

Bioavailability of cadmium and zinc following salinization of a degraded Isohyperthermic Arenic Kandiudult

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Abstract

This study was conducted in 2006 to investigate bioavailability of cadmium (Cd) and zinc (Zn) using 5 levels of salt treatments. It was a greenhouse study using maize (*Zea mays* L.) as indicator crop in a Completely Randomized Design (CRD). Soil data were analyzed using analysis of variance (ANOVA) in a SAS Computer package. Results showed that high rates of salinity decreased total Zn (Zn_T) and decreased Zn content of maize shoots. Application of $ZnSO_4$ decreased Cd in uptake shoots of maize (*Zea mays* L.) and increased shoot dry matter weight and yield. [Life Science Journal. 2007; 4(2): 64 – 68] (ISSN: 1097 – 8135).

Keywords: bioaccessibility; degradation; heavy metals; Kandiudult; remediation

1 Introduction

Zinc (Zn) is a micronutrient required for the maintenance of the integrity of biomembranes (Marschner, 1995). Obasi (2003) reported that application of Zn alone or in combination with copper (Cu) and iron (Fe) significantly reduced harvest drop and increased fruit weight, length, diameter and total yield of orange tree (*Citrus sinensis*). However, Zn complexes are formed under acidic pH range (Isirimah *et al*, 2003) and the element sorbs, thereby becoming unavailable when soils become alkaline (Tinker and Lauchi, 1984).

Cadmium (Cd) is a biotoxic heavy metal regarded as an important environmental pollutant in agricultural soils. Because of the potential adverse effects, it may pose to food quality, soil health and the environment (Gray *et al*, 2004). But it is the labile fraction rather than the total Cd content that is critical when assessing Cd availability in soils, and consequent bioaccessibility in other forms of life.

Zinc and Cd are chemically similar as shown in chemical and physiological processes they undergo in soils and plants (Jalil *et al*, 1994). The bioavailability of cadmium

element depends on the concentration form of the metal, concentration of Zn, soil pH, organic matter content, clay content, presence of other cations, complexing ligands and fertilization practices (Norvel *et al*, 2000).

In the humid tropics and indeed developing countries, scanty studies have been conducted on the above enumerated factors influencing biotoxic Cd. Onweremadu *et al* (2007) investigated the temporal variability of selected heavy metals in automobile soils and found that Cd concentrations in these soils are above critical limit. This study did not evaluate effects of these heavy metals on nutrient uptake.

Amidst limited attention given to Zn-fertilization (Takar and Walker, 1993), it becomes necessary to investigate the effects of sanization in the uptake of Zn in cadmium polluted arable soils. The aim of this study was to assess the phytoavailability of cadmium and zinc when varying rates of sodium chloride (NaCl) were added in soils using maize (*Zea mays* L.) as an indicator crop.

2 Materials and Methods

2.1 Study area

The study was conducted at the University Teaching and Research Farm of Federal University of Technology, Owerri, Nigeria. The site is located on latitude 5°

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27° 50' .230 N and longitude 7° 2' 49' .330 E, with an elevation of 55 metres above mean sea level (Handheld Global Positioning System-GPS) receiver (Garmin Ltd, Kansas, United States of America). Soils are derived from Coastal Plain Sands (Benin formation). In an earlier study (Onweremadu *et al*, 2006) soils were classified as Isohyperthermic Arenic Kandiudult (Soil Survey Staff, 2003). The study area has over 2,500 mm of total annual rainfall. Temperatures are high and change only slightly during the year (mean daily temperature is about 27°C). Rainforest vegetation predominates, although anthropogenic activities, especially urbanization, deforestation and farming have altered the original biodiversity.

2.2 Soil sampling

Surface soil samples were collected from cadmium-contaminated arable soil proximal to an automobile service station in 2006. Soils were classified as Isohyperthermic Arenic Kandiudult. Cadmium accumulation in the soils of the area is attributed to automobile wastes and inorganic fertilizer applications.

