### Heat Tolerance in Tomato

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Abstract: High temperature is the prevalent characteristic of subtropical and tropical regions and higher temperature has become an important limiting element for tomato production and yield. Although, tomato crops exhibit anatomical, morphological, physiological, phonological, and molecular responses to tackle with heat stress, but their reproductive stage and yields are extremely influenced by the high temperature. Various QTL's, heat shock proteins, and genes were detected in terms of heat resistance in tomato however a few stress-resistant tomato varieties are developed through traditional breeding ways. This is because the complicatedness of heat resistant characteristics that may be handled by the activity of different genes whose expression patterns are induced by several environmental elements. Furthermore, resistance to heat stress is developmentally regulated, stage-special event and resistance at one step of crop improvement is sometimes not related to resistance at other growth steps. Therefore, to produce of tomato under heat stress successfully, resistance may be required at whole imperative steps of crop growth such as germination of seed, reproductive and vegetative steps. Recently, various molecular and classical markers for heat resistance were screened and MAS (Marker-Assisted Selection) may be applied to improve tolerance of tomato to heat stress via biotechnological and molecular methods. To study heat tolerance in tomato appropriately, this paper will be an appropriate material and will assist for future studies.

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#### 1. Introduction

Tomato (Lycopersicon esculentum Mill.) is usually a fruit but it is sometimes called as a vegetable and vastly grown in the world and forms an imperative industry for agriculture. Throughout the world, after potato, it is mostly used as a second vegetable (FAOSTAT, 2005) and is definitely the most prominent garden plant. Tomatoes are consumed straightly as a raw vegetable or combined with other various food items such as tomatoes that are completely peeled, paste, diced products and diverse sorts of juice, soups and sauces (Foolad, 2007). In various parts of the world, tomatoes are an imperative section of a variant and balanced diet (Willcox et al., 2003). Tomato does not have a higher rate in nutrition value; one average tomato that is fresh (135gm) prepares vitamin C (47 percent RDA), vitamin A (22 percent RDA), and 25 calories energy. In the USA diet, for instance, tomato is the first among all vegetables and fruits as a rich origin of minerals and vitamins (Rick, 1980) and antioxidants that are phenolic (Vinson et al., 1998). Moreover, tomatoes which are fresh are the wealthy origins of the antioxidant lycopene (Nguyen and Schwartz, 1999) that preserves cells of plants from oxidants which have been related to cancer (Giovannucci, 1999).

Tomato crops are developed in vast sorts of environments with diverse climatic in the universe from the tropical areas to some degrees of the Arctic Circle. The biggest tomato producing nations involve China, USA, India, Turkey, Egypt, Italy, Spain, Brazil, Iran, Mexico, Canada, Greece and Russia (FAOSTAT, 2005). Although tomato has a good potential to be cultivated every location in the universe but it confronts lots of abiotic stress and high temperature is a crucial problem nowadays. According to the Intergovernmental Panel on Climatic Change (IPCC), in each decade, worldwide average temperature will be enhanced by 0.3 °C (Jones et al., 1999) and reached to around 1°C and 3°C higher than the current temperature by the years of 2025 and 2100, respectively and led to warming of the globe. AVDRC proposed that "in environments that are tropical, high temperature situations are sometimes common during the growing season and with climatic alteration, tomato plants in this region will be issued to enhanced temperature stress". Climatic analysis in areas in which tomato is grown proposes that temperatures are enhancing and the intensity and quantity of abovenormal temperature will rise in the next decades (Bell et al., 2000). In this condition, tomato production that is resistant to heat is extremely required.

Heat stress is identified as the enhancement in temperature below a threshold level for some time is enough to prompt irreversible harm to crop growth and improvement. As a whole, a temporary increase in temperature 10-15 C above normal, can lead to heat stress or shock (Wahid et al., 2007). Heat resistance refers to the capability of the crop to develop and create economic production in high temperatures. However, heat stress because of high temperatures is an important problem to plant yield throughout the world (Hall, 2001). Heat stress has been considered as one of the most imperative prompt of alteration in biochemical, morphology, and physiology facets of crops that decreases normal growth in diverse plants, involving tomato (Thomas and Prasad, 2003; Wahid et al., 2007). When temperature is up, injury of cellules and death may happen within minutes that could be related to a disturbance of cellular structure (Schoffl et al., 1999). When temperature is optimum, damages or death may happen after long term exposure. Direct damages can be happened in high temperatures such as denaturation and aggregation of protein, and enhanced membrane lipids liquidity. Indirect or slower heat damages can be occurred in terms of enzymes inactivation in chloroplast and mitochondria, limitation in of protein production, degradation of protein and loss of integrity of membrane (Howarth, 2005). Furthermore, in tomato, high temperatures can lead to remarkable losses in its yield because of the diminished fruit set, small size, and fruits low quality (Stevens and Rudich, 1978). Heat stress before anthesis period is linked with developmental alterations in the anthers, especially disorders in epidermis and endothecium, shortage in stromium opening and poor formation of pollen (Sato et al., 2002). Hazra et al. (2007) clarified that, in tomato, the signals which cause fruit set failure at high temperatures involves bud drop, abnormal flower growth, poor pollen creation, poor inflorescence and viability, abortion of ovule and reduced carbohydrate prohibition existence. Moreover, marked of photosynthesis happens at temperatures above average, causing remarkable decrease in yield. Intense heat stress (45°C, 20 min) in tomato that is mature-green leads to programmed cell death (PCD) in terms of fragmentation of DNA, cytochrome c release, and activity of special enzymes which are caspase-like (Ou et al., 2009). It is properly detected that reproductive organs of crops have higher susceptibility to heat stress in comparison with vegetative organs (Ruan et al., 2010; Zinn et al., 2010).

Crop species have different susceptibility in reaction to abiotic stress. Medium temperature for growth and reproduction is markedly different between crop species and their lines (Bohnert et al., 1995). However, in some occasions, tomato crops demonstrate special HSPs for presenting resistance to heat stress. The resistance is dedicated by HSPs which causes to promoted physiological parameters including photosynthesis, better use of water and nutrient, and integrity of membrane (Camejo et al., 2005; Ahn and Zimmerman, 2006; Momcilovic and Ristic, 2007). Such developments cause tomato growth to be feasible in heat stress situation. However, all of the lines within species have different abilities in tolerance to the heat stress but there are severe differences between and within species that provide chances to promote tomato tolerance to heat stress via genetic instruments. Vegetative and reproductive developments in tomatoes are intensively manipulated by temperature or various environmental elements (Abdalla and Verkerk, 1968).

Various endeavors to promote tomato heatresistant lines through traditional plant breeding methods have become prosperous (Ehlers and Hall, 1998; Camejo et al., 2005). But conventional breeding ways prepare low information on the locations of chromosomes that control complicated characteristics, the contemporary impacts of every chromosomal location on other characteristics (Epistasis, Pleiotropy or Linkage), or the genetic origin of such yield related characteristics due to dominance or over-dominance nature (Semel et al., 2006). If there is merely phenotype analysis, selection by conventional breeding instruments is hard when there are huge interactions between genotype and environment. There is no trustworthy field screening method that can be applied year by year or race by race (Kamel et al., 2010). Nevertheless, progressive methods of genetic engineering and molecular breeding have prepared further instruments that could be used to promote tomato with developed tolerance to heat. Molecular markers are used for both evaluating diversity of genes in germplasm collections and detecting varieties within population. Kantety et al. (1995) exhibited that ISSR method was capable to distinguish variations among inbred lines that are closely related and also individual population. Thin ISSR is very helpful to study genotypes of tomato. One technique to simplify polygenic characteristics' selection and breeding is to detect traits of interest through genetic markers. DNA markers have eased QTL (quantitative trait locus) mapping researches in populations which are segregated and exhibited certain genomic locations resulted from wild germplasm which have good potential to promote characteristics that are related to fruit (Gur and Zamir, 2004). Discovering of RAPD markers on tomato's genome map is advantageous to develop programs for breeding plants. It provides the easiest and most rapid technique for distinguishing a huge number of genome markers (Edwards et al., 1992). Michelmore et al. (1991) promoted the bulked segregant analysis of F<sub>2</sub> crops as an easier alternative method to analysis of isogenic line where the greatest and lowest extremes of the  $F_2$  population are bulked for the improvement of RAPD and SSR molecular markers required for QTL-assisted selection. ISSR markers have already found to be highly variable, require less time, money and labor than other ways and have the capability to be inherited (Wolfe and Liston, 1998). However, to make sure that this strategy is

successful, endeavors of crop physiologists, breeders and biologists are crucial (Wahid et al., 2007).

This review paper concentrates on responses of tomato to high temperature stress at the whole organs of crop, cellular and sub-cellular levels, mechanisms and methods of resistance for genetic development of tomato with resistance to heat stress that will be a substantial material for more studies.

#### 2. Threshold levels of heat-stress in tomato

A threshold temperature can be defined as value of daily average temperature in which a decrease in crop growth starts. Upper and lowermost developmental threshold temperatures have been detected for various crop genotypes in laboratory and field via controlled experiments. A less developmental threshold or an origin temperature is one below which growth of plant stops. At the same time, an overhead developmental threshold is the temperature above which growth ceases. Identifying a steady overhead threshold temperature is hard as the crop behavior may be different depending on other environmental situations (Miller et al., 2001). In tomato, for instance, when the environment temperature is higher than  $35^{\circ}$ C, its germination, seedling and vegetative stage, flowering and fruit set and ripening phase of fruit are inappropriately impacted (Miller et al., 2001). In general, basis and overhead threshold temperatures are varied in various crops which belong to diverse environments. However, Camejo et al. (2005) clarified that 30°C as overhead threshold temperature in emergence phase is harmful for tomato. Therefore, it is extremely favorable to estimate threshold temperatures for various steps of tomato crops to hinder harms by adverse temperatures in the crop ontogeny.

