Aquaculture feeds*. Kluwer academic publishers. Dordrecht. Netherlands. Pp. 482-483.
38. Hidalgo, F., Alliot, E. and Thebault, H. (1987). Influence of water temperature on food intake, food efficiency and gross composition of juvenile sea bass, *Dicentrarchus labrax*. *Aquaculture*, 64: 199-207.
39. Higgs, D. A.; Markert, J. R.; Plotnikoff, M. D.; McBride, J. R. and Dosanjh, B. S. (1985). Development of nutritional and environmental strategies for maximizing the growth and survival of juvenile pink salmon (*Oncorhynchus gorbuscha*). *Aquaculture*, 47: 113-130.
40. Hillestad, M. and Johnsen, F. (1994). High-energy/low-protein diets for Atlantic salmon: effects on growth, nutrient retention and slaughter quality. *Aquaculture*. 124: 109-116.
41. Hung, P.D. and Mao, N. D. (2010). Effect of different trash fish with alginate binding on growth and body composition of juvenile cobia (*Rachycentron canadum*). *Aquaculture Asia Magazine*, Volume XV No.2, April-June.
42. Hung, S.S.O.; Conte, F.S. and Hallen, E.K. (1993). Effects of feeding rates on growth, body composition and nutrient metabolism in striped bass (*Morone saxatilis*) fingerlings. *Aquaculture*, 112: 349-361.
43. Huy, N.Q. (2002). Status of cobia (*Rachycentron canadum*) reproduction and culturing in Vietnam. *Journal of Fisheries (In Vietnamese with English abstract)* 7, 14-15.
44. IBM SPSS Statistic. (2011). IBM SPSS Statistic Program. Version 19 Statistical software Packages. IBM Corporation. New York.
45. Kaushik, S. J. and Luquet, P. (1984). Relationship between protein intake and voluntary energy intake as affected by body weight with an estimation of maintenance needs in rainbow trout. *Z. Tierphysiol. Tierernahr. U. Futtermittelkd.*, 51: 57-69.
46. Kaushik, S. J.; Coves, D.; Dutto, G. and Blanc, D. (2004). Almost total replacement of fish meal by plant protein sources in the diet of a marine teleost, the European sea bass, (*Dicentrarchus labrax*). *Aquaculture* 230, 391-404.
47. Krogdahl, A.; Hemre, G. I. and Mommsen, T. P. (2005). Carbohydrates in fish nutrition: digestion and absorption in post larval stages. *Aquac. Nutr.*, 11: 103-122.
48. Lasiak, T.A. (1983). Aspects of the reproductive biology of the southern mullet. *Liza richardsoni*, from Algoa Bay, South Africa. *South African Journal of Zoology* 18: 89 -95.
49. Lee, S. M.; Cho, S. H. and Kim, D. J. (2000). Effects of feeding frequency and dietary energy level on growth and body composition of juvenile flounder *Paralichthys olivaceus* (Temminck and Schlegel). *Aquacult. Res.*, 31: 917-921.
50. Li, P.; Wang, X.X.; Hardy, R.W. and Gatlin 111, D.M. (2004). Nutritional value of fisheries by-catch and by-product meals in the diet of red drum (*Sciaenops ocellatus*). *Aquaculture*, 236: 485-496.
51. Lovell, R. (1979). Factors affecting voluntary food consumption by channel catfish. *Proc. World Symposium on Finfish Nutr. and finfish Tech. Hamburg 20-23 June 1978. Vol. 1. Berlin*.
52. Lovell, R.T. (1989). *Nutrition and Feeding of Fish*. Van Nostrand Reinhold, New York.
53. Lupatsch, I.; Kissil, G. W. and Sklan, D. (2001). Optimization of feeding regimes for European sea

- bass, *Dicentrarchus labrax*: a factorial approach. *Aquaculture*, 202: 289-302.
54. Maack, G. and Segner, H. (2003). Morphological development of the gonads in zebra fish. *Journal of fish Biology*, 62: 895-906.