2.3 Greenhouse experiment

Bulk soil sample weighing about 300 kg was air-dried, crushed, thoroughly mixed and sieved to remove particle sizes greater than 2 mm. Sieved sample were brought to the greenhouse. Uniform soil weighing 3.0 kg was put into polyethylene pots (28 cm height, 19 cm diameter). 5-cm layer of well-washed sand was first filled in the pots to enhance drainage. Two levels of Zn (0 and 15 mg Zn/Kg dry soil, in the form of zinc sulphate) and 5 salinity levels were achieved using irrigation water with salinities of 0, 50, 100 and 150 mM NaCl, and 100 mM NaNO₃, respectively. An inclusion of NaNO₃ (sodium nitrate) helped to distinguish between effects of osmotic stress and chloro-cadmium complexation in solution on plant uptake. At planting, uniform rates of nitrogen and potassium fertilizers, 100 mg/kg N and K each ammonium sulphate and potassium sulphate respectively were applied to each pot. The fertilizer application was as recommended by The Fertilizer Procurement and Distribution Division (Federal Ministry of Agriculture and Natural Resources, 1990). These fertilizers were mixed thoroughly with soil before planting. Soil moisture content was maintained at field capacity (20%) gravimetric water using deionized water.

Maize seeds were sown and thinned to 2 plants per pot after 7 days of planting. Planting of maize was done in a Completely Randomized Design (CRD) with factorially arranged treatments and three replications. At harvest, shoots were cut at the soil surface, washed and dried at 70°C.

2.4 Laboratory analysis

2.4.1 Soil samples: Particle size distribution was determined by hydrometer method (Gee and Or, 2002). Soil pH was estimated using a pH meter (Model 691 Metrohm AG Herisau, Switzerland) in a 1.2 soil/water ratio, while electrical conductivity in soil saturation extracts was determined with an FC-meter (Model Ohm-644 Metrohm AG, Herisau, Switzerland). Available phosphorus was extracted from the soil with 0.5 M NaHCO₃ and determined according to the procedure of Olsen and Sommers (1990). Available potassium was extracted with ammonium acetate and estimated using flame-photometer (Chapman and Pratt, 1961).

Total Cd and Zn contents were measured by microwave digestion using method 3051 (USEPA, 1995). In this method 0.5 g of air-dried subsample (less than 150 µm) was digested in XP1500 plus Teflon-PFA microwave vessel (CEM Corp, Matthews NC) using 9 ml of nitric acid and 3 ml of hydrochloric acid then filtered through Whatman No. 42 filters. This was transferred to 50 ml volumetric flasks and diluted with deionized distilled water (DDW). Chelate-extractable Cd and Zn were extracted using 0.005 M DTPA-TEA (Lindsay and Norvele, 1978) and then measured on a graphite furnace atomic absorption spectrophotometer (GFAAS) (Perkin-Elmer 3400, Perkin Elmer Willsely, MA).

2.4.2 Pant samples: The dried shoot materials were ground and ashed at a temperature of 550°C for 7 hours and the ash was dissolved in hydrochloric acid. The digest solutions were used in the determination of Cd and Zn by GFAAS.

2.4.3 Post-harvest soil tests: After harvesting the maize, a sample of 800 g soil was collected from the soil surface of each pot, air-dried, crushed and sieved using a 2-mm sieve. Each soil sample was saturated with deionized water, mixed to a paste of uniform consistency and kept for 24 hours. After this, it was transferred to the suction flask for extraction of the soil solution. Soil pH and EC of the saturated extract, concentrations of Ca, Mg, Na, K, SO₄²⁻, HCO₃⁻, P, NO₃⁻, Cd, Zn and Fe were determined using the methods earlier described.

2.4.4 Data analysis: Soil and plant data were analyzed by analysis of variance using the SAS Computer programme (Little *et al*, 1996).

3 Results

3.1 Soil properties

Results of soil properties are shown in Table 1, indicating a sandy loam texture. Soil pH was extremely low.

Salinity level did not significantly ($P < 0.05$) affect soil pH (Table 2). Sodium cation concentration increased almost proportionally to the applied sodium chloride and sodium nitrate (Table 2). This trend was followed by chloride anions. Results also showed that Cd concentrations in soil solutions increase significantly ($P < 0.05$) as sodium chloride rate increased (Table 2), and this statement is true for both experiments (that is, without Zn and with Zn treatment). Sodium nitrate application resulted to little rise in Cd concentration of saturated extract. Treatment with zinc sulphate ($ZnSO_4$) had little or no effect on dissolved Zn, whereas there was outstanding increase in sulphate anion concentration, resulting from the application of Zn to the soil system (Table 2).

3.2 Effects of salinization and zinc application on metallic concentrations in the maize shoot

Concentrations of Cd and Zn in maize shoots resulting from the treatments are presented on Table 3. Zinc treatment lowered Cd concentration in the shoot while it did not in the without Zn treatment. Again, increasing NaCl concentration significantly ($P < 0.05$) led to a rise in Cd concentrations in shoots in both zinc treatments (that is, without Zn and with Zn treatments). On the other hand, application of sodium nitrate had no significant effect on Cd concentrations in shoots of maize.