#### 3. Reactions of tomato to heat stress

# 3.1. Anatomical and morphological reactions of tomato

In tropic climates, extra radiation and great temperatures are sometimes the most prohibiting elements that affect plant development and final yield. Greater temperatures can lead to remarkable pre- and post-harvest harms, involving burning of twigs and leaves, leaves sunburns, stems and branches, senility of leaf and abscission, prohibition in the development of shoot and root, discoloration of fruit, and diminished production (Guilioni et al., 1997; Ismail and Hall, 1999; Vollenweider and Gunthardt-Goerg, 2005). Abdelmageed and Gruda (2009) perceived that morphological traits including fruits and flowers number per crop, percentage of fruit fresh weight and set were diverse in heat resistant and heat susceptible tomato lines and the outcomes were differed in field and glasshouse environments in 11 lines of tomato. There were vast levels of differentiations between the diverse varieties in their flowers number in glasshouse. 'CLN-1-0-3' created the greatest flowers number in each plant, but 'Omdurman' and 'UC-82-B' generated the lowermost numbers. Such impressiveness of high temperature is basically because of the reduction in bud or flower production and drop of flower. This outcome was similar to that of El-Ahmadi and Stevens (1979) where heat susceptible cultivar created only dropped flowers at high temperature. Fruits number in each plant was high in 'CLN-16-B'and 'CLN-1-0-3' but 'CLN-26-D', 'Summerset' and 'UC-82-B' had an average fruits number per crop, but the other lines had a few fruits in glasshouse. In contrast, under open field environments, the fruits number was 'zero' in the heat susceptible line 'UC-82-B' and the heat resistant line 'Summerset' vielded the biggest fruits number. Concerning percentage of fruit set; there were marked variations between the various lines. 'Summerset' presented the greatest percentage of fruit set, but 'UC-82-B' the heat susceptible variety had no fruits. Other lines generated low and 'CLN-1-0-3' was average. Percentage of fruit set exhibited a similar result as in the fruits number per crop. Satti and Abdalla (1984) and Dane et al. (1991) perceived the same outcomes in their own trials. In fruit fresh weight estimation, 'Summerset' demonstrated the greatest fruit weight and this is followed by 'Drd-85-F1', 'Omdurman', 'Kervic-F<sub>1</sub>' and 'Maverick-F<sub>1</sub>', while the other lines were either medium or low. This finding proves previous discoveries of El-Ahmadi and Stevens (1979). Sato et al. (2000) and Abdelmageed and Gruda (2009).

Under high temperatures, alterations in tomato anatomy were not explored in detail and a little information was accessible. In general, it is obvious that high temperature influences markedly plants anatomy at the tissue, cellular, and sub-cellular levels. The additional impacts of all these alterations in high temperature stress can lead to crop low growth and yield (Wahid et al., 2007). In all plant organs, there is a common trend of closure of stomata and loss of curtailed water, diminished size of cell, enhanced densities of stomata, and higher root and shoot's xylem vessels (Anon et al., 2004). The tomato flower happens in the three patterns that are organizational and flowers that are simple appear as well as branched and simple cymes. Flowers number that appears in inflorescence is based on environmental elements including temperature (David et al., 1996).

# **3.2.** Reactions of tomato in reproductive stage

Camejo et al. (2005) claimed that the medium temperatures for tomato plantation during the photoperiod are between 25 C and 30 C and during the dark period is 20 C. However, only 2-4 C rise in optimal temperature improperly influenced gamete development and prohibited the capability of pollinated flowers into seeded fruits and therefore diminished crops yields (Peet et al., 1997; Sato et al., 2001; Firon et al., 2006). Recently, Miller et al. (2001) clarified that heat stress higher than 35°C became a major blockade element for germination of seed, vegetative growth and seedling, flowering stage, fruit set and ripening in tomato. Peet et al. (1997) claimed that heat stress inappropriately impacts meiosis and germination in pollen, development of ovule and improvement and viability of embryo. Foolad (2005) also mentioned that meiosis in male and female organs, germination of pollen and development of pollen tube, viability of ovule, style and stigmatic situations, pollen grains number that are maintained by the stigma, fertilization and post-fertilization trends, endosperm development, pre-embryo and fertilized embryo influenced improperly by high temperature in tomato. Moreover, the most outstanding impact of high temperatures on reproductive stages in tomato is the production of an exserted style (i.e., stigma is elongated beyond the anther cone), that may hinder self-pollination. Critical period of susceptibility to optimize high temperature  $(32^{2}/26^{\circ}C)$  is 7 to 15 days before anthesis (Sato et al., 2002). High temperatures also participated in development of floral bud which caused to abortion of flower. Pollen grains numbers that created by the heat resistant varieties stayed more than susceptible lines (Abdelmageed et al., 2003).

Pollen viability and production are so susceptible to small rises in temperature higher than the medium (Thomas and Prasad, 2003). A decrease in generation of pollen, release, viability, germination capability, fruit set, and production in tomato at temperatures above medium levels has mentioned by various scientists (Peet et al., 1997; Sato et al., 2000; Pressman et al., 2002). Pollen viability germination and percentage capability decreased markedly in moderate and high temperature environments. It was mentioned that pollen grains which are germinated diminished 13 times when the temperature enhanced gradually from optimum (Pressman et al., 2002). Both the pollen germination and release capability in high temperature are imperative elements to identify the capability of fruit set. This is because a fail in pollen germination or release can hinder creation of fruit set even the pollen is viable (Sato et al., 2000). Pollination, growth of pollen tube and fertilization, and pollen germination must take place prosperously for good fruit set (Kinet and Peet, 1997). The fruit set decline under optimum high temperature stress is mainly because of a decrease in release and viability of pollen but not in generation of pollen (Sato et al., 2006) formerly, Sato et al. (2000) did not detect marked linkage among produced pollen grains number and fruit set. Eventually, they finalized that both pollen release and viability are the most imperative elements that effect fruit set in high temperature condition. Pressman et al. (2002) claimed that the impact of heat stress on viability of pollen was linked with

metabolism of carbohydrate during growth of anther. Under medium temperature, in pollen, concentration of soluble sugar enhanced slightly. Consistent high temperature hindered concentration of starch to be increased and caused soluble sugar content in mature pollen to be reduced. These probably lead to a reduction in livability of pollen. Poor fruit set has also been related to low amount of carbohydrates and growth regulators distributed in sink tissues of plant at high temperature (Kinet and Peet, 1997). Growth chamber and trainings of greenhouse propose that when flowers are first visible high temperature is most detrimental and susceptibility goes on for 10 to 15 days. Release of pollen and capability of germination can be a suitable standard for identifying crop reaction to high temperature and this is applied as a standard for selection in programs for breeding to choose heat resistant varieties (Comlekcioglu and Soylu, 2010).

# **3.3.** Phenological reactions of tomato

Heat stress can induce changes in crops directly such as existing physiological trends or indirect like changes of developmental patterns. These reactions may be different in one phenological step to one other (Weaich et al., 1996). Moreover, stress resistance is adjusted extensionally, stage-special event; resistance at one step of crop growth cannot be linked to resistance at other growth steps. For instance, in tomato, although crops are susceptible to high temperatures at entire ontogeny of crop, fruit set and flowering are highly susceptible steps; fruit set is slightly influenced by temperatures above 20/26 C day/night and is intensely impacted by above 26/35°C (Berry and Rafique-Uddin, 1988). Perception of alterations in phenology of crop in reaction to heat stress can disclose a proper understanding of the crop atmosphere interactions. and stress Diverse phonological steps are different in their susceptibility to high temperature, but this is based on species and varieties (Wollenweber et al., 2003; Howarth, 2005). In the growth step, when crop is under stress the intensity of feasible harms is experienced by plant. It is unknown whether damaging influences of heat episodes are cumulative that occur at various developmental steps (Wollenweber et al., 2003).

Alsadon et al. (2006) perceived remarkable variations in heat resistance in twenty tomato lines at diverse steps of growth. At the vegetative step, the remarkable greatest average values for EC (electrical conductivity) were detected in Edkawi variety (63.12 µmho/cm) and this is followed by Pakmore VF, Castle Rock, Chico, Pakmore and Tnshet Star, respectively. They also identified that these lines had higher susceptibility to heat stress at vegetative step, in contrast, the marked lowermost average value for EC was distinguished in Pearson, Super Strain-B, Queen, VFN-8 and Strain-B varieties, which indicate that

these lines had the best function and were resistant to heat stress at vegetative step. The other nine genotypes exhibited average mean values for EC were detected to be mildly resistant to heat stress. In the next step, heat susceptible varieties presented the greatest average values and heat resistant varieties had the lowermost average values. The rest varieties, that demonstrated average values for EC, were noticed to be moderately resistant to heat stress. In the fruiting step, those varieties that had the greatest EC average values were detected to be rather susceptible to heat stress. In contrast, those varieties that had the lowermost EC values were noticed as the best heat resistant varieties and the rest varieties, that showed medium values, were noticed to be moderately resistant to heat stress. The same findings were perceived by Saadella et al. (1990), Kuo et al. (1993) and Ismail and Hall (1999) in cowpea and wheat.

#### **3.4.** Physiological reactions of tomato

Physiologists and geneticists express that most stress resistant characteristics are complicated and handled by rather than one gene and impacted highly by different environments (Blum, 1988). In tomato crop water relations, concentration of compatible osmolytes, cell membrane thermo integrity, photosynthesis, and alterations in hormones are important physiological reactions to heat stress.

### 3.4.1. Waters relations

Crop water situation is an imperative variable under changing environmental temperatures (Mazorra et al., 2002). Heat stress, in tomato, disturbed hydraulic conductivity of root and the leaf water relationships (Morales et al., 2003). During daytime, increased transpiration affects water deficit in crops, prompting a reduction in potentiality of water and causing disturbance of lots of physiological parameters (Tsukaguchi et al., 2003). High temperatures can prompt crops to face more loss of water during daytime compared to nighttime (Wahid et al., 2007).