 55. Mahmoud, H. H. (2009). Gonadal Maturation and histological observations of *Epinephelus areolatus* and *lethrinus nebulosus* in Halaieib/ Shalatién area "Red Sea", Egypt. *Global Veterinaria*, 3 (5): 414-423.
 56. Martinez-Ilorens, S.; Vidal, A. T.; Monino, A. V.; Torres, M. P. and Cerda, M. J. (2007). Effects of dietary soybean oil concentration on growth, nutrient utilization and muscle fatty acid composition of gilthead sea bream (*Sparus aurata* L.). *Aquaculture Research*, 38: 76-81.
 57. Mihelakakis, A.; Tsolkas, C. and Yoshimatsu, T. (2002). Optimization of feeding rate for hatchery-Produced Juvenile gilthead sea bream, *Sparus aurata*. *Journal of the world Aquaculture society*. Vol. 33, No. 2, June: 169-175.
 58. Millamena, O. M. (2002). Replacement of fish meal by animal by-product meal in a practical diet for grow – out culture of grouper, *Epinephelus coioides*. *Aquaculture* 204, 75-84.
 59. Munsiri, P. and Lovell, R. T. (1993). Comparison of satiate and restricted feeding of channel catfish with diets of varying protein quality in production ponds. *J. world Aquacult. Soc.* 24: 459-465.
 60. Murdoch, W. J.; Hansen, T. K. and Mc Pherson, L. A. (1993). A review-role of eicosanoids in vertebrate ovulation. *Prostaglandins* 46: 85-115.
 61. Navas, J. M.; Mananos, E.; Thrush, M.; Ramos, J.; Zanuy, S.; Carrillo, M.; Zohar, Y. and Bromage, N. (1998). Effect of dietary lipid composition on vitellogenin, 17a – estradiol and gonado tropin plasma levels and spawning performance in captive sea bass (*Dicentrarchus labrax*). *Aquaculture*, 165: 65-79.
 62. Ng, W. K.; Lu, K. S.; Hashim, R. and Ali, A. (2000). Effects of feeding rate on growth, feed utilization an body composition of a tropical catfish. *Aquac. Int.*, 8: 19-29.
 63. Nolan, M.; Jobling, S.; Brighty, G.; Sumpter, J. P. and Tyler, C. R. (2001). A histological description of inter sexuality in the roach. *Journal of fish Biology*, 58: 160-176. doi: 10.1006/J fbi. 2000.1431.
 64. Oberleas, D. and Harland, B. F. (1977). Nutritional agents which affect metabolic Zinc status. In: *Zinc Metabolism: Current Aspects in Health and Disease*. Alan R. Liss, Inc., New York, PP. 11-24.
 65. Papadki, M.; piferrer, F.; Zanuy, S.; Maingot, E.; Divanach, P. and Mylonaa, C.C. (2005). Growth, sex differentiation and gonad and plasma levels of sex steroids in male and female-dominant populations of *Dicentrarchus labrax* obtained through repeated size grading. *Journal of fish Biology* 66, 938-956.
 66. Parpoura, A.C.R. and Alexis, M.N. (2001). Effect of different dietary oils in sea bass (*Dicentrarchus Labrax*) nutrition. *Aquaculture international* 9: 463-476.
 67. Partridge, G. J. and Jenkins, G. I. (2002). The effect of salinity on growth and survival of juvenile black bream (*Acanthopagrus butcheri*). *Aquaculture*, 210: 219-230.
 68. Peres, H. and Oliva-Teles, A (1999). Influence of temperature on protein utilization in Juvenile European sea bass (*Dicentrarchus Labrax*). *Aquaculture* 170: 337-348.
 69. Perez, L.; Gonzalez, H.; Jover, M. and Fernandez-Carmona, J. (1997). Growth of European sea bass fingerlings (*Dicenturarchus labrax*) fed extruded diets containing varying levels of protein, lipid and carbohydrate. *Aquaculture*, 156: 183-193.