Table 1. Selected soil properties of the surface soil sample (0 – 30 cm) used in the greenhouse study

| Soil property | Value |
|-------------------------|--------------|
| Total sand | 800.0 g/kg |
| Silt | 40.0 g/kg |
| Clay | 160.0 g/kg |
| pH _{water} | 4.0 |
| EC | 2.1 mmhos/cm |
| Available P | 15.4 g/kg |
| Available K | 160.0 g/kg |
| DTPA-TEA extractable Zn | 0.9 g/kg |
| DTPA-TEA extractable Cd | 1.3 g/kg |
| Total Zn | 26.4 g/kg |
| Total Cd | 14.9 g/kg |

Salinization decreased Zn content in shoots while $NaNO_3$ treatment had little or no effect on zinc concentration in shoots (Table 3). However, Zn content increased in maize plants receiving $ZnSO_4$ when compared with those that did not.

Maize dry matter weight decreased significantly ($P < 0.05$) with increasing rate of NaCl application. Zinc application increased dry matter weight when compared without Zn treatment irrespective of salinity level (Table 4).

Table 2. Values of total ions in the saturated paste soil solution

| Salinity level | Concentration | | | | | | | | | |
|--------------------------------|--------------------------|--------------------------|------------------------|-------------------------|---------------------------------------|-------------------------|---------------------------------------|---------------------------|---------------------------|---------------|
| | Ca ²⁺ (mM) | Mg ²⁺ (mM) | K ⁺ (mM) | Na ⁺ (mM) | HCO ₃ ⁻ (mM) | Cl ⁻ (mM) | SO ₄ ²⁻ (mM) | Cd _T (mg/L) | Zn _T (mg/L) | pH (water) |
| Without Zn treatment | | | | | | | | | | |
| 0 NaCl | 23 | 3.8 | 0.4 | 8 | 1.3 | 12 | 12 | 0.02 | 0.14 | 4.0 |
| 50 NaCl | 24 | 4.3 | 0.6 | 18 | 1.3 | 27 | 13 | 0.02 | 0.11 | 4.1 |
| 100 NaCl | 25 | 4.8 | 0.9 | 30 | 1.8 | 42 | 14 | 0.11 | 0.14 | 4.0 |
| 150 NaCl | 25 | 4.8 | 1.3 | 46 | 2.3 | 58 | 16 | 0.41 | 0.12 | 4.0 |
| 100 NaNO ₃ | 25 | 4.8 | 0.9 | 33 | 1.9 | 13 | 22 | 0.03 | 0.13 | 4.2 |
| With Zn treatment | | | | | | | | | | |
| 0 NaCl | 22 | 4.9 | 1.3 | 11 | 1.1 | 17 | 12 | 0.01 | 0.15 | 4.0 |
| 50 NaCl | 23 | 4.9 | 0.8 | 20 | 1.2 | 29 | 17 | 0.04 | 0.13 | 4.0 |
| 100 NaCl | 24 | 4.9 | 0.8 | 28 | 1.8 | 32 | 22 | 0.12 | 0.16 | 4.1 |
| 150 NaCl | 24 | 5.0 | 1.8 | 33 | 1.9 | 56 | 21 | 0.41 | 0.16 | 4.0 |
| 100 NaNO ₃ | 23 | 5.6 | 0.8 | 28 | 1.8 | 22 | 27 | 0.04 | 0.16 | 4.0 |
| LSD _{salinity (0.05)} | 2.7 | 0.3 | 0.2 | 1.8 | 0.1 | 2.2 | 3.1 | 0.20 | 0.03 | NS |
| LSD _{Zn (0.05)} | 1.6 | 0.2 | 0.1 | 1.1 | 0.1 | 1.3 | 2.0 | 0.01 | 0.03 | NS |

NS = Not significant, Cd_T = total Cd in the paste, Zn_T = total Zn in the paste

Table 3. Concentrations of Cd and Zn in maize shoots at different salt rates

| Treatment | Salinity level (mM) | Cd (mg/kg) | Zn (mg/kg) |
|----------------------|-----------------------|------------|------------|
| Without Zn treatment | 0 NaCl | 24.6 | 1.0 |
| | 50 NaCl | 22.6 | 1.1 |
| | 100 NaCl | 21.5 | 1.3 |
| | 150 NaCl | 16.3 | 2.9 |
| | 100 NaNO ₃ | 24.8 | 1.2 |
| With Zn treatment | 0 NaCl | 56.5 | 0.1 |
| | 50 NaCl | 48.0 | 1.0 |
| | 100 NaCl | 36.9 | 1.0 |
| | 150 NaCl | 39.0 | 2.2 |
| | 100 NaNO ₃ | 49.8 | 0.2 |

LSD_{salinity (0.05)}: Cd, 0.3, Zn, 0.1; LSD_{Zn (0.05)}: Cd, 0.2, Zn, 0.1.