# **3.4.2.** Compatible osmolytes accumulation

An important adaptive system in lots of crops developed under abiotic stresses, involving salinity, lack of water and severe temperatures, is cumulating of organic compounds that have low molecular mass, commonly called compatible osmolytes (Hare et al., 1998; Sakamoto and Murata, 2002). Under stress, various crop species may cumulate different varieties of osmolytes including sugar alcohols (polyols), and sugars, quaternary and tertiary ammonium, proline, and tertiary sulphonium compounds (Sairam and Tyagi, 2004). In high temperature conditions, fruit set reduced in tomato crops because of the disturbance in metabolism of sugar and transport of proline during the narrow window of male reproductive growth (Sato et al., 2006). To summarize, due to remarkable functions of osmolytes in reaction to environmental stresses in

crops, (e.g., heat) resistance may be increased by enhanced cumulating of solutes that are compatible via conventional crop breeding, MAS (marker-assisted selection) or GE (genetic engineering) techniques (Ashraf and Foolad, 2007).

### 3.4.3. Photosynthesis

Changes in several photosynthetic approaches in heat stress are proper indexes of thermo resistance of the crop as they are correlated with growth. When photosynthesis is limited, crop development can be prohibited at high temperatures. Photochemical responses in thylakoid lamellae and metabolism of carbon in chloroplast stroma have been proposed as the primitive areas of damage at high temperatures (Wise et al., 2004). Rising leaf temperatures and density of photosynthetic photon flux affect thermo resistance adjustments of PSII, displaying their potentials to optimize photosynthesis in different environmental situations since the overhead thermal ranges do not exceed (Salvucci and Crafts-Brandner, 2004; Marchand et al., 2005). In tomato varieties that had different capacities to thermo resistance in enhanced chlorophyll a: b proportion and diminished chlorophyll, carotenoids proportion were perceived in the resistant varieties in high temperatures, showing that these alterations were linked to tomato's thermo resistance (Camejo et al., 2005; Wahid and Ghazanfar, 2006). Moreover, in high temperatures, reduction in chlorophyll a and b was rather proved in progressed in comparison with developing leaves (Karim et al., 1997, 1999). These impacts on photosynthetic machinery or chlorophyll were proposed to be related to the creation of active oxygen species (Camejo et al., 2006; Guo et al., 2006). PSII is intensely thermo labile and its function is highly diminished or relatively halted in high temperatures (Bukhov et al., 1999; Camejo et al., 2005) that may be because of the exclusivity of thylakoid membranes wherever PSII is situated (Mcdonald and Paulsen, 1997). Heat shock decreases the number of photosynthetic pigments (Todorov et al., 2003), rubisco binding proteins (RBP), soluble proteins, and large and small subunits (SS) of rubisco in darkness but enhances them in light, exhibiting their functions as HSPs and chaperones (Kepova et al., 2005). Photosynthesis is detected as a physiological index that is highly susceptible to high temperatures, and a rise in the content of atmospheric CO<sub>2</sub> will make temperature to be enhanced and this may present a remarkable effect on the yield and distribution of lots of crop genotypes in the future (Wahid et al., 2007).

# 3.4.4. Cell membrane thermo stability

Maintained role of cellular membranes under stress is fundamental for trends like respiration and photosynthesis (Blum, 1988). Heat stress fastens the kinetic energy and motion of molecules in membranes which lose chemical bonds in biological membranes molecules. This causes the biological membranes' lipid bilayer to be rather liquid by either proteins denaturation or a rise in fatty acids that are unsaturated (Savchenko et al., 2002). The stability and roles of biological membranes are susceptible to high temperature, as heat stress changes membrane proteins' tertiary and guaternary structures. These changes increase the penetrance of membranes, as obvious from enhanced loss of electrolytes. The enhanced solute leakage, as a symptom of diminished cell membrane thermo stability (CMT), has long been applied as an indirect estimation of heat-stress resistance in different crop species, involving potato and tomato (Chen et al., 1982), soybean (Martineau et al., 1979), cotton (Ashraf et al., 1994), cowpea (Ismail and Hall, 1999), wheat (Blum et al., 2001), sorghum (Marcum, 1998), and barley (Wahid and Shabbir, 2005).

# 3.4.5. Alterations in hormone

Crops have the capability to monitor and adjust to inappropriate environmental situations, although the adaptability or tolerance degree to special stresses differs between species and genotypes. Hormones have an imperative function in this issue. Under heat stress condition, hormonal homeostasis, biosynthesis content. stability. and compartmentalization are changed (Maestri et al., 2002). Stress hormones such as abscisic acid (ABA) and ethylene  $(C_2H_4)$ , are included in the regulation of various physiological properties by performing as signal molecules. Diverse environmental stresses, such as high temperature, leads to enhanced ABA levels (Larkindale and Huang, 2005). Other researches also propose that various HSPs (e.g., HSP70) induction by ABA can be one system whereby it confers thermo resistance (Pareek et al., 1998). Another kind of hormone, brassinosteroids have already been presented to confer thermo resistance to oilseed rape (Brassica napus) and tomato, but not to cereals (Dhaubhadel et al., 1999). The potential functions of other phytohormones in tomato for thermo resistance are unclear vet.

# **3.5. Molecular reactions of tomato**

Tomato exhibits molecular reactions to heat stress by creating heat shock proteins.

# 3.5.1. Heat shock proteins

Production and cumulating of special proteins are inquired when heat stress is rapid and these proteins are identified as HSPs. Enhanced production of HSPs happens when crops experience either sudden or slow rise in temperature (Nakamoto and Hiyama, 1999; Schoffl et al., 1999). HSPs induction seems to be a worldwide reaction to temperature stress, being perceived in various organisms from bacteria to human (Vierling, 1991). In semiarid and arid areas, crops can

produce and cumulate remarkable levels of HSPs. Under cyclic or developmental control, certain HSPs can also be expressed in various cells (Hopf et al., 1992). In this regard, HSPs expression is limited to certain steps of growth, including germination, embryogenesis, growth of pollen, and maturation of fruit (Prasinos et al., 2005). Three sorts of proteins, as detected by molecular weight, account for most HSPs, viz., HSP90, HSP70 and less molecular weight proteins of 15–30 kDa. The ratios of these protein sorts vary between crop species (Feussner et al., 1997). In reaction to high temperatures, special HSPs have been distinguished in various crop species. For instance, HSP68, that is located in mitochondria and usually expressed incorporately, was detected to have enhanced expression under heat stress in barley, tomato, maize, potato, and sovbean cells (Neumann et al., 1993). The gene for a nuclear-encoded HSP, Hsa32, that encode a 32 kDa protein, has been cloned in tomato (Liu et al., 2006). Immune-localization researches have identified that HSPs naturally link to specific cellular structures, like chloroplasts, cell wall, mitochondria, and ribosomes (Nieto-Sotelo et al., 2002; Yang et al., 2006). In tomato crops which suffer from heat stress. HSPs gather into a granular structure in the cytoplasm, probably preserving the protein bioproduction machinery (Miroshnichenko et al., 2005). Presence of HSPs can hinder other proteins denaturation that can be impacted by high temperature. The conformational dynamism and aggregate state of small HSPs may be vital for their roles in thermoprotection of crop cells from harmful influences of heat stress (Sch"offl et al., 1999; Iba, 2002). The specific significance of small HSPs in crops is proposed by their abnormal diversity and abundance. The capability of small HSPs to gather into heat shock granules (HSGs) and their decomposition is a prerequisite for crops cells survival under constant stress environments at sub-lethal temperatures (Miroshnichenko et al., 2005). LMW-HSPs may have structural functions in stability of cell membrane. LMW-HSPs localization in chloroplast membranes proposed that these proteins preserved the PSII from improper impacts of heat stress and played a function in transport of photosynthetic electron (Barua et al., 2003). Recently, in tomato, dual function of LMW HSP21 has been expressed as conserving PSII from oxidative harm and taking part in fruit color alteration during storage at low temperatures (Neta-Sharir et al., 2005).

# 4. QTLs for heat tolerant in tomato

In tomato, while substantial endeavors have been appropriated to the detection and mapping of QTLs conferring resistance to environmental stresses including drought, low temperatures, and salinity less mapping study has been performed on high temperatures (Foolad, 2005). Kadirvel (2010) showed an AVDRC report regarding two QTLs for heat resistant in tomato in chromosome 6 and 12. They exhibited that in Chromosome 6 the QTL is TES0111-SLM6-5; LOD score is 2.3; Variance10.6%; Additive influence 9.12; Donor CLN1621L and in chromosome 12 the QTL is SLM12-31-SLM12-50; LOD score is 2.6; Variance 13.0%; Additive influence 5.81; Donor CLN1621L. However, it seems that in tomato less improvement has been done in breeding and detection of QTLs for heat resistance than breeding for resistance to any other environmental stresses. This scenario proposed to a greater importance on breeding and detection of QTLs for heat resistance in tomato.

# 5. Molecular markers and classical genetic markers for heat tolerant tomato

To define, any characteristic which is expressed in multifold forms and inherited in a simple Mendelian fashion can be regarded and applied as a genetic marker. In tomato, there are more than 1300 morphological, physiological (e.g., male sterility, fruit abscission, fruit ripening), and disease tolerance genes (Kalloo, 1991) of them less than 400 have been mapped (Mutschler et al., 1987; Tanksley, 1993; Chetelat, 2002). The genetic markers' second generation, isozymes have been famous in 1970s and early 1980s. In tomato, 41 isozymic genes that correspond to 15 separate enzymatic responses have been detected, among them 36 have been mapped onto the 12 tomato chromosomes (Tanksley, 1993; Tanksley and Bernatzky, 1987). Despite their huge benefits, isozyme markers are so restricted in number and sometimes are not polymorphic between lines which are highly-related (Foolad et al., 1993; Tanksley and Orton, 1983).