 70. Plascencia-Jatomea, M.; Olvera-Novoa, M.A. Arredondo-Figueroa, J.L.; Hall, G. M. and Shirai, K. (2002). Feasibility of fish meal replacement by shrimp head silage protein hydrolysate in Nile tilapia (*Oreochromis piloticus*) diets. *Journal of Science of Food Agriculture* 82: 753-759.
 71. Posadas, B. C. (1988). Economic analysis of various prawn farming systems. In: Chiu, Y. N.; Santos, L. M., Juliano, R. O. (Eds). *Technical Considerations for the Management and Operations of intensive prawn Farm*. U. P. aquaculture, Society. I Loilo. City, Philippines, PP. 12-24.
 72. Poston, H. A. (1986). Response of lake trout and rainbow trout to dietary cellulose, fish wild I. *Tech. Rep., Us Fish wild I. servv.* 5: 6 P.
 73. Robaina L.; Izquierdo, M.S.; Moyano, F.J.; Socorro, J.; Vergara, J. M.; Montero, D. and Fernandez-palacios, H. (1995). Soybean and lupin seed meals as protein sources in diets for gilthead seabream (*Sparus aurata*): nutritional and histological implications. *Aquaculture*, 130: 219-233.
 74. Russell, N. R.; Fish, J. D.; and wootton, K. J. (1996). Feeding and growth of juvenile sea bass: the effect of ration and temperature on growth rate and efficiency. *J. Fish Biol.* 49: 206-220.
 75. Saillant, E.; Fostier, A.; Haffray, P.; Menu, B.; Laureau, S.; Thimonier, J. and Chatain, B. (2003). Effects of rearing density, size grading and parental factors on sex ratios of the sea bass (*Dicentrarchus labrax*) in intensive aquaculture. *Aquaculture* 221: 183 – 206.
 76. Schackley, S.E. and King, P.E. (1977). Oogenesis in a marine teleosts, *Blennius pholis* L. *Cell Tissue Research*. 181: 105 – 128.
 77. Schwarz, F. J.; Plank, J. and Kirchgessner, M. (1985). Effects of protein energy restriction with subsequent realimentation on performance parameters of carp (*Cyprinus carpio* L.). *Aquaculture*. 48: 23-33.

78. Shapawi, R.; Mustafa, S. and Ng, W. K. (2011). A comparison of the growth performance and body composition of the humpback grouper, *Cromileptes altivelis* fed on farm-made feeds, Commercial feeds or trash fish. *Journal of fisheries and Aquatic Science*, 6 (5): 523-534.
79. Shiau, S.Y. and Liang, H. S. (1994). Nutrient digestibility and growth of hybrid tilapia, *Oreochromis niloticus* X *O. aureus*, as influenced by agar supplementation at two dietary protein levels. *Aquaculture*, 127: 41-48.
80. Stirling, H.P. (1977). Growth, food utilization and effect of social interaction in European sea bass (*Dicentrarchus labrax*). *Marine Biology*, 40, 173 – 184.
81. Stone, D. A. J. (2003). Dietary carbohydrate utilization by fish. *Reviews Fish. Sci.*, 11: 337-369.
82. Sugiyama, S.; Staples, D. and Funge-Smith, S. (2004). Status and potential of Fisheries and Aquaculture in Asia and the Pacific. Asia-Pacific Fishery Commission, RAP Publication 2004/25.
83. Suzuki, H.I.; Agostinho, A.A and Winemiller, K. O. (2000). Relationship between oocyte morphology and reproductive strategy in *Loricariid catfishes* of the Panama River, Brazil. *Journal of Fish Biology* 57: 791 – 807.
84. Tacon, A. G. J. (1995). The potential for fish meal substitution in aqua feeds. *INOFISH Int.* 3 (95), 29-34.
85. Teng, S. and Chua, T. (1978). Effect of stocking on the growth of estuary grouper, *Epinephelus Salmoides* Maxwell, cultured in floating net cages. USM/IFS/CTE. Project. Report #2. School of Biological Sciences. Universiti Sains Malaysia, Penang. Malaysia.