Table 4. Shoot dry weight of maize in relation of salinity level

| Treatment | Salinity level (mM) | Shoot weight (g/plant) |
|----------------------|-----------------------|------------------------|
| Without Zn treatment | 0 NaCl | 14.8 ^a |
| | 50 NaCl | 14.3 ^{ab} |
| | 100 NaCl | 11.4 ^{bc} |
| | 150 NaCl | 7.8 ^c |
| | 100 NaNO ₃ | 12.6 ^d |
| With Zn treatment | 0 NaCl | 22.1 ^a |
| | 50 NaCl | 19.6 ^{ab} |
| | 100 NaCl | 16.2 ^{bc} |
| | 150 NaCl | 12.3 ^c |
| | 100 NaNO ₃ | 13.5 ^c |

Means in the same column followed by the same letter(s) are not significantly different for each treatment.

4 Discussion

The total Cd content (Table 1) was very much above critical limits of 10 mg/kg (EPA/ROC, 1989), and 0.2 mg/kg (Roberts *et al*, 1994). The high sand content did not promote poor retentivity or there was very severe pollution of soils of the site by Cd, thus high values of Cd in the sites. Low pH values suggest complexation of Zn and its unavailability and this is in line with the findings of Foth (1984) that most essential nutrients become avail-

able at slightly acidic and neutral pH ranges (pH = 6.0 – 6.2) since these are of low base status soils. Because, the studied soils are not saline, high uptake of Cd is not suspected if crop are grown in the field. Yet treatment of soils at 5 salinity levels did not significantly ($P < 0.05$) alter soil pH.

Application of sodium chloride on maize through irrigation water added sodium cations which competed with other cations, such as hydrogen and aluminium cations for sorption in the exchange sites. Also addition of chloride anions may have attracted positively charged Cd ions to form Cd-complexes. These complexes may reduce Cd in soil solution available for maize uptake. Whereas NaNO₃ treatment did not increase Cd concentration in soil solution, higher salinity levels did.

Without zinc treatment, salinity influenced K ions in the saturated paste soil solutions (Table 2), indicating that cation exchange effect resulted from salinizing the irrigation water used in the experiment. The implication is that salinity levels altered not only K but other cations although in a negligible manner.

Salinized irrigation water significantly ($P < 0.05$) increased Cd content in the shoots of maize (*Zea mays* L.) as indicated in Table 3. It can be argued that this increase in shoot content of Cd is attributable to osmosis, but 120 mM sodium nitrate treatment had no significant effect on Cd plant concentrations in an experiment conducted by Smolders *et al* (1998).

There was a proportionate decline in maize dry matter weight and yield resulting from salt applications (Table 4). This condition requires an application of zinc fertilizer to augment for deficiencies created by salinization. Such Zn fertilizers reduce plant uptake of boron, sodium and chlorine, and the associated low yield of maize.

The level of Cd in soil and maize shoots suggested chemical immobilization methods which rapidly increase exchange sites for Cd-adsorption (Chen, 1997). Applications of zeolite reduces the concentration of Cd that dissolves in polluted soils (Mench *et al*, 1994) while heavy applications of phosphates to concentrated soils can reduce Zn dissolved in soil solution by precipitation (Saeed and Fox 1979).

5 Conclusions

Salinization enhanced the mobilization of Cd in soils and this promoted bioavailability. As a consequence, there was increased Cd in maize shoots. The study also revealed that fertilization of soils with zinc decreased Cd concentrations in shoots of maize.