With the arrival of DNA marker technique in 1980s (Botstein et al., 1980) and early 1990s, lots of restrictions linked to isozyme and morphological markers were conquered and genetic mapping entered to a new exciting and developed era with the promise to remarkably enhanced efficiency of crop breeding and genetics study. A DNA marker is usually branched from a small area of DNA that exhibits sequence polymorphism between individuals within or between species. DNA markers, that are phenotypically neutral and identically unrestricted in number, have permitted scanning of whole genome and assigning landmarks in high density on each chromosome in lots of crop species, involving tomato. During the past two decades, several sorts of molecular markers have been improved and progressed, like, but not restricted to, randomly amplified polymorphic DNAs (RAPDs) (Williams et al., 1990), simple sequence repeats (SSRs or microsatellites) (He et al., 2003; Tautz, 1989), amplified fragment length polymorphisms (AFLPs) (Vos et al., 1995), cleaved amplified polymorphic

sequences (CAPS) (Konieczny and Ausubel, 1993), restriction fragment length polymorphisms (RFLPs) (Botstein et al., 1980), variable number of tandem repeats (VNTRs or minisatellites) (Jeffreys et al., 1985), sequence characterized amplified regions (SCARs) (Paran and Michelmore, 1993), expressed sequence tags (ESTs) (Adams et al., 1991), conserved ortholog sets (COS) (Fulton et al., 2002), single-strand conformation polymorphisms (SSCPs) (Orita et al., 1989), insertion deletions (InDels), and single nucleotide polymorphisms (SNPs) (Landegren et al., 1998). Kamel et al. (2010) secluded DNA from the two contrasting parents, LSSS1 as a heat resistant parent and Super Strain B as a heat susceptible parent, their subsequent F1 and DNA bulks of the resistant and susceptible groups of F<sub>2</sub> segregating population were experimented against 10 preselected primers. All of the primers had polymorphisms with the genotypes studied. Primers A16 and Z13 presented 2 positive molecular markers that were only detected in the resistant parent (LSSS1),  $F_1$  and the resistant  $F_2$  bulk with 100 bp molecular sizes for primers A16 and 500 bp for primer Z13, while they were absent in the susceptible parent (Super Strain B) and the susceptible  $F_2$  bulk. In contrast, primers C02, C03, C05, C08, C14 and C15 demonstrated 8 molecular markers that were detected only in the susceptible F<sub>2</sub> bulk with 500 bp molecular size and 1500 bp for primer C02, 1750 bp and 750 bp for primer C03, 2400 bp for primer C05, 550 bp for primer C08, 400 bp for primer C14 and 650 bp for primer C15. Zhang et al. (1994) and Mackay and Caligari (2000) claimed that analysis of RAPD that is mixed with BSA has been applied to screen for markers associated with genes of interest. Furthermore, Lin et al. (2006) detected 14 random amplified polymorphic DNA (RAPD) markers associated with heat resistance characteristics in tomatoes under heat stress with the use of the bulked segregant analysis. Various RAPD markers were unique to one special characteristic, and the rest were related to two characteristics while several markers demonstrate one polymorphic band and the others two polymorphic band. They also made use of 22 genetic markers as indirect selection linked to morphological traits and exhibited polymorphic bands, 13 were special to the susceptible parent i.e. C09 marker presented 1.5 kb for high number of flower and 1.0 kb for low number of fruit; D06 marker's 0.3 kb for high number of fruit and 1.0 kb for low number of flower: D11 marker's 0.3 kb for high number of flower and 0.3 kb for high fruit weight; D12 marker's 1.0 kb for high number of flower; K06 marker's 1.1 kb for high number of flower and 1.3Kb for low number of fruit; K14 marker's 0.5 kb for high number of flower; P06 marker's 0.5 kb for high yield ; X01marker's 0.4 kb for high fruit weight and 0.7 kb for low number of flower while 9 were specific to the resistant parent, like D08 marker's 1.0 kb for low number of flower; K02 marker's 1.6 kb for low number of flower and 0.5 kb for low number of fruit; K08 marker's 0.6 kb for low number of flower; K20 marker's 0.9 kb for low fruit weight; P08 marker's 1.2 kb for low number of flower and 0.8Kb for low yield; S13 marker's 1.2 kb for low weight of fruit and 1.3 kb for low weight of fruit. Kamel et al. (2010) also detected that 844A as a primer presented as positive molecular marker that was only distinguished in the resistant parent (LSSS1),  $F_1$ and the resistant  $F_2$  bulk with 650 bp molecular sizes. These findings were similar to those of Lin et al. (2010) who made use of 160  $F_2$  tomato crops segregating population to detect ISSR markers that were related to fruit characteristics in the tomato which exposed to high temperatures. Lin et al. (2010) screened 100 ISSR-PCR primers in the parents and 51 were identified to be polymorphic and of them 42 markers were segregated in a Mendelian fashion. The greatest (14) and lowermost (3) band numbers were created by primers 884 and 814, respectively. Lin et al. (2010) created 127 AFLP bands with fragment sizes that ranged from 50 to 500 bp with the use of 2 ECoRIMseI primer pair combinations. Of these, 50 polymorphic bands with an average number of 25 bands per primer pair were disclosed. Among detected 50 polymorphic fragments, 26 AFLP loci were identified to be associated with the genetic map. Mansour et al. (2009) detected differentiations in tomato varieties that were grown under heat stress and distinguished only 15 ISSR (814, 844A, 844B, 17898A, 17898B, 17899A, 17899B, HB8-15) and 20 RAPD (P1-20) primers that could distinguish intraspecific differentiations.

# 6. Marker-assisted selection (MAS) for heat tolerance in tomato

Marker-Assisted Selection is defined as a selection for a characteristic that depend on the genotype of an associated marker more than the characteristic itself. In essence, a marker that is associated can be applied as a criterion for selecting indirectly. The potential of MAS as an instrument for plant progress has been vastly investigated (Tanksley et al., 1989; Ribaut et al., 2002; Servin et al., 2004). Despite the utility of MAS for manipulating singlegene characteristics is straightforward and has been properly documented, its usefulness for complicated characteristics has also been distinguished (Stuber and Edward, 1986; Edwards and Johnson, 1994; Eathington et al., 1997; Schneider et al., 1997; Knapp, 1998; Toojinda et al., 1998; Stuber et al., 1999; Zhu et al., 1999; Hospital et al., 2000; Bouchez et al., 2002; Tar'an et al., 2003; Zhou et al., 2003; Jiang et al., 2004). However, it should be understood that MAS for polygenic characteristic progress is in its primary step

and transitory process and the field is on the verge of producing convincing outcomes. Based on most simulation studies and empirical outcomes, it seems that characteristic heritability (h<sup>2</sup>) and the number-of-OTLs are the most imperative elements impacting the impressiveness of MAS. MAS seems to be most useful for characteristics with low  $h^2$  (0.1–0.3) and that are handled by rather small numbers of QTLs with huge impacts. In general, it is accepted that in most cases, for a trait that has a low-heritability, MAS will have better selection outcomes than selection of phenotypic (Stuber et al., 1999). Previous researches presented that heat resistance exhibits low heritability so that MAS can be applied for producing of heat resistant tomato. The stages are needed for the progress of markers to be used in MAS and various benefits of MAS are expressed in a review by Collard et al. (2005). Polymorphism level that is distinguished in molecular marker followed by the use of markerassisted selection (MAS) has been certified to be proper alternative way of the agronomic selection, where it provides crop breeders with environmentalindependent genetic markers for certain economic characteristics.

### 7. Development of heat-stress tolerance of tomato

Under agricultural systems, crops adaptation or their resistance to environmental stresses can be manipulated by different methods. Generally, the negative influences of abiotic stresses on agricultural yield are diminished by a composition of genetic development and cultural practices (Wahid et al., 2007). Genetic improvement involves progress of varieties that can resist to environmental stresses and generate economic yield. However, genetic progress of crops for stress resistance is an economically constant solution for generation of plants in stressful conditions (Blum, 1988). The relatives of the planted tomato have certified to be valuable origines of favorable genes for better genetic development (Rick, 1986) and prosperous inter-generic crosses have also been made among planted tomato and its nearly related Solanum species (Rick, 1960; Stoeva and zagorska, 1987; Wann and Johnson, 1963). Hybrid lines also seemed to have a proper performance consistency especially under stress than optimal growing environments (Yordanov, 1983). Both traditional and hybrid breeding ways, that benefit of additively acting genes and genetic interactions, should be useful in tomato heat resistance breeding. In favor of hybrid breeding, around 1/3 of the diallel hybrid progenies from the foregoing study had better fruit set than the better heat resistant parents (AVDRC, 1988). In another related study, crosses among heat resistant stocks were better in fruit setting capability and yield than their crosses with heat susceptible parents from the diallel test (Opena et al., 1987).

Adjustment or alterations in cultural practices, like the time of planting, crop density, and management of soil and irrigation can reduce stress impacts, for instance Hanna et al. (1997) identified development and yield reactions of heat resistant tomatoes to depth of transplant, daily irrigation time and color of polyethylene mulch. They cultivated fiveweek-old tomato seedlings to a depth of 15.0 cm and perceived remarkable rise in marketable yield but mean fruit mass was not affected by transplant depth, in contrast, crop dry mass was markedly enhanced by deeper transplanting. Irrigation in morning enhanced the marketable and total yields, average fruit mass in 1994, and dry mass of crop in 1995. White-surface mulch had the same impact on fruit mass and yield. They finalized that a rise in yield of heat resistant tomatoes can be performed by deeper transplanting, irrigation in morning, making use of white-surface polyethylene mulch, or a mixture of all three. Practically, to be prosperous in developing agricultural yield in stress conditions, both genetic progress and adjustment in cultural practices must be done simultaneously (Wahid et al., 2007). In below, a summary of such endeavors and improvements is discussed and demonstrated.