86. Teruel, MNB.; Millamena, OM. and Fermin AC. (2001). Reproductive performance of hatchery-bred donkey's ear abalone, *Haliotis asinine* Linne, fed natural and artificial diets. *Aquac. Res.*, 32: 249-254.
87. Tibaldi, E.; Hakim, Y.; Uni, Z.; Tulli, F.; DeFrancesco, M.; Luzzana, U. and Harpaz, S. (2006). Effects of the partial substitution of dietary fish meal by differently processed soybean meals on growth performance, nutrient digestibility and activity of intestinal brush border enzymes in the European sea bass (*Dicentrarchus Labrax*). *Aquaculture*, 261: 182-193.
88. Tubongbanua, E.S. (1987). Development of artificial feeds for sea bass (*Lates Calcarifer*). Management of wild and Cultured sea bass/ Barramundi: 186-188.
89. Van Ham, E. H., Berntssen, M. H. G.; Imsladn, A. K.; Parpoura, A. C.; Wenderlaar Bongar, S. E.; and Stefansson, S. O. (2003). The influence of temperature and ration on growth, feed conversion, body composition and nutrition retention of juvenile turbot (*Scophthalmus maximus*). *Aquaculture*, 217: 547-558.
90. Venkatesh, B.; Mukherji, A. p.; Mukhopadhyay, P.K. and Dehadrai, P.V. (1985). Growth and metabolism of the catfish, *Clarias batrachus* (Linn.) fed with different experimental diets. *Proc. Indian Academy of Sci.* 95, 457-462.
91. Wade, M. G.; Van Der Kraak, G.; Gerrits, M.F. and Ballatyne, J. S. (1994). Release and steroidogenic actions of polyunsaturated fatty acids in the goldfish testis. *Biol. Reprod.* 51: 131-139.
92. Watanabe, T.; Fujimura, T.; Lee, M.; Fukusho, K.; Satoh, S. and Takeuchi, T. (1991a). Effect of polar and nonpolar lipids from krill on quality of eggs of red sea bream, *Pagrus major*. *Nippon Suisan Gakkaishi*, 57: 695-698.
93. Watanabe, T.; Lee, M.; Mizutani, J.; Yamada, T.; Satoh, S.; Takeuchi, T.; Yoshida, N.; Kitada, T. and Arakawa, T. (1991b). Effective components in cuttlefish meal and raw krill for improvement of quality of red sea bream, *Pagrus major* eggs. *Nippon Suisan Gakkaishi*, 57: 681-694.
94. Weber, P. C. (1990). The modification of the arachidonic acid cascade by n-3 fatty acids. In: Samuelson, B., Dahlen, S.E.; Fritsch, J. and Hedqvist, P. (Eds.). *Advances in Prostaglandin, Thromboxane and Leukotriene Research*, Vol. 20. Raven Press, New York, Pp. 232-240.
95. Wiegand, M.D. (1982). Vitellogenesis in fishes. In: Richter. C.C.J. and Goos, J.J Th>, (Eds.). *Proceedings of international Symposium on Reproductive Physiology in fish*. pp: 136 – 146. Wageningen: Pudoc.
96. XU, Z.; Lin, X.; Lin, Q; Yang, Y. and Wang, Y. (2007). Nitrogen, Phosphorus, and energy waste outputs of four marine cage-cultured fish fed with trash fish. *Aquaculture*, 263: 130-141.
97. Xue, M. and Cui, Y. B. (2001). Effect of several feeding stimulants on diet preference by juvenile gibel carp (*Carassius auratus gibelio*), fed diets with or without partial replacement of fish meal by meat and bone meal. *Aquaculture*, 198: 281-292.
98. Yigit, M.; Koshio, S.; Aral, O.; Karaali, B. and karayucel, S. (2003). Ammonia nitrogen excretion rate- an index for Evaluating protein Quality of three feed fishes, for the black sea turbot. *The Israeli journal of Aquaculture-Bamidgeh*, 55 (1) 69-76.