References

- Chapman HD, Pratt PF. Methods of analysis for soils, plants and waters. University of California. Division of Agricultural Science, Riverside. 1961.
- Chen ZS. Assessment and experience on the soil remediation techniques used in the soils contaminated with cadmium and lead in Taiwan. In: Z.S. Chen Proceedings of the 5th Workshop on soil pollution and Prevention: Symposium on Soil Remediation Technologies on Contaminated Soils. 1997; 75 – 88.
- EPA/ROC. Final reports of heavy metals contents in Taiwan Agricultural Soils. 4 Volumes. Tapei, Taiwan ROC. 1989.
- Federal Ministry of Agriculture and Natural Resources Literature Review on Soil Fertility Investigations in Nigeria. Five volumes, FMANR, Lagos, Nigeria. 1990.
- Foth HD. Fundamentals of soil science. 7th edition John Wiley and Sons, New York. 1984.
- Gee GW, Or D. Particle size analysis. In: Dane JH and Topp GC (eds.). Methods of soil analysis, part 4. physical methods. Soil Sci. Soc. Am. Book Series No. 5 ASA and SSSA Madison, WI, 2002; 255 – 93.
- Gray CW, McLaren RG, Gunther D, Sinaj S. An assessment of cadmium availability in cadmium-contaminated soils using Isotope Exchange Kinetics. Soil Sci Soc Am J 2004; 68: 1210 – 7.
- Isirimah NO, Dickson AA, Igwe CA. Introductory Soil Chemistry and Biology for Agriculture and Biotechnology OSIA Int’L. Publishers Ltd, Port Harcourt, Nigeria 2003.
- Jalil AF, Siles, Clark JM. Effect of Cd on growth and uptake of Cd and other elements by durum wheat. J plant Nutr 1994; 17: 1839 – 58.
- Lindsay WI, Norvell WA. Development of a DTPA soil test for zinc, iron, manganese and copper. Soil Sci Soc Am J 1978; 42: 421 – 8.
- Little RC, Milliken GA, Stroup WW, Wolfinger RC. SAS system for mixed models. Statistical System Inc., Cary North Carolina, USA, 1996; 633.
- Marschner H. Mineral Nutrition of Higher Plants. 2nd edition Academic Press, San Diego, CA. 1995.
- Mench M, Didier VL, Loffler M, Gomez A, Masson P.A mimicked in-situ remediation study of metal-contaminated soils with emphasis on cadmium and lead. J Environ Qual 1994; 23: 58 – 63.
- Norvell WA, Wu J, Hopkins DG, Welch RM. Association of cadmium in durum in wheat grain with soil chloride and chelate-extractable soil cadmium. Soil Sci Soc Am J 2000; 64: 2162 – 8.
- Obasi MO. Influence of foliar applied area and micronutrients on leaf parameters, fruit yield and characteristics of sweet orange. Proceedings of the 28th Annual Conference of the Soil Science Society of Nigeria, National Root Crops Research Institute, Umudike Nigeria 4 – 7 November 2003; 51 – 6.
- Olsen SR, Sommers LE. Phosphorus. In: A. L. Page (ed.). Methods of soil analysis, Part 2 ed. Agron. Monogr. No. 9 ASA Madison, WI 1990; 403 – 31.
- Onweremadu EU, Opara CC, Nkwopara U, Duruigbo CI, Ibeawuchi II. Yield response of a cowpea variety on ground seashells on Isohyperthermic Arenic Kandiodult of Owerri, Southeastern Nigeria. Int J Soil Sci 2006; 1 (3): 251 – 7.
- Onweremadu EU, Eshett ET, Osuji GE. Temporal variability of selected heavy metals in automobile soils. Int J Environ Sci Tech 2007; 4(1): 35 – 41.
- Roberts AHC, Longhurst RD, Brown MW. Cadmium status of soils, plants and grazing animals on New Zealand. NZJ Agric Res 1994; 37:119 – 29.
- Saeed M, Fox RL. Influence of phosphate fertilization on zinc adsorption by tropical soils. Soil Sci Soc Am J 1979; 43: 683 – 6.
- Smolders E, Lambergts RM, McLaughlin MJ, Tiller KG. Effect of soil solution chloride on cadmium availability to Swiss chard. J Environ Qual 1998; 27: 426 – 31.
- Soil Survey Staff. Keys to Soil Taxonomy. 9th edition. United States Department of Agriculture. 2003; 332.
- Takkar PN, Walker CD. The Distribution and Correction of Zinc Deficiency. In: A. D. Robson (ed.). Zinc in Soils and Plants. Kluwer Academic Publishers, Dordrecht, The Netherlands. 1993;151 – 65.
- Tinkler PB, Lauchli A. Advances in Plant Nutrition. Academic Publishers, San Diego, CA 1984.
- USEPA (United States Environmental Protection Agency). Method 3051. Microwave assisted acid digestion of sediments, sludges, soils and oils. Available online at <http://www.epa.gov/sw.84/pdfs/3051pdf> (Verified 30th August 2005), USEPA Washington, DC. 1995.