### 7.1. Traditional breeding strategies

Traditional breeding of heat resistant crops basically based on selection and a common technique of selecting crops for heat stress resistance has been to grow breeding materials in a hot target production environment and detect individuals/lines with higher yield (Ehlers and Hall, 1998). A proposed method has been detected in selection criteria during early steps of crop growth that can be linked to heat resistance during reproductive steps. In tomato, a potent positive correlation has been perceived between vield and fruit set under high temperature. Therefore, estimation of germplasm to detect sources of heat resistance has regularly been performed by screening for fruit set under high temperature (Berry and Rafique-Uddin, 1988). Among various other characteristics that are influenced by high temperature, the non-reproductive trends involve efficiency of photosynthesis, assimilate translocation, mesophyll tolerance, and cellular membranes disorganization (Chen et al., 1982). Breeding to develop such characteristics under high temperatures can lead to improvement of varieties with heat resistance approaches. Various other concerns when applying conventional breeding protocols to promote heat resistant crops are as follows:

-Detection of genetic resources with heat resistance approaches. In lots of crop species, for instance tomatoes and soybeans, restricted genetic differentiations exist within the cultivated species necessitating detection and use of wild accessions (Foolad, 2005). -In different crop species, heat resistance is sometimes linked to various unfavorable agronomical or horticultural traits. In tomato, for instance, two unfavorable traits generally perceived in heat resistant lines are small fruit and limited foliar canopy (Scott et al., 1997).

-The small fruit production is mostly because of improper impacts of high temperature on the creation of auxins in the fruit and the poor canopy is for the sake of the highly reproductive nature of the heat resistant varieties (Scott et al., 1997).

Heat resistance breeding is yet in its primitive step and needs more attention in comparison with the past. Unfortunately, the literature has partially less information on breeding for heat resistance in various species. However. although plant all the complicatedness of heat resistance and hardships confronted during transfer of resistance, various heat resistant inbred lines and hybrid varieties with commercial acceptability have been improved and released in tomato (Scott et al., 1986; Scott et al., 1995).

# 7.2. Molecular and biotechnological strategies

Recent genetic researches and endeavors to convince high-temperature resistance of crops with the use of conventional protocols and transgenic attributes have vastly detected that crop heat stress resistance is a polygenic characteristic. Various ingredients of resistance, handled by various sets of genes, are vital for heat resistance at various steps of crop growth or in diverse tissues (Howarth, 2005; Bohnert et al., 2006). Therefore, the use of genetic stocks with diverse levels of heat resistance, co-segregation and correlation analyses, molecular biology methods and molecular markers to detect resistance, QTLs are promising attributes to dissect the genetic source of thermo-(Maestri et al., 2002). Recently, resistance biotechnology has assisted substantially to a proper understanding of the genetic source of heat resistance. For instance, various genes which are responsible for inducing the HSPs synthesis, have been detected and secluded in diverse crop species, involving maize and tomato (Liu et al., 2006; Sun et al., 2006; Momcilovic and Ristic, 2007). It has also been exhibited that tomato MT-sHSP has a molecular chaperone role in vitro (Liu and Shono, 1999) and recently it has been presented that MT-sHSP gene shows thermo-resistance in transformed tobacco with the tomato MT-sHSP gene (Sanmiya et al., 2004) at the crop level. Experimental data gained from transgenic, reverse-genetics and mutation attributes in non-cereal species prove causal involvement of HSPs in thermo-resistance in crops (Queitsch et al., 2000).

# 7.3. Induction of heat resistance of Tomato

Though genetic methods may be advantageous in the production of heat resistant crops,

it is probable that the recently produced crops are yielded low in comparison with near-isogenic heat susceptible crops. Therefore, substantial attention has been devoted to the induction of heat resistance in existing high-vielding varieties. Among the various techniques to achieve this target, foliar application of, or pre-sowing seed treatment with, low concentrations of inorganic salts, osmoprotectants, signaling molecules (e.g., growth hormones) and oxidants (e.g.,  $H_2O_2$ ) as well as preconditioning of crops are common attributes. Preconditioned tomato crops presented better osmotic adjustment by keeping the osmotic potential and stomatal conductance and better development than non-conditioned crops (Morales et al., 2003). Similarly, heat acclimated, in comparison with non-acclimated, turf grass leaves revealed higher thermo-stability, lower lipid peroxidation product malondialdehyde (MDA) and lower harm to chloroplast in exposure to heat stress (Xu et al., 2006). In tomato, it was exhibited that heat treatment administered to crops prior to chilling stress resulted in diminished incidence and intensity of chilling injury in fruit and other organs (Whitaker, 1994). Therefore, to promote heat resistant tomato crop, alternative methods to genetic means would involve pre-treatment of crops or seeds with heat stress or certain mineral or organic compounds. The success of such method, however, based on tomato plant and genotypes and must be studied on case basis.

#### 8. Conclusions and future prospects

Already substantial improvement has been performed in tomato research, involving development of molecular markers, mapping of specific genes and QTLs, comparative analysis of different characters, fine-mapping and map-based cloning of genes and genome sequencing and organization. Molecular mapping can be applied as criteria for indirect selection and tomato improvement. However, little information is available for the use of markers in tomato breeding especially for the development of complex characteristics like heat resistance. However, depending on the most recent discoveries and research progresses, it is clear that the future of routine application of markers in heat resistant tomato breeding is prospective. But the most imperative problem is the improvement of appropriate markers for the breeding programs. PCR based molecular markers that can distinguish polymorphism between closely related genotypes can be used in marker-assisted breeding for heat resistant tomato. Furthermore, the complete sequencing of the tomato genome will assist to progress sequence-based high-resolving markers. This will make MAS as a routine procedure in tomato breeding programs especially for improvement of many complicated characteristics. For complex traits i.e. heat resistance obtaining a reliable phenotypic data for QTL mapping may not be proper on the other hand, partitioning of the total genetic variation for heat resistant characteristic into its physiological and developmental components would lead to detection of QTLs for individual components that may be more useful. The importance of such progresses is well distinguished by the geneticists and plant breeders and lots of research programs have commenced such activities. As heat resistant tomato is a demandable criterion in tropical and subtropical environment in future, a combination of traditional breeding protocols and marker assisted breeding will become a routine procedure for heat resistant tomato production.

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### References

- 1. FAOSTAT. F.A.O. Statistical Databases. Food and Agriculture Organization of the United Nations, Statistics Division, Rome, Italy, 2005.
- 2. Foolad MR. Genome mapping and molecular breeding of tomato. Int J Plant Genomics 2007: 64358
- 3. Willcox JK, Catignani GL, Lazarus S. "Tomatoes and cardiovascular health," Critical Reviews in Food Science and Nutrition 2003;43(1):1–18.
- 4. Rick CM. "Tomato," In: Hybridization of Crop Plants, American Society of Agronomy/Crop Science Society of America, Madison,Wis, USA, 1980;669-680.
- 5. Vinson JA, Hao Y, Su X, Zubik L. "Phenol antioxidant quantity and quality in foods: vegetables." J Agr Food Chem 1998;46(9):3630–3634.
- Nguyen ML, Schwartz SJ. "Lycopene: chemical chemical and biological properties." Food Technol 1999;53(2):38–45.
- Giovannucci E. "Tomatoes, tomato-based products, lycopene and cancer: review of the epidemiologic literature." J Natl Cancer Inst 1999;91(4):317–331.
- Jones PD, New M, Parker DE, Mortin S, Rigor IG. Surface area temperature and its change over the past 150 years. Rev Geophys 1999;37:173–199.
- Bell GD, Halpert MS, Schnell RC, Higgins RW, Lowrimore J, Kousky VE, Tinker R, Thiaw W, Chelliah M, Artusa A. Climate Assessment for 1999. Supplement June 2000, Bull Am Meteorol Soc 2000;81.
- Wahid A, Gelani S, Ashraf M, Foolad MR. Heat tolerance in plants: An overview. Environ Exp Bot 2007;61:199–223.
- 11. Hall AE. Crop Responses to Environment. CRC Press LLC, Boca Raton, Florida, 2001.
- 12. Thomas JMG, Prasad PVV. Plants and the Environment /Global Warming Effects. University of Florida, Gainesville, FL, USA, 2003.

- Schoffl F, Prandl R, Reindl A. Molecular responses to heat stress. In: Shinozaki K, Yamaguchi-Shinozaki K, (eds). Molecular Responses to Cold, Drought, Heat and Salt Stress in Higher Plants. R.G. Landes Co., Austin, Texas, 1999;81–98.
- Howarth CJ. Genetic improvements of tolerance to high temperature. In: Ashraf M, Harris PJC, (eds). Abiotic Stresses: Plant Resistance Through Breeding and Molecular Approaches. Howarth Press Inc., New York, 2005.
- 15. Stevens MA, Rudich J. "Genetic potential for overcoming physiological limitations on adaptability, yield and quality in tomato." Hort Science 1978;13(6):673–678.
- Sato S, Peet MM, Thomas JF. Determining critical preand post- anthesis periods and physiological processes in *Lycopersicon esculentum* Mill. exposed to moderately elevated temperatures. J Exp Bot 2002;53:1187-1195.
- 17. Hazra P, Samsul HA, Sikder D, Peter KV. Breeding tomato (*Lycopersicon esculentum* Mill.) resistant to high temperature stress. Int J Plant Breed 2007;1:1.
- Qu GQ, Liu X, Zhang YL, Yao D, Ma QM, Yang MY, Zhu WH, Yu S, Luo YB. Evidence for programmed cell death and activation of specific caspase-like enzymes in the tomato fruit heat stress response. Planta 2009;229:1269–1279.
- 19. Ruan YL, Jin Y, Yang YJ, Li GJ, Boyer JS. Sugar input, metabolism and signaling mediated by invertase: roles in development, yield potential, and response to drought and heat. Mol Plant 2010;3:942–955.
- 20. Zinn KE, Tunc-Ozdemir M, Harper JF. Temperature stress and plant sexual reproduction: uncovering the weakest links. J Exp Bot, 2010;61:1959–1968.
- 21. Bohnert HJ, Nelson DE, Jensenay RG. Adaptations to Environmental Stresses. The Plant Cell 1995;7:1099– 1111.
- Camejo D, Rodriguez P, Morales AM, Amico JM, Torrecillas A, Alarcon JJ. High temperature effects on photosynthetic activity of two tomato cultivars with different heat susceptibility. J Plant Physiol 2005;162:281–289.
- Ahn YJ, Zimmerman JL. Introduction of the carrot HSP17.7 into potato (*Solanum tuberosum* L.) enhances cellular membrane stability and tuberization *in vitro*. Plant Cell Environ 2006;29:95–104.
- 24. Momcilovic I, Ristic Z. Expression of chloroplast protein synthesis elongation factor, EF-Tu, in two lines of maize with contrasting tolerance to heat stress during early stages of plant development. J Plant Physiol 2007;164:90–99.
- 25. Abdalla AA, Verkerk K. Growth, flowering and fruitset of the tomato at high temperature. Neth J Agr Sci 1968;16:71–76.
- 26. Ehlers JD, Hall AE. Heat tolerance of contrasting cowpea lines in short and long days. Field Crops Res 1998;55:11–21.
- Semel Y, Nissenbaum J, Menda N, Zinder M, Krieger U, Issman N. Overdominant quantitative trait loci for yield and fitness in tomato. Proc Natl Acad Sci 2006;103:12981–12986.

- Kamel MA, Soliman SS, Mandour AE, Mahassen SSA. Genetic Evaluation and Molecular Markers for Heat Tolerance in Tomato (*Lycopersicon esculentum* Mill.). J Am Sci 2010;6(12):364-374.
- Kantety RV, Zeng X, Bennetzen JL, Zehr BE. Assessment of genetic diversity in dent and popcorn (*Zea mays* L.) inbred lines using inter-simple sequence repeat (ISSR) amplification. Mol Breed 1995;1:365-373.
- Gur A, Zamir D. Unused natural variation can lift yield barriers in plant breeding. PLoS Biol 2004;2:1610–1615.
- Edwards MD, Helntijaris TT, Wright S, Stuber CW. Molecular marker facilitated investigation of quantitative trait loci in maize. 4. analysis based on genome saturation with isozyme and restriction fragment length polymorphism markers. Theor Appl Genet 1992;83:765–774.
- 32. Michelmore RW, Paran I, Kessel RV. Identification of markers linked to disease resistance genes by bulked segregant analysis: a rapid method to detect markers in specific genomic regions by using segregating populations. Proc. Natl Acad Sci 1991;88:9829-9832.
- Wolfe AD, Liston A. RAPD markers. In: Soltis DE, Solties PS, Doyle JJ, (eds). Molecular Systematics of Plants II: DNA sequencing. Boston, 1998;43-86.
- Miller P, Lanier W, Brandt S. Using Growing Degree Days to Predict Plant Stages. Ag/Extension Communications Coordinator, Communications Services, Montana State University-Bozeman, Bozeman, MO, 2001.
- 35. Guilioni L, Wery J, Tardieu F. Heat stress-induced abortion of buds and flowers in pea: is sensitivity linked to organ age or to relations between reproductive organs. Ann Bot 1997;80:159–168.
- Ismail AM, Hall AE. Reproductive-stage heat tolerance, leaf membrane thermo-stability and plant morphology in cowpea. Crop Sci 1999;39:1762–1768.
- Vollenweider P, Gunthardt-Goerg MS. Diagnosis of abiotic and biotic stress factors using the visible symptoms in foliage. Environ Pollut 2005;137:455– 465.
- Abdelmageed AHA, Gruda N. Growth and gas exchange rate of some tomato cultivars under controlled heat stress conditions. Eur J Hortic Sci 2009;74(4):152–159.
- El-Ahmadi AB, Stevens MA. Reproductive responses of heat-tolerant tomatoes to high temperatures. J Am Soc Hort Sci 1979;104(5):686–691.
- 40. Satti SME, Abdalla AA. Temperature and nitrogen fertilization in relation to flowering and fruit set in tomatoes under arid conditions of the Sudan. Acta Hort 1984;143:289–297.
- 41. Dane F, Hunter AG, Chambliss OL. Fruit set, pollen fertility and combining ability of selected tomato genotypes under temperature field conditions. J Am Soc Hort Sci 1991;116(5):906–910.
- 42. Sato S, Peet MM, Thomas JF. Physiological factors limit fruit set of tomato (*Lycopersicon esculentum* Mill.) under chronic mild heat stress. Plant, Cell and Environment 2000;23:719–726.

- 43. Anon S, Fernandez JA, Franco JA, Torrecillas A, Alarcon JJ, Sanchez-Blanco MJ. Effects of water stress and night temperature preconditioning on water relations and morphological and anatomical changes of *Lotus creticus* plants. Sci Hortic 2004;101:333–342.
- David FD, Heidmann J, Heinen C, Lamp B, Martin J, Rost T, Silady R, Tong K, Tassel DV. Section of Plant Biology Division of Biological Sciences, University of California, DAVIS,1996.
- 45. Peet MM, Willits DH, Gardner RG. Responses of ovule development and post pollen production processes in male-sterile tomatoes to chronic, sub-acute high temperature stress. J Exp Bot 1997;48:101–111.
- 46. Sato S, Peet MM, Gardner RG. Formation of partenocarpic fruit, undeveloped flowers and aborted flowers in tomato under moderately elevated temperatures. Sci Horticult 2001;90:243–254.
- 47. Firon N, Shaked R, Peet MM, Phari DM, Zamski E, Rosenfeld K, Althanand L, Pressman NE. Pollen Grains of Heat Tolerant Tomato Cultivars Retain Higher Carbohydrate Concentration Under Heat Stress Conditions. Sci Horticult 2006;109:212–217.
- Foolad MR. "Breeding for abiotic stress tolerances in tomato," In: Abiotic Stresses: Plant Resistance Through Breeding and Molecular Approaches, Ashraf M, Harris PJC, (eds). The Haworth Press, New York, NY, USA, 2005;613-684.
- Abdelmageed AH, Gruda N, Geyer B. Effect of high temperature and heat shock on tomato (*Lycopersicon esculentum* M.) genotypes under controlled conditions. Conference on International Agricultural Research for Development. Deutscher Tropentag, Göttingen, October 8–10, 2003.
- 50. Pressman E, Peet MM, Phar DM. The Effect of heat stress on tomato pollen characteristics is associated with changes in carbohydrate concentration in developing anthers. Ann Bot 2002;90:631–636.
- Kinet JM, Peet MM. Tomato. In: Wien HC, (ed). The Physiology of Vegetable Crops. CAB International, Wallingford, UK, 1997;207–258.
- 52. Sato S, Kamiyama M, Iwata T, Makita N, Furukawa H, Ikeda H. Moderate increase of mean daily temperature adversely affects fruit set of *Lycopersicon esculentum* by disrupting specific physiological processes in male reproductive development. Ann Bot 2006;97:731–738.
- Comlekcioglu N, Soylu MK. Determination of high temperature tolerance via screening of flower and fruit formation in tomato. J Agric Sci 2010;20(2):123-130.
- Weaich K, Briston KL, Cass A. Modeling preemergent maize shoot growth. II. High temperature stress conditions. Agric J 1996;88:398–403.
- 55. Berry SZ, Rafique-Uddin M. Effect of high temperature on fruit set in tomato cultivars and selected germplasm. Hort Science 1988;23:606–608.
- Wollenweber B, Porter JR, Schellberg J. Lack of interaction between extreme high temperature events at vegetative and reproductive growth stages in wheat. J Agron Crop Sci 2003;189:142–150.
- Alsadon AA, Wahb-allah MA, Khalil SO. *In vitro* Evaluation of Heat Stress Tolerance in Some Tomato Cultivars. J King Saud Univ 2006;19(1):13-24.

- Saadella MM, Shanahan JF, Quick JS. "Heat Tolerance in Winter Wheat: 1. Hardining and Genetic Effects on Membrane Thermostability." Crop Sci 1990;30:1243-1247.
- 59. Kuo CG, Chen HM, Sun HC. "Membrane Thermostability and Heat Tolerance of Vegetable Leaves." In: Kuo CG, (ed). Adaptation of Food Crops to Temperature and Water Stress. Shanhua, Taiwan, Asian Veg Res Dev Center 1993;160-168.
- 60. Blum A. Plant Breeding for Stress Environments. CRC Press Inc., Boca Raton, Florida, 1988;223.
- 61. Mazorra LM, Nunez M, Echerarria E, Coll F, S'anchez-Blanco MJ. Influence of brassinosteriods and antioxidant enzymes activity in tomato under different temperatures. Plant Biol 2002;45:593–596.
- 62. Morales D, Rodr'ıguez P, Dell'amico J, Nicol'as E, Torrecillas A, S'anchez- Blanco MJ. High-temperature preconditioning and thermal shock imposition affects water relations, gas exchange and root hydraulic conductivity in tomato. Biol Plant 2003;47:203–208.
- 63. Tsukaguchi T, Kawamitsu Y, Takeda H, Suzuki K, Egawa Y. Water status of flower buds and leaves as affected by high temperature in heat tolerant and heat-sensitive cultivars of snap bean (*Phaseolus vulgaris* L.). Plant Prod Sci 2003;6:4–27.
- Hare PD, Cress WA, Staden JV. Dissecting the roles of osmolyte accumulation during stress. Plant Cell Environ 1998;21:535–553.
- 65. Sakamoto A, Murata N. The role of glycine betaine in the protection of plants from stress: clues from transgenic plants. Plant Cell Environ 2002;25:163–171.
- Sairam RK, Tyagi A. Physiology and molecular biology of salinity stress tolerance in plants. Curr Sci 2004;86:407–421.
- 67. Ashraf M, Foolad MR. Roles of glycine betaine and proline in improving plant abiotic stress resistance. Environ Exp Bot 2007;59:206–216.
- Wise RR, Olson AJ, Schrader SM, Sharkey TD. Electron transport is the functional limitation of photosynthesis in field-grown Pima cotton plants at high temperature. Plant Cell Environ 2004;27:717– 724.
- Salvucci ME, Crafts-Brandner SJ. Inhibition of photosynthesis by heat stress: the activation state of Rubisco as a limiting factor in photosynthesis. Physiol Plant 2004;120:179–186.
- Marchand FL, Mertens S, Kockelbergh F, Beyens L, Nijs I. Performance of high arctic tundra plants improved during but deteriorated after exposure to a simulated extreme temperature event. Global Change Biol 2005;11:2078–2089.
- Wahid A, Ghazanfar A. Possible involvement of some secondary metabolites in salt tolerance of sugarcane. J Plant Physiol 2006;163:723–730.
- Karim MA, Fracheboud Y, Stamp P. Heat tolerance of maize with reference of some physiological characteristics. Ann Bangladesh Agri 1997;7:27–33.
- 73. Karim MA, Fracheboud Y, Stamp P. Photosynthetic activity of developing leaves in maize is less affected by heat stress than that of developed leaves. Physiol Plant 1999;105:685–693.

- Camejo D, Jim'enez A, Alarc'on JJ, Torres W, G'omez JM, Sevilla F. Changes in photosynthetic parameters and antioxidant activities following heat-shock treatment in tomato plants. Funct Plant Biol 2006;33:177–187.
- 75. Guo YP, Zhou HF, Zhang LC. Photosynthetic characteristics and protective mechanisms against photo-oxidation during high temperature stress in two citrus species. Sci Hort 2006;108:260–267.
- Bukhov NG, Wiese C, Neimanis S, Heber U. Heat sensitivity of chloroplasts and leaves: leakage of protons from thylakoids and reversible activation of cyclic electron transport. Photosyn Res 1999;59:81–93.
- 77. Mcdonald GK, Paulsen GM. High temperature effects on photosynthesis and water relations of grain legumes. Plant Soil 1997;196:47–58.
- Todorov DT, Karanov EN, Smith AR, Hall MA. Chlorophyllase activity and chlorophyll content in wild type and *eti 5* mutant of *Arabidopsis thaliana* subjected to low and high temperatures. Biol Plant 2003;46:633– 636.
- 79. Kepova KD, Holzer R, Stoilova LS, Feller U. Heat stress effects on ribulose-1,5-bisphosphate carboxylase/oxygenase, Rubisco bindind protein and Rubisco activase in wheat leaves. Biol Plant 2005;49:521–525.
- Savchenko GE, Klyuchareva EA, Abrabchik LM, Serdyuchenko EV. Effect of periodic heat shock on the membrane system of etioplasts. Russ J Plant Physiol 2002;49:349–359.
- 81. Chen THH, Shen ZY, Lee PH. Adaptability of crop plants to high temperature stress. Crop Sci 1982;22:719–725.
- Martineau JR, Specht JE, Williams JH, Sullivan CY. Temperature tolerance in soybean. I. Evaluation of technique for assessing cellular membrane thermostability. Crop Sci 19679;19:75–78.
- Ashraf M, Saeed MM, Qureshi MJ. Tolerance to high temperature in cotton (*Gossypium hirsutum* L.) at initial growth stages. Environ Exp Bot 1994;34:275– 283.
- 84. Blum A, Klueva N, Nguyen HT. Wheat cellular thermo-tolerance is related to yield under heat stress. Euphytica 2001;117:117–123.
- 85. Marcum KB. Cell membrane thermo-stability and whole plant heat tolerance of Kentucky bluegrass. Crop Sci 1998;38:1214–1218.
- 86. Wahid A, Shabbir A. Induction of heat stress tolerance in barley seedlings by pre-sowing seed treatment with glycinebetaine. Plant Growth Reg 2005;46:133–141.
- Maestri E, Klueva N, Perrotta C, Gulli M, Nguyen HT, Marmiroli N. Molecular genetics of heat tolerance and heat shock proteins in cereals. Plant Mol Biol 2002;48:667–681.
- Larkindale J, Huang B. Effects of abscisic acid, salicylic acid, ethylene and hydrogen peroxide in thermotolerance and recovery for creeping bentgrass. Plant Growth Regul 2005;47:17–28.
- Pareek A, Singla SL, Grover A. Proteins alterations associated with salinity, desiccation, high and low temperature stresses and abscisic acid application in

seedlings of Pusa 169, a high-yielding rice (*Oryza sativa* L.) cultivar. Curr Sci 1998;75:1023–1035.

- Dhaubhadel S, Chaudhary S, Dobinson KF, Krishna P. Treatment with 24-epibrassinolide, a brassinosteroid, increases the basic thermo-tolerance of *Brassica napus* and tomato seedlings. Plant Mol Biol 1999;40:333– 342.
- Nakamoto H, Hiyama T. Heat-shock proteins and temperature stress. In: Pessarakli M, (ed). Handbook of Plant and Crop Stress. Marcel Dekker, New York, 1999;399–416.
- Vierling E. The role of heat shock proteins in plants. In: Annu Rev Plant Physiol Plant Mol Biol 1991;42:579–620.
- 93. Hopf N, Plesofskv-Vig N, Brambl R. The heat response of pollen and other tissues of maize. Plant Mol Biol 1992;19:623–630.
- Prasinos C, Krampis K, Samakovli D, Hatzopoulos P. Tight regulation of expression of two Arabidopsis cytosolic *Hsp90* genes during embryo development. J Exp Bot 2005;56:633–644.
- Feussner K, Feussner I, Leopold I, Wasternack C. Isolation of a cDNA coding for an ubiquitinconjugating enzyme UBC1 of tomato-the first stressinduced UBC of higher plants. FEBS Lett 1997;409:211–215.
- Neumann DM, Emmermann M, Thierfelder JM, Zur NU, Clericus M, Braun HP, Nover L, Schmitz UK. HSP68-a DNA K-like heat-stress protein of plant mitochondria. Planta 1993;190:32–43.
- 97. Liu N, Ko S, Yeh KC, Charng Y. Isolation and characterization of tomato Hsa32 encoding a novel heat-shock protein. Plant Sci 2006;170:976–985.
- Nieto-Sotelo J, Mart'inez LM, Ponce G, Cassab GI, Alag'on A, Meeley RB, Ribaut JM, Yang R. Maize HSP101 plays important roles in both induced and basal thermo-tolerance and primary root growth. Plant Cell 2002;14:1621–1633.
- 99. Yang KA, Lim CJ, Hong JK, Park CY, Cheong YH, Chung WS, Lee KO, Lee SY, Cho MJ, Lim CO. Identification of cell wall genes modified by a permissive high temperature in Chinese cabbage. Plant Sci 2006;171:175–182.
- Miroshnichenko S, Tripp J, Nieden U, Neumann D, Conrad U, Manteuffel R. Immuno-modulation of function of small heat shock proteins prevents their assembly into heat stress granules and results in cell death at sublethal temperatures. Plant J 2005;41:269– 281.
- Iba K. Acclimative response to temperature stress in higher plants: approaches of gene engineering for temperature tolerance. Annu Rev Plant Biol 2002;53:225–245.
- 102. Barua D, Downs CA, Heckathorn SA. Variation in chloroplast small heat-shock protein function is a major determinant of variation in thermo-tolerance of photosynthetic electron transport among ecotypes of *Chenopodium album*. Funct Plant Biol 2003;30:1071–1079.
- 103. Neta-Sharir I, Isaacson T, Lurie S, Weiss D. Dual role for tomato heat shock protein 21: protecting photosystem ii from oxidative stress and promoting

color changes during fruit maturation. Plant Cell 2005;17:1829-1838.

- 104. Kadirvel P. Molecular breeding in vegetable crops: Opportunities and challenges presented In: 2<sup>nd</sup> National Workshop on Marker-Assisted Selection for Crop Improvement. ICRISAT, Patancheru, A.P. India, 2010.
- 105. Kalloo G. "Breeding for environmental resistance in tomato," In: Genetic Improvement of Tomato, Kalloo G, (ed). Of Monographs on Theoretical and Applied Genetics, Springer, Berlin, Germany, 1991;14:153–165.
- 106. Mutschler MA, Tanksley SD, Rick CM. "Linkage maps of the tomato (*Lycopersicon esculentum*)." Report of the Tomato Genetics Cooperative, 1987;37:5–34.
- 107. Tanksley SD. "Linkage map of the tomato (*Lycopersicon esculentum*) (2N =24)," In: Genetic Maps: Locus Maps of Complex Genomes, O'Brian SJ, (ed). Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, USA, 1993;6.3–6.15.
- 108. Chetelat RT. "Revised list of monogenic stocks," Report of the Tomato Genetics Cooperative, 2002;52:41-62.
- 109. Tanksley SD, Bernatzky R. "Molecular markers for the nuclear genome of tomato," In: Plant Biology, Vol. 4, Tomato Biotechnology, Nevins DJ, Jones RA, (eds). Alan R. Liss, New York, NY, USA, 1987;37–44.
- Foolad MR, Jones RA, Rodriguez RL. "RAPD markers for constructing intraspecific tomato genetic maps." Plant Cell Reports 1993;12(5):293–297.
- 111. Tanksley SD, Orton TJ. Isozymes in Plant Genetics and Breeding, Elsevier, Amsterdam, The Netherlands, 1983.
- 112. Botstein D, White RL, Skolnick M, Davis RW. "Construction of a genetic linkage map in man using restriction fragment length polymorphisms." Am J Hum Genet 1980;32(3):314–331.
- 113. Williams JGK, Kubelik AR, Livak KJ, Rafalski JA, Tingey SV. "DNA polymorphisms amplified by arbitrary primers are useful as genetic markers," Nucleic Acids Res 1990;18(22):6531–6535.
- 114. He C, Poysa V, Yu K. "Development and characterization of simple sequence repeat (SSR) markers and their use in determining relationships among *Lycopersicon esculentum* cultivars." Theor Appl Genet 2003;106(2):363–373.
- 115. Tautz D. "Hypervariability of simple sequences as a general source for polymorphic DNA markers." Nucleic Acids Res 1989;17(16):6463–6471.
- 116. Vos P, Hogers R, Bleeker M. "AFLP: a new technique for DNA fingerprinting," Nucleic Acids Res 1995;23(21):4407–4414.
- 117. Konieczny A, Ausubel FM. "A procedure for mapping *Arabidopsis* mutations using co-dominant ecotypespecific PCR-basedmarkers." Plant Journal 1993;4(2):403–410.
- Jeffreys AJ, Wilson V. Thein SL. "Hypervariable 'minisatellite' regions in human DNA." Nature 1985;314(6006):67–73.
- Paran I, Michelmore RW. "Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce," Theor Appl Genet 1993;85(8):985– 993.

- Adams MD, Kelley JM, Gocayne JD. "Complementary DNA sequencing: expressed sequence tags and human genome project." Science 1991;252(5013):1651–1656.
- 121. Fulton TM, Van der Hoeven R, Eannetta NT, Tanksley SD. "Identification, analysis and utilization of conserved ortholog set markers for comparative genomics in higher plants." The Plant Cell 2002;14(7):1457–1467.
- 122. Orita M, Iwahana H, Kanazawa H, Hayashi K, Sekiya T. "Detection of polymorphisms of human DNA by gel electrophoresis as single-strand conformation polymorphisms." Proc Natl Acad Sci 1989;86(8):2766–2770.
- Landegren U, Nilsson M, Kwok PY. "Reading bits of genetic information: methods for single-nucleotide polymorphism analysis." Genome Research 1998;8(8):769–776.
- 124. Zhang Q, Shen BZ, Dai BZ. Using bulked extremes and recessive class to map genes for photoperiodsensitive genetic make sterility in rice. Proc Natl Acad Sci 1994;91:8675-8679.
- 125. Mackay IJ, Caligari PDS. Efficiencies of F<sub>2</sub> and backcross generations for bulked segregant analysis using dominant markers. Crop Sci 2000;40:626–630.
- 126. Lin KH, Lo HF, Leo SP, Kuo CG, Chen JT, Yeh WL. RAPD markers for the identification of yield traits in tomatoes under heat stress via bulked segregant analysis. Hereditas 2006;143:142–154.
- 127. Lin KH, Yeh WL, Chen HM, Lo HF. Quantitative trait loci influencing fruit-related characteristics of tomato grown in hightemperature conditions. Euphytica 2010;13 Feb. (online)
- 128. Mansour A, Ismail HM, Ramadan MF, Gyulai G. Variations in tomato (*Lycopersicon esculentum*) cultivars grown under heat stress. J Verbr Lebensm 2009;4:118–127.
- 129. Tanksley SD, Young ND, Paterson AH, Bonierbale MW. "RFLP mapping in plant breeding: new tools for an old science." Bio/Technology 1989;7:257–264.
- Ribaut JM, Jiang C, Hoisington D. "Simulation experiments on efficiencies of gene introgression by backcrossing." Crop Sci 2002;42(2):557–565.
- 131. Servin B, Martin OC, M'ezard M, Hospital F. "Toward a theory of marker-assisted gene pyramiding." Genetics 2004;168(1):513–523.
- 132. Stuber CW, Edward MD. "Genotypic selection for improvement of quantitative traits in corn using molecular marker loci," In: Proceedings of the 41st Annual Corn and Sorghum Sorghum Industry Research Conference, American Seed Trade Association, Chicago, Ill, USA, 1986;70–83.
- 133. Edwards M, Johnson L. "RFLPs for rapid recurrent selection," In: Proceedings of Symposium on Analysis of Molecular Marker Data, Am Soc Hort Sci, CSSA, Corvallis, Ore, USA, 1994;33–40.
- Eathington SR, Dudley JW, Rufener II GK. "Usefulness of marker-QTL associations in early generation selection." Crop Science 1997;37(6):1686– 1693.
- 135. Schneider KA, Brothers ME, Kelly JD. "Markerassisted selection to improve drought resistance in common bean." Crop Sci 1997;37(1):51–60.

- 136. Knapp SJ. "Marker-assisted selection as a strategy for increasing the probability of selecting superior genotypes." Crop Science 1998;38(5):1164–1174.
- 137. Toojinda T, Baird E, Booth A. "Introgression of quantitative trait loci (QTLs) determining stripe rust resistance in barley: an example of marker-assisted line development." Theor Appl Genet 1998;96(1):123–131.
- 138. Stuber CW, Polacco M, Senior ML. "Synergy of empirical breeding, marker-assisted selection and genomics to increase crop yield potential." Crop Sci 1999;39(6):1571–1583.
- 139. Zhu H, Brice no G, Dovel R. "Molecular breeding for grain yield in barley: an evaluation of QTL effects in a spring barley cross." Theor Appl Genet 1999;98(5):772–779.
- 140. Hospital F, Goldringer I, Openshaw S. "Efficient marker-based recurrent selection for multiple quantitative trait loci." Genetical Research 2000;75(3):357–368.
- 141. Bouchez A, Hospital F, Causse M, Gallais A, Charcosset A. "Marker-assisted introgression of favorable alleles at quantitative trait loci between maize elite lines,"Genetics 2002;162(4):1945–1959.
- 142. Taran B, Michaels TE, Pauls KP. "Marker-assisted selection for complex trait in common bean (*Phaseolus vulgaris* L.) using QTL-based index." Euphytica 2003;130(3):423–432.
- 143. Zhou PH, Tan YF, He YQ, Xu CG, Zhang Q. "Simultaneous improvement for four quality traits of Zhenshan 97, an elite parent of hybrid rice, by molecular marker assisted selection." Theor Appl Genet 2003;106(2):326–331.
- 144. Jiang GH, Xu CG, Tu JM, Li XH, He YQ, Zhang QF. "Pyramiding of insect- and disease-resistance genes into an elite *indica*, cytoplasm male sterile restorer line of rice, 'Minghui 63'," Plant Breeding 2004;123(2):112–116.
- 145. Collard BCY, Jahufer MZZ, Brouwer JB, Pang ECK. An introduction to markers, quantitative trait loci (QTL) mapping and marker-assisted selection for crop improvement: The basic concepts. Euphytica 2005;142:169–196.
- 146. Rick CM. Germplasm resources in the wild tomato species. Acta Hort 1986;190:39-47.
- 147. Rick CM. Hybridization between Lycopersicon esculentum and Solanum pennellii, Phylogenetic and cytogenetic significance. Proc Natl Acad Sci 1960;46:78-82.
- 148. Stoeva PK, Zagorska N. Hybrids between *Solanum lycopersicoides* and self compatible species of *Lycopersicon*. Rept Tomato Genet Coop 1987;37:73-74.
- Wann EV, Jhonson KW. Intergeneric hybridization in involving species of *Solanum* and *Lycopersicon*. Bot Gaz (Chicago) 1963;124:451-455.
- 10/15/2012

- 150. Yordanov M. Heterosis in tomato. Monogr. Theor Appl Genet 1983;6:189-219.
- 151. AVDRC. Progress report 1986. Asian Vegetable Research and Development Center. Shanhua, Taiwan, 1988;541.
- 152. Opena RT, Kuo GC, Yoon JY. Breeding for stress tolerance under tropical conditions in tomato and heading Chinese cabbage. ASPAC-FFTC. Taipei, Taiwan, 1987;88-109.
- 153. Hanna HY, Millhollon EP, Herrick JK, Fletcher CL. Increased yield of heat tolerant Tomatoes with Deep Transplanting, Morning Irrigation and White Mulch. Hort Science 1997;32(2):224-226.
- 154. Scott JW, Bryan HH, Ramos LJ. High temperature fruit setting ability of large-fruited, jointless pedicel tomato hybrids with various combinations of heat-tolerance. Proc Fla State Hortic Soc 1997;110:281–284.
- 155. Scott JW, Volin RB, Bryan HH, Olson SM. "Use of hybrids to develop heat tolerant tomato cultivars." In: Proceedings of the Florida State Horticultural Society, 1986;99:311–315.
- 156. Scott JW, Olson SM, Howe TK, Stoffella PJ, Bartz JA, Bryan HH. "Equinox' heat-telerant hybrid tomato." Hort Science 1995;30:647–648.
- 157. Bohnert HJ, Gong Q, Li P, Ma S. Unraveling abiotic stress tolerance mechanisms-getting genomics going. Curr Opin Plant Biol 2006;9:180–188.
- 158. Sun A, Yi S, Yang J, Zhao C, Liu J. Identification and characterization of a heat-inducible ftsH gene from tomato (*Lycopersicon esculentum* Mill.). Plant Sci 2006;170:551–562.
- Liu J, Shono M. Characterization of mitochondrialocated small heat shock protein from tomato (*Lycopersicon esculentum*). Plant Cell Physiol 1999;40:1297–1304.
- Sanmiya K, Suzuki K, Egawa Y, Shono M. Mitochondrial small heat shock protein enhances thermo-tolerance in tobacco plants. FEBS Lett 2004;557:265–268.
- 161. Queitsch C, Hong SW, Vierling E, Lindquest S. Heat shock protein 101 plays a crucial role in thermo-tolerance in *Arabidopsis*. Plant Cell 2000;12:479–492.
- 162. Xu S, Li J, Zhang X, Wei H, Cui L. Effects of heat acclimation pretreatment on changes of membrane lipid peroxidation, antioxidant metabolites and ultrastructure of chloroplasts in two cool-season turfgrass species under heat stress. Environ Exp Bot 2006;56:274–285.
- 163. Whitaker BD. A reassessment of heat treatment as a means of reducing chilling injury in tomato fruit. Postharvest Biol Technol 1994;4:75–